

1 **No sex differences in oxygen uptake or extraction kinetics in the moderate or heavy**
2 **exercise intensity domains**

3

4

5 Maria Solleiro Pons¹, Lina Bernert^{1,2}, Emily Hume¹, Luke Hughes¹, Zander Williams³, Mark
6 Burnley⁴, Paul Ansdell¹

7

8

9 ¹ Department of Sport, Exercise and Rehabilitation, Faculty of Health and Life Sciences,
10 Northumbria University, UK

11 ² Institute of Sport and Exercise Sciences, University of Münster, Germany

12 ³ Department of Respiratory Medicine, Royal Brompton Hospital, UK

13 ⁴ School of Sport, Health and Exercise Sciences, Loughborough University, UK

14

15

16

17 **Running Title:** The influence of sex on oxygen uptake and extraction

18

19

20

21 **Corresponding Author:**

22 Paul Ansdell, PhD

23 Department of Sport, Exercise and Rehabilitation

24 Faculty of Health and Life Sciences

25 Northumbria University

26 Newcastle upon Tyne

27 NE1 8ST

28 UK

29 p.ansdell@northumbria.ac.uk

30 **Abstract**

31 The integrative response to exercise differs between sexes, with oxidative energy
32 contribution purported as a potential mechanism. The present study investigated whether
33 this difference was evident in the kinetics of oxygen uptake ($\dot{V}O_2$) and extraction (HHb+Mb)
34 during exercise.

35 Sixteen adults (8 males, 8 females, age: 27 ± 5 years) completed three experimental visits.
36 Incremental exercise testing was performed to obtain lactate threshold and $\dot{V}O_{2peak}$.
37 Subsequent visits involved three six-minute cycling bouts at 80% of lactate threshold and
38 one 30-minute bout at a work rate 30% between the lactate threshold and power at $\dot{V}O_{2peak}$.
39 Pulmonary gas exchange and near-infrared spectroscopy of the vastus lateralis were used
40 to continuously sample $\dot{V}O_2$ and HHb+Mb, respectively. The phase II $\dot{V}O_2$ kinetics were
41 quantified using mono-exponential curves during moderate and heavy exercise. Slow
42 component amplitudes were also quantified for the heavy intensity domain.

43 Relative $\dot{V}O_{2peak}$ values were not different between sexes ($p=0.111$). Males achieved ~30%
44 greater power outputs ($p=0.002$). In the moderate and heavy intensity domains, the relative
45 amplitude of the phase II transition was not different between sexes for $\dot{V}O_2$ (~24 and ~40%
46 $\dot{V}O_{2peak}$, $p \geq 0.179$) and HHb+Mb (~20 and ~32% ischemia, $p \geq 0.193$). Similarly, there were no
47 sex differences in the time constants for $\dot{V}O_2$ (~28 s, $p \geq 0.385$) or HHb+Mb (~10s, $p \geq 0.274$).
48 In the heavy intensity domain, neither $\dot{V}O_2$ ($p \geq 0.686$) or HHb+Mb ($p \geq 0.432$) slow component
49 amplitudes were different between sexes.

50 The oxidative response to moderate and heavy intensity exercise did not differ between
51 males and females, suggesting similar dynamic responses of oxidative metabolism during
52 intensity-matched exercise.

53

54 **New and Noteworthy**

55 This study demonstrated no sex differences in the oxidative response to moderate and
56 heavy intensity cycling exercise. The change in oxygen uptake and deoxyhaemoglobin were
57 modelled with mono-exponential curve fitting, which revealed no differences in the rate of
58 oxidative energy provision between sexes. This provides insight into previously reported sex
59 differences in the integrative response to exercise.

60 Introduction

61 The transition from rest to exercise involves an integrated response from the pulmonary,
62 cardiovascular, and muscular systems to rapidly increase the supply and utilisation of
63 oxygen for oxidative adenosine triphosphate (ATP) provision (Poole & Jones, 2012). The
64 speed at which this process can occur can be quantified using pulmonary oxygen uptake
65 ($\dot{V}O_2$) kinetics, and is thought to determine metabolic stability and exercise tolerance across
66 the spectrum of athletic performance and disease (Burnley & Jones, 2007; Grassi *et al.*,
67 2011). The $\dot{V}O_2$ response can be broken down into three phases, beginning with the initial
68 cardio-dynamic phase (phase I, 10-20 s) which represents an increased venous return via
69 the muscle pump effect, as well as increased pulmonary blood flow (Grassi *et al.*, 1996).
70 Thereafter, increases in pulmonary $\dot{V}O_2$ are considered to reflect increased muscle oxygen
71 uptake in response to exercise (phase II), until the energy demand of exercise is met by
72 oxidative phosphorylation and $\dot{V}O_2$ reaches a steady state (Hughson *et al.*, 2001). A steady
73 state response is attainable quickly within the moderate intensity domain, whereas in either
74 heavy or severe intensity domains, a further rise in $\dot{V}O_2$ is observed before the steady state
75 is attained (heavy) or $\dot{V}O_{2max}$ is reached (severe), termed the slow component. This three-
76 phase response is ubiquitous in exercising humans; however, the biological characteristics
77 of the individual can influence the rates at which they occur (for review see Poole & Jones,
78 2012).

79

80 The time constant of phase II kinetics are considered to be a crucial determinant of the
81 decrease in contractile function experienced by the exercising individual (Temesi *et al.*,
82 2017; Goulding *et al.*, 2021), and could explain previously observed sex differences in the
83 integrative response to exercise (Ansdell *et al.*, 2020b). As oxidative phosphorylation does
84 not immediately meet the demand for ATP, substrate-level phosphorylation is required
85 (Burnley & Jones, 2007). Intuitively, the rate at which oxidative metabolism can be
86 upregulated at the onset of exercise is inversely linked with the accumulation of deleterious
87 metabolites such as hydrogen ions [H^+] and inorganic phosphate [P_i], as well as the
88 depletion of phosphocreatine [PCr] stores, which all interfere with excitation-contraction
89 coupling (Allen *et al.*, 2008). However, the relationship between the aforementioned
90 metabolites and the upregulation of oxidative phosphorylation is multi-faceted, with the
91 progressive change in the phosphate energy state (i.e., the [ATP]/[ADP]/[P_i] balance) driving
92 the rate at which mitochondrial respiration increases, whilst [H^+] accumulation concurrently
93 inhibits anaerobic glycolysis at the onset of exercise (Tschakovsky & Hughson, 1999;
94 Korzeniewski & Rossiter, 2015). Accordingly, Temesi *et al.* (2017) demonstrated a positive
95 correlation between the time constant ($\tau\dot{V}O_2$) of the phase II response and the decrease in
96 quadriceps potentiated twitch force. Similarly, elite endurance athletes demonstrate faster

97 $\tau\dot{V}O_2$ (Koppo *et al.*, 2004) and lesser declines in contractile function (Ducrocq *et al.*, 2021)
98 compared to untrained individuals when exercising at similar relative intensities.

99

100 A consistent finding in studies comparing males and females exercising at the same
101 metabolic intensity is that females experience a lesser degree of contractile impairment of
102 the knee-extensors (Ansdell *et al.*, 2019; Ansdell *et al.*, 2020a; Azevedo *et al.*, 2021).
103 Previously, this has been suggested to be a result of sex differences in skeletal muscle
104 composition, whereby females consistently demonstrate a greater proportional area of type I
105 fibres (Staron *et al.*, 2000; Roepstorff *et al.*, 2006). The consequences of this fibre type
106 difference are multi-factorial; for-instance, it is well established that type I fibres are more
107 fatigue-resistant (Schiaffino & Reggiani, 2011). Additionally, female *vastus lateralis* capillary
108 density is ~23% greater in females compared to males (Roepstorff *et al.*, 2006), while
109 females also demonstrate greater mitochondrial oxidative function and intrinsic respiratory
110 rates than males of equivalent training status (Cardinale *et al.*, 2018). One factor that
111 remains unexplored is whether these physiological sex differences result in differences in the
112 metabolic response to exercise. Conceivably, the superior aerobic phenotype of female
113 skeletal muscle could imply that females might be able to meet the ATP demand of exercise
114 through oxidative means faster than males. Despite this, *ex vivo* evidence suggests that
115 female skeletal muscle fibres have a lower ADP sensitivity of mitochondrial respiration
116 (Miotto *et al.*, 2018), which could result in a slower rate of oxidative phosphorylation at the
117 onset of exercise. Therefore, *in vivo* assessment of $\dot{V}O_2$ kinetics would provide insight into
118 the balance between these morphological and cellular sex differences.

119

120 The $\dot{V}O_2$ slow component is underpinned by different mechanisms to the phase II kinetics
121 and describes the increase in $\dot{V}O_2$ during constant-load exercise. This increase in $\dot{V}O_2$
122 implies an impairment of efficiency and is likely an amalgamation of several concurrent
123 physiological changes. Within skeletal muscle, the accumulation of metabolites (e.g., [Pi])
124 and associated contractile dysfunction is linked with the loss of efficiency (Grassi *et al.*,
125 2015). Of relevance here, is that female skeletal muscle has consistently been demonstrated
126 to be more fatigue-resistant (Ansdell *et al.*, 2019; Ansdell *et al.*, 2020a), and shows lesser
127 increases in the surface electromyogram in states of fatigue (Ansdell *et al.*, 2017; Ansdell *et al.*
128 *et al.*, 2019). Furthermore, it is well-established that females have a greater reliance on lipid
129 metabolism during sustained exercise at similar relative work rates (Cano *et al.*, 2022),
130 which could be related to the more oxidative phenotype of skeletal muscle, as described
131 above. Combined, these physiological sex differences could represent a slower loss of
132 efficiency during constant load exercise in females, however, this remains unexplored.

133

134 Despite more aerobically-suited skeletal muscle, females have lower levels of haemoglobin
135 (Murphy, 2014), which is thought to impair O₂ carrying capacity during exercise (Harms *et*
136 *al.*, 1998; Diaz-Canestro *et al.*, 2022). During exercise where O₂ delivery and utilisation are
137 both limiting factors (e.g., cycling, Goulding & Marwood, 2023), these factors are thought to
138 counteract each other to enable comparable relative metabolic thresholds (i.e., critical
139 power) between the sexes (Ansdell *et al.*, 2020b). To date, the only investigation to
140 systematically investigate the oxidative adjustment at the onset of exercise between sexes
141 did so during low intensity treadmill walking (Beltrame *et al.*, 2017). Data from this study
142 suggested faster O₂ extraction in females, quantified as the change in de-oxyhaemoglobin
143 and myoglobin (HHb+Mb) signal in near-infrared spectroscopy (NIRS), fitting with the notion
144 that phase II $\dot{V}O_2$ kinetics are influenced by intramuscular factors in healthy humans (Poole
145 & Jones, 2012). However, the demands of treadmill walking differ to those of high-intensity
146 cycling exercise, where it has recently been argued that all levels of the O₂ cascade are
147 considered to be influential in determining metabolic responses to exercise (Goulding &
148 Marwood, 2023). Thus, given the lack of evidence regarding physiological responses to
149 exercise in females (James *et al.*, 2023), it remains to be determined how sex differences in
150 convective and diffusive contributions to O₂ delivery mediate the rate of $\dot{V}O_2$ adjustment to
151 exercise.

152

153 Accordingly, the present study employed a multi-method approach of measuring pulmonary
154 gas exchange and near-infrared spectroscopy simultaneously to compare the kinetics of
155 pulmonary $\dot{V}O_2$ as well as muscle oxygen extraction in both sexes during moderate and
156 heavy intensity exercise. Previously this experimental approach has been used to obtain
157 information about O₂ delivery and utilisation to gain insights into the integrative response to
158 exercise (DeLorey *et al.*, 2003). It was hypothesised that females would demonstrate a
159 smaller value for the phase II time constant (i.e., faster kinetics) for $\dot{V}O_2$ and HHb+Mb at the
160 onset of exercise, and a smaller slow component amplitude in the heavy intensity domain.

161

162 **Methods**

163 *Ethical Approval*

164 This study received institutional ethical approval from the Northumbria University Health and
165 Life Sciences Research Ethics Committee (submission reference: 49189) and was
166 conducted according to all aspects of the Declaration of Helsinki, apart from pre-registration
167 in a database. Participants volunteered for the study and provided written informed consent.

168

169 *Participants*

170 Using the effect size for the sex difference in *vastus lateralis* tissue oxygenation during
171 heavy intensity exercise ($\eta^2 = 0.509$) from (Ansdell *et al.*, 2020a), an *a priori* sample size
172 calculation determined a minimum of 14 participants (seven females and seven males) were
173 required to detect an effect ($\alpha = 0.05$, power = 0.95). Therefore, eight males (mean \pm SD
174 age: 27 ± 3 years; stature: 182 ± 5 cm; body mass: 75.3 ± 10.2 kg) and eight females (mean
175 \pm SD age: 27 ± 7 ; stature: 163 ± 4 cm; body mass: 61.8 ± 5.9 kg) volunteered to take part in
176 the study. Hormonal status was not an exclusion criterion or controlled for in this study.
177 Female participants were tested in any phase of their menstrual cycle and there were no
178 restrictions on hormonal contraceptive usage. This decision was based on evidence from
179 (Mattu *et al.*, 2020) who demonstrated no hormonal effects on $\dot{V}O_2$ kinetics during cycling
180 exercise. Of the eight females, two were using combined oral contraceptive pills (Lucette
181 and Rigevidon) and six were naturally menstruating. All participants were free from
182 cardiovascular, respiratory, and neurological disease as well as musculoskeletal injury.

183

184

185 *Experimental Design*

186 All participants visited the laboratory on three occasions across an average of 10 ± 4 days
187 (range: 5 – 21 days). During the first visit, participants were familiarised with the
188 experimental procedures and completed two incremental exercise tests to quantify lactate
189 threshold and peak oxygen uptake ($\dot{V}O_{2peak}$). The second and third visits were identical and
190 involved three six-minute bouts of moderate intensity exercise (80% of lactate threshold, LT),
191 separated by six minutes of unloaded pedalling. Thereafter, a single bout of heavy intensity
192 exercise ($30\% \Delta LT - \dot{V}O_{2peak}$) was performed for 30 minutes.

193

194 *Visit 1: Familiarisation & Incremental Testing*

195 The first visit began with participants completing a screening questionnaire to ensure
196 inclusion criteria were met. Thereafter, participants moved onto the cycle ergometer
197 (Velotron, SRAM, Chicago, IL, USA) which was set up with the seat height aligned with the
198 hip, and handlebar height set according to the participants' comfort, these measurements

199 were recorded and replicated for subsequent trials. The breath-by-breath gas exchange
200 mask was then placed over the participant's mouth and nose, and an air-tight seal was
201 ensured before resting data was recorded. Following resting measures of pulmonary gas
202 exchange and muscle oxygenation, participants completed five minutes of warm-up cycling
203 at a light intensity (60 W) at a self-selected cadence between 70-100 rpm, before
204 commencing an incremental exercise test. The first incremental exercise test began at 75 W
205 and increased by 25 W every five minutes. At the end of each stage, a capillary blood
206 sample was drawn from the participants' fingertip and immediately analysed to determine
207 whole blood lactate concentration ($\text{mmol}\cdot\text{L}^{-1}$, Biosen C-Line, EKF Diagnostics, Germany).
208 The test was terminated once LT was identified as the first work rate at which a non-linear
209 increase in blood lactate concentration was observed (Faude *et al.*, 2009), after which,
210 participants were provided 20 minutes of passive rest.

211

212 Next, participants began the second incremental test with five minutes of warm-up cycling at
213 a light intensity (60 W) at the same self-selected cadence as before. Thereafter, a ramp test
214 beginning at 75 W commenced, with power output increasing 1 W every 2.4 seconds (25
215 $\text{W}\cdot\text{min}^{-1}$). This test was terminated at volitional exhaustion, defined as cadence falling >10
216 rpm for five seconds. Strong verbal encouragement was provided to participants throughout.
217 A final blood lactate sample was drawn immediately after volitional exertion. The greatest 30
218 second average $\dot{V}\text{O}_2$ value was used to quantify $\dot{V}\text{O}_{2\text{peak}}$, whilst the final power output was
219 used to quantify maximal ramp test power (P_{max}).

220

221 *Visits 2 & 3: Square-Wave Exercise Bouts*

222 Visits two and three were identical, and performed with a minimum of 24 h between visits.
223 The visits involved continuous sampling of pulmonary gas exchange and near infrared
224 spectroscopy (NIRS) of the *vastus lateralis*. Trials commenced with participants performing
225 three minutes of unloaded pedalling on the cycle ergometer. Thereafter, participants
226 performed three repetitions of six-minute cycling bouts at 80% of the work rate associated
227 with LT (moderate intensity exercise), interspersed with six minutes of unloaded pedalling.
228 Following this, participants cycled for 30 minutes at a work rate 30% between the LT and
229 $\dot{V}\text{O}_{2\text{peak}}$ (30% Δ , heavy intensity exercise). Throughout this visit, participants were asked to
230 replicate their self-selected cadence from visit 1, which was monitored by an experimenter
231 throughout. Exercise intensity was altered abruptly in a 'square-wave' fashion for each
232 repetition.

233

234 Immediately following all exercise, a 'physiological calibration' of NIRS signals was
235 performed as per recommendations from Barstow (2019). Participants were laid supine on a

236 physiotherapy table, with their leg placed horizontal, and an automatic personalised
237 tourniquet system for blood flow restriction (Delfi Medical Innovations Inc., Vancouver BC,
238 Canada) was placed around the thigh via a nylon cuff (11.5 cm × 86 cm, 5 mm thick),
239 proximal to the NIRS optode. The cuff was inflated for five minutes at 120% of limb occlusion
240 pressure in order to occlude blood flow, and the mean pressure was not different between
241 males and females (120% limb occlusion pressure: 248 ± 20 vs. 249 ± 30 mmHg, $p =$
242 0.780). This system automatically measures limb occlusion pressure, defined as the
243 minimum pressure required for complete restriction of arterial blood flow in a limb, and
244 maintains the pressure during inflation to ensure consistent occlusion. Following this,
245 pressure in the cuff was released and the hyperaemic response measured (see *Near*
246 *Infrared Spectroscopy*). This protocol allowed all NIRS data to be expressed as a % of an
247 individual's physiological minimum and maximum values. Physiological calibration negates
248 any potential influence of adipose tissue thickness on NIRS data (Barstow, 2019).

249

250 *Pulmonary Gas Exchange*

251 During all visits, expired gas was analysed breath-by-breath using an online system (Vyntus
252 CPX, Jaeger, CareFusion, Germany). Oxygen (O_2) and carbon dioxide (CO_2) concentrations
253 were quantified via a paramagnetic chemical fuel cell and non-dispersive infrared cell
254 respectively. Before each test, the analysers were calibrated using ambient air and a gas of
255 known O_2 (15.00%) and CO_2 (4.97%) concentrations. Ventilatory volumes were inferred from
256 measurement of gas flow using a digital turbine transducer (volume 0 to 10 L, resolution 3
257 mL, flow 0 to $15 \text{ L} \cdot \text{s}^{-1}$) and calibrated prior to each test (Hans Rudolph Inc. Kansas City,
258 USA).

259

260 *Near Infrared Spectroscopy*

261 A multi-distance, continuous-wave, single channel NIRS (NIRO-200NX, Hamamatsu) was
262 used to evaluate changes in vastus lateralis muscle deoxyhaemoglobin and myoglobin
263 (HHb+Mb), as well as oxyhaemoglobin and myoglobin (HbO_2+MbO_2) concentrations,
264 sampled at a rate of 5 Hz. Tissue oxygenation index (TOI) was calculated as $HbO_2+MbO_2 \div$
265 $[HbO_2+MbO_2 + HHb+Mb] \times 100$. The light-emitting probe comprised of light emitting diodes
266 operating at three wavelengths (735, 810, and 850 nm). The probe was placed on the *vastus*
267 *lateralis*, 20 cm above the fibular head. Optodes were held in place by an elasticised
268 bandage and covered by an opaque, dark material to avoid motion and ambient light
269 influences.

270

271 *Data Analysis*

272 *$\dot{V}O_2$ Kinetics*

273 The breath by breath data was manually filtered to remove outlying breaths, defined as
274 breaths deviating more than 500 ml.min⁻¹ from the mean value from the preceding five
275 breaths. Thereafter, breath by breath data was linearly interpolated to provide second-by-
276 second values. The multiple repetitions of the square-wave exercise bouts were then
277 averaged, and $\dot{V}O_2$ responses were time aligned to the onset of exercise. Data from the
278 onset of the transition to 20 s was removed, then the resultant data was modelled with a
279 monoexponential curve, including data from -60 to 360 seconds (moderate) or -60 to 120
280 seconds (heavy), with the following equation:

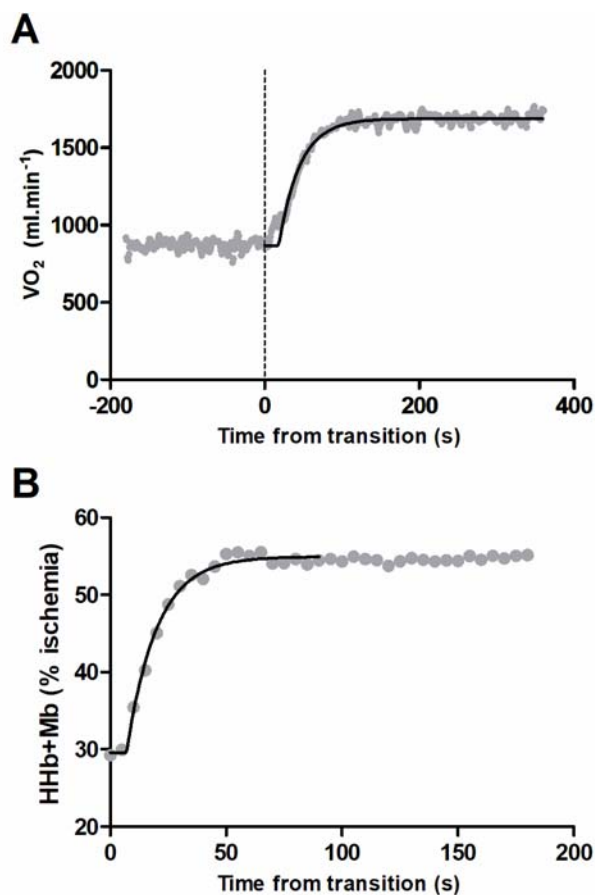
281

$$282 \quad \dot{V}O_2(t) = \dot{V}O_2(b) + A_p(1 - e^{-(t-TD_p)/\tau_p})$$

283

284 Where $\dot{V}O_2(t)$ is the $\dot{V}O_2$ at time t ; $\dot{V}O_2(b)$ is the baseline $\dot{V}O_2$ measured in the 60 s
285 preceding the transition in work rate; and A_p , TD_p , and τ_p are the amplitude, time delay, and
286 the time constant of the phase II response, respectively. We chose to constrain the
287 modelling of heavy intensity domain onset kinetics to 120 seconds in order to minimise the
288 influence of the slow component, however this cannot be guaranteed (Burnley *et al.*, 2006).
289 For exercise in the heavy intensity domain, the amplitude of the $\dot{V}O_2$ slow component was
290 determined by subtracting the phase II amplitude from the highest 30 s average of $\dot{V}O_2$
291 during the 30 min bout (Rossiter *et al.*, 2001). To facilitate comparisons between sexes,
292 amplitudes were also normalised to each individual's $\dot{V}O_{2peak}$ and end-exercise $\dot{V}O_2$ as well
293 as being presented in L.min⁻¹. The O₂ cost of the transitions was estimated by calculating the
294 $\dot{V}O_2$ gain (Porcelli *et al.*, 2016), where the amplitude of the phase II response was divided by
295 work rate (ml.min⁻¹.W⁻¹).

296



297

Figure 1: Visualisation of the monoexponential curve fitting procedures for a representative participant's data in the moderate intensity domain. Panel A describes the $\dot{V}O_2$ data (1 Hz) and Panel B describes the HHb+Mb data (0.2 Hz).

298

299 Deoxyhaemoglobin Kinetics

300 Prior to curve fitting, data from NIRS were normalised to the minimum values during, and the
 301 maximum values following the five minute arterial occlusion (Ryan *et al.*, 2012), then
 302 averaged into 1 s and 5 s bins. The multiple repetitions of the square-wave exercise bouts
 303 were then averaged, and HHb+Mb responses were time aligned to the onset of exercise.
 304 The TD for the HHb+Mb response was determined using the 1 s averaged data as the time
 305 between exercise onset and the first point at which HHb+Mb signal started to systematically
 306 increase. This was performed for each transition individually, with all TDs averaged to
 307 provide a single value. The 5 s averaged data was then modelled with a monoexponential
 308 curve in the same manner as $\dot{V}O_2$ data, including data up to 90 s after the transition (Murias
 309 *et al.*, 2010). Other NIRS-derived variables (TOI and HbO₂+MbO₂) were quantified as 30 s
 310 averages at the following time points: the 30s of unloaded pedalling immediately prior to the
 311 transition, 90 s following the transition (heavy intensity domain only), and the final 30s of the
 312 square wave bout of exercise.

313

314 *Statistical Analysis*

315 Data are presented as mean \pm SD within the text and figures. Normal distribution of data
316 was confirmed with the Shapiro-Wilk test. As all variables had normally distributed data,
317 males and females were compared with independent samples t tests for variables with a
318 single value or time point. For repeated measures variables during exercise, two-way (sex \times
319 time) repeated measures ANOVA were performed, followed by Bonferroni-corrected post
320 hoc tests if significant main effects were observed. Effect sizes for comparisons were
321 calculated as Cohen's *d*. The significance level for all statistical tests was set at $p < 0.05$.

322

323 **Results**

324 *Incremental Exercise Testing*

325 Anthropometric data and outcome variables from the two incremental exercise tests
 326 performed in the first visit are presented in Table 1. As expected, males had a greater
 327 stature and body mass than females ($p \leq 0.006$) as well as a greater absolute $\dot{V}O_{2peak}$ (mean
 328 difference: 39%, $p = 0.002$). However, when $\dot{V}O_{2peak}$ was expressed relative to body mass,
 329 no sex difference was observed (mean difference: 14%, $p = 0.111$). Males also exercised at
 330 greater power outputs than females, with Pmax and LT being ~30% greater in males ($p \leq$
 331 0.023), however when LT was expressed as a % of Pmax, no sex difference was observed
 332 ($p = 0.373$). This resulted in the power outputs for the moderate and heavy intensity bouts
 333 being greater in males compared to females ($p \leq 0.023$).

334

335 *Table 1: Anthropometric data and outcome variables from incremental exercise testing.*

	Males (n = 8)	Females (n = 8)	P value	Cohen's d
Age (years)	27 ± 3	27 ± 7	0.734	0.117
Stature (cm)	182 ± 5	163 ± 4	< 0.001	1.383
Body Mass (kg)	75.3 ± 10.2	61.8 ± 5.9	0.006	0.501
$\dot{V}O_{2peak}$ (L.min ⁻¹)	3.47 ± 0.58	2.50 ± 4.42	0.002	0.627
Relative $\dot{V}O_{2peak}$ (ml.kg ⁻¹ .min ⁻¹)	46.2 ± 6.6	40.5 ± 6.7	0.111	0.326
Pmax (W)	328 ± 54	236 ± 43	0.002	0.646
Pmax (W.kg ⁻¹)	4.4 ± 0.8	3.8 ± 0.5	0.118	0.253
Power at LT (W)	153 ± 25	119 ± 29	0.023	0.528
LT (% Pmax)	47 ± 6	50 ± 7	0.373	0.183
LT (W.kg ⁻¹)	2.0 ± 0.2	1.9 ± 0.4	0.440	0.127
80% LT (W)	123 ± 20	95 ± 23	0.023	0.528
80% LT (W.kg ⁻¹)	1.6 ± 0.2	1.5 ± 0.3	0.440	0.110
30% Δ (W)	206 ± 30	154 ± 32	0.005	0.653
30% Δ (W.kg ⁻¹)	2.7 ± 0.4	2.5 ± 0.4	0.198	0.213

LT: lactate threshold, Pmax: Maximal ramp test power output, $\dot{V}O_{2peak}$: Maximal rate of oxygen consumption.

336

337

338

339 $\dot{V}O_2$ Kinetics

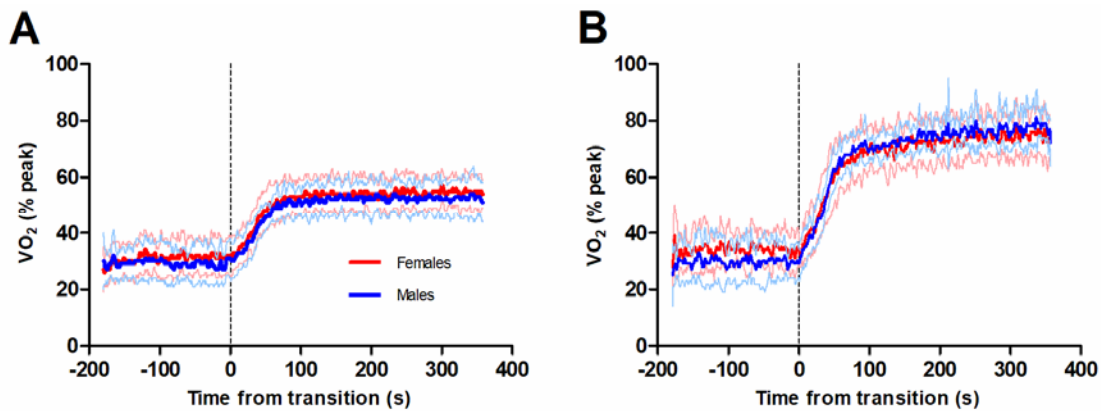
340 The transition from unloaded pedalling to moderate and heavy intensity cycling elicited an
 341 increase in $\dot{V}O_2$ (see Figure 2), and the monoexponential curve used to describe the
 342 increase in $\dot{V}O_2$ in males and females demonstrated excellent r^2 values (see Table 2).

343 *Table 2: Data from the monoexponential modelling of $\dot{V}O_2$ kinetics during moderate and heavy*
 344 *intensity transitions.*

	Males (n = 8)	Females (n = 8)	P value	Cohen's d
<i>Moderate Intensity Domain</i>				
TD (s)	16.6 ± 4.1	17.9 ± 4.9	0.575	0.121
Baseline $\dot{V}O_2$ (L.min ⁻¹)	0.97 ± 0.05	0.77 ± 0.09	< 0.001	1.516
Amplitude (L.min ⁻¹)	0.83 ± 0.19	0.60 ± 0.18	0.024	0.477
Amplitude (ml.kg ⁻¹ .min ⁻¹)	11.0 ± 1.6	9.7 ± 2.6	0.230	0.323
Amplitude (% $\dot{V}O_{2peak}$)	24 ± 3	24 ± 5	0.949	0.018
$\dot{V}O_2$ gain (ml.min ⁻¹ .W ⁻¹)	10.2 ± 0.7	10.5 ± 1.1	0.587	0.130
τ (s)	27.9 ± 7.5	24.8 ± 6.6	0.385	0.160
r^2	0.965 ± 0.032	0.947 ± 0.034		
<i>Heavy Intensity Domain</i>				
TD (s)	15.0 ± 4.8	14.5 ± 4.5	0.858	0.033
Baseline $\dot{V}O_2$ (L.min ⁻¹)	1.00 ± 0.06	0.85 ± 0.11	0.005	0.894
Amplitude (L.min ⁻¹)	1.50 ± 0.38	0.96 ± 0.25	0.005	0.541
Amplitude (ml.kg ⁻¹ .min ⁻¹)	19.9 ± 4.7	15.6 ± 3.7	0.060	0.351
Amplitude (% $\dot{V}O_{2peak}$)	43 ± 5	38 ± 7	0.179	0.355
$\dot{V}O_2$ gain (ml.min ⁻¹ .W ⁻¹)	9.4 ± 0.8	9.3 ± 0.9	0.766	0.059
τ (s)	28.8 ± 7.9	27.2 ± 4.4	0.633	0.075
r^2	0.961 ± 0.031	0.927 ± 0.030		
SC Amplitude (L.min ⁻¹)	0.38 ± 0.16	0.28 ± 0.12	0.158	0.250
SC Amplitude (% $\dot{V}O_{2peak}$)	11.9 ± 6.9	10.8 ± 3.2	0.686	0.061
SC Amplitude (% end exercise $\dot{V}O_2$)	13.6 ± 6.5	12.9 ± 3.9	0.822	0.035

SC: slow component, τ : time constant, TD: time delay, $\dot{V}O_2$: rate of oxygen consumption

345



346

347 *Figure 2: Group mean $\dot{V}O_2$ data from males (blue, $n=8$) and females (red, $n=8$) during moderate*
348 *(Panel A) and heavy (Panel B) intensity transitions. The bold lines represent group means and the*
349 *thin lines represent standard deviation.*

350

351 In absolute units ($L \cdot \text{min}^{-1}$) males experienced greater amplitudes of $\dot{V}O_2$ during the phase II
352 kinetics ($p \leq 0.024$), however when this was made relative to the individual ($\dot{V}O_2$ gain and %
353 $\dot{V}O_{2\text{peak}}$) no sex differences were observed ($p \geq 0.179$). Similarly, the $\dot{V}O_2$ slow component
354 amplitude was not different between sexes in relative units ($p = 0.686$). As visualised in
355 Figure 1, there were no sex differences in $\tau\dot{V}O_2$ in either the moderate ($p = 0.385$) or heavy
356 ($p = 0.633$) intensity domains. Baseline $\dot{V}O_2$ was slightly elevated at the onset of heavy
357 intensity exercise compared to the onset of moderate intensity exercise for females (mean
358 difference: $0.08 L \cdot \text{min}^{-1}$, $p = 0.028$), whereas male baseline $\dot{V}O_2$ was not different ($p =$
359 0.312).

360

361 *Oxygen Extraction Kinetics*

362 The transition from unloaded pedalling to moderate and heavy intensity cycling elicited an
363 increase in HHb+Mb concentration (Figure 3), and the monoexponential curve used to
364 describe the increase in HHb+Mb in males and females demonstrated excellent r^2 values
365 (Table 3). One female's data had to be removed due to issues with the NIRS signal,
366 resulting in $n = 7$ females being used for NIRS analyses.

367

368 Table 3: Data from the monoexponential modelling of deoxyhaemoglobin kinetics during moderate
 369 and heavy intensity transitions.

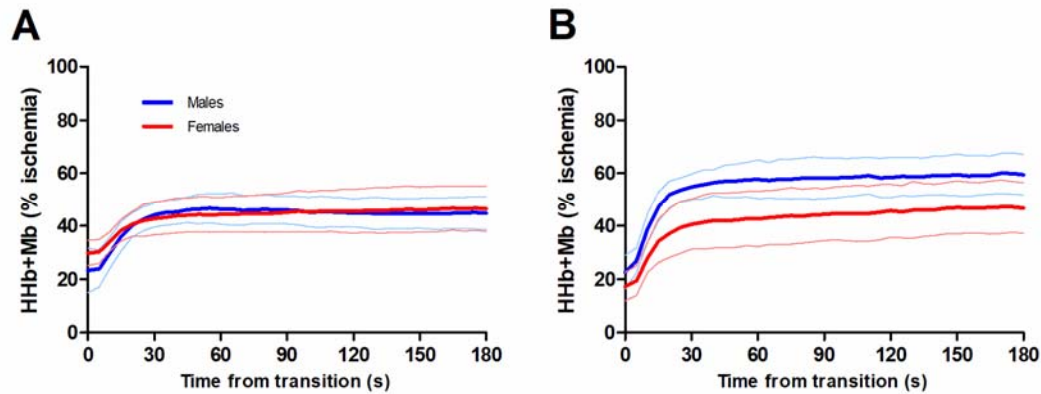
	Males (n = 8)	Females (n = 7)	P value	Cohen's d
<i>Moderate Intensity Domain</i>				
TD (s)	8.7 ± 1.5	9.6 ± 2.6	0.447	0.212
Baseline HHb+Mb (% ischemia)	23 ± 8	29 ± 7	0.186	0.251
Amplitude (% ischemia)	21 ± 7	17 ± 5	0.225	0.219
τ (s)	8.1 ± 2.8	10.0 ± 3.8	0.274	0.260
r ²	0.974 ± 0.022	0.956 ± 0.037		
<i>Heavy Intensity Domain</i>				
TD (s)	6.2 ± 2.8	7.0 ± 2.3	0.582	0.101
Baseline HHb+Mb (% ischemia)	23 ± 6	17 ± 7	0.146	0.332
Amplitude (% ischemia)	36 ± 9	30 ± 8	0.193	0.252
τ (s)	11.3 ± 3.7	12.2 ± 4.0	0.665	0.090
r ²	0.983 ± 0.011	0.979 ± 0.023		
SC Amplitude (% ischemia)	14 ± 5	16 ± 4	0.432	0.142

HHb+Mb: deoxyhaemoglobin and myoglobin, SC: slow component, τ: time constant, TD: time delay consumption

370

371 As shown in Figure 3, the phase II amplitude of HHb+Mb increase in both intensity domains
 372 was not different between sexes ($p \geq 0.193$). Similarly, there was no sex difference in the
 373 amplitude of the slow component in the heavy intensity domain ($p = 0.432$). The time
 374 constant for phase II HHb+Mb kinetics ($\tau_{\text{HHb+Mb}}$) was also not different between males and
 375 females in both intensity domains ($p \geq 0.274$). Baseline HHb+Mb was slightly lower at the
 376 onset of heavy intensity exercise for females compared to the onset of moderate exercise
 377 (mean difference: 12% ischemia, $p = 0.002$), but not for males ($p = 0.583$).

378



379

380 *Figure 3: Group mean HHb+Mb data from males (blue, n=8) and females (red, n=7) during moderate*
 381 *(Panel A) and heavy (Panel B) intensity transitions. The bold lines represent group means and the*
 382 *thin lines represent standard deviation.*

383

384 *Near-infrared spectroscopy*

385 During the moderate intensity cycling, a significant effect of sex was observed for
 386 $\text{HbO}_2 + \text{MbO}_2$ ($F_{1,13} = 10.85, p = 0.006$), but no time ($p = 0.804$) or sex \times time interaction ($p =$
 387 0.054) effects were observed. For TOI, significant time ($F_{1,13} = 48.59, p < 0.001$) and sex
 388 ($F_{1,13} = 12.57, p = 0.004$) effects were observed, but no sex \times time interaction effect ($p =$
 389 0.095). Post-hoc tests revealed that females had greater values before (6%, $p = 0.002$) and
 390 during the stage (10%, $p = 0.007$). However, when TOI values were normalised as %
 391 ischemia, there was a main effect of time ($F_{1,6} = 115.09, p < 0.001$), but neither the main
 392 effect of sex ($p = 0.181$) or the sex \times time interaction effect ($p = 0.381$) were evident.

393

394 *Table 4: Data from near-infrared spectroscopy before and during the moderate and heavy intensity*
 395 *exercise transitions in males (n = 8) and females (n = 7).*

		Moderate Intensity		Heavy Intensity		
		Unloaded Pedalling	End Stage	Unloaded Pedalling	90 secs	End Stage
HbO₂ + MbO₂ (%ischemia)	Male	57 ± 6	53 ± 7	68 ± 11*	54 ± 10**	47 ± 10#
	Female	41 ± 10	44 ± 9	51 ± 7	41 ± 4#	53 ± 8
TOI (%)	Male	69 ± 3	61 ± 5#	71 ± 4	57 ± 6#	50 ± 8#§
	Female	75 ± 3*	71 ± 6**	78 ± 4*	70 ± 8**	69 ± 9**
TOI (% ischemia)	Male	72 ± 4	60 ± 7#	76 ± 6	53 ± 9#	39 ± 10#§
	Female	67 ± 9	53 ± 10#	77 ± 6	52 ± 9#	51 ± 17#

HbO₂+MbO₂: oxygenated haemoglobin and myoglobin; TOI: tissue oxygenation index; * = greater than the opposite sex ($p < 0.05$); # = lower than unloaded pedalling ($p < 0.05$); § = lower than 90 secs.

396

397 During heavy intensity cycling, a main effect of time was observed for HbO₂+MbO₂ ($F_{2,26} =$
398 $17.39, p < 0.001$), as well as sex \times time interaction effect ($F_{2,26} = 15.67, p < 0.001$), but no
399 main effect of sex ($p = 0.052$). Post-hoc tests revealed that females had lower values before
400 (-17% ischemia, $p = 0.004$) and 90 seconds after the transition (-13% ischemia, $p = 0.005$),
401 but not at the end of the stage ($p = 0.216$). Furthermore, HbO₂+MbO₂ decreased in both
402 sexes from unloaded pedalling to 90 seconds into the transition ($p < 0.001$), but for females,
403 this returned to baseline by the end of the stage ($p = 0.056$) whereas males remained
404 decreased ($p = 0.002$). For TOI, main effects of time ($F_{2,26} = 70.21, p < 0.001$) and a sex \times
405 time interaction effect ($F_{2,26} = 15.67, p < 0.001$) were observed, but no main effect of sex ($p =$
406 0.052). Post-hoc tests revealed that females had greater values than males at all timepoints
407 ($p \leq 0.005$). Additionally, while males demonstrated a progressive decrease in TOI at each of
408 the three time points ($p \leq 0.001$), females only decreased from unloaded pedalling to 90
409 seconds ($p < 0.001$), then no further decrease was observed at 30 minutes ($p = 1.000$).
410 When TOI was normalised to % ischemia, a main effect of time ($F_{1,37,17.78} = 49.50 p < 0.001$)
411 remained, however the sex ($p = 0.266$) and sex \times time interaction effect ($p = 0.112$) were not
412 observed.

413

414 **Discussion**

415 This study aimed to compare the kinetics of $\dot{V}O_2$ and HHb+Mb during moderate and heavy
416 intensity exercise in males and females. In contrast to the hypothesis, at the onset of
417 exercise the phase II time constants (τ) for $\dot{V}O_2$ and HHb+Mb were not different between the
418 sexes, implying that both males and females were able to increase oxidative phosphorylation
419 at comparable rates. In absolute units, males had larger amplitude increases than females,
420 however when normalised to the individuals' maximum values, the rise in $\dot{V}O_2$ and HHb+Mb
421 was not different. Combined, these data demonstrate that the oxidative response to exercise
422 is not different between sexes, which provides mechanistic insight into previously observed
423 sex differences in the integrative response to exercise.

424

425 Previous literature investigating sex differences in the onset kinetics of oxygen transport and
426 utilisation conflicts with the present data, with Beltrame *et al.* (2017) demonstrating quicker
427 $\tau\dot{V}O_2$ and $\tau\text{HHb+Mb}$ in females compared to males. One potential explanation for this
428 discrepancy could be that Beltrame *et al.* utilised a treadmill walking task, compared to
429 cycling. In tasks where O₂ delivery is not a limiting factor, females often outperform males.
430 For instance, Ansdell *et al.* (2019) showed female knee-extensors had a greater relative
431 critical torque than males during single-limb exercise. Whereas during cycling, where O₂
432 delivery is a determinant of critical power (Goulding & Marwood, 2023), this metabolic

433 threshold was not different between sexes (Ansdell *et al.*, 2020a). Whilst consensus on
434 whether O₂ delivery does (Hughson *et al.*, 2001) or does not (Grassi, 2001) limit $\tau\dot{V}O_2$ has
435 not been reached, it is conceivable that during tasks where O₂ delivery and utilisation are
436 both determinants in the metabolic response to exercise, the superior female skeletal muscle
437 oxidative capacity (Cardinale *et al.*, 2018) and vasodilatory response to exercise (Parker *et al.*,
438 *et al.*, 2007) is counteracted by an inferior O₂ carrying capacity (Murphy, 2014). Within the
439 present data, this balance manifests as a comparable $\tau\dot{V}O_2$ in males and females, which
440 agrees with data from do Nascimento Salvador *et al.* (2019), who demonstrated no sex
441 difference in $\tau\dot{V}O_2$ during a transition from unloaded pedalling to 'very heavy' (60% Δ) cycling
442 exercise.

443

444 Data from incremental exercise suggests that the poorer O₂ delivery in females results in a
445 greater degree of O₂ extraction to compensate (Murias *et al.*, 2013). The present data
446 contradicts this notion, as the amplitude of phase II HHb+Mb kinetics was not different
447 between sexes (see Table 3). However, it is important to note that Murias *et al.* noted that
448 this sex difference only occurred once incremental exercise exceeded the respiratory
449 compensation point (i.e., the severe intensity domain), whereas the present study compared
450 sexes in the moderate and heavy intensity domains. The lack of a sex difference in the
451 phase II amplitude for HHb+Mb kinetics contradicts previously published NIRS data that
452 demonstrated a smaller rise in HHb+Mb and lesser decrease in TOI in females compared to
453 males during constant-load exercise (Ansdell *et al.*, 2020a). The crucial difference in
454 methodologies employed between the previous study and the present study is the
455 application of a 'physiological calibration' to negate the influence of adipose tissue thickness
456 on NIRS signals (Ryan *et al.*, 2012). Previously, the sex difference in the rise in HHb+Mb
457 was suggested to reflect a lower oxygen cost of muscle contraction in female knee-
458 extensors, however the present data, with more rigorous methodologies employed, refutes
459 this. One aspect of the modelling that did demonstrate a sex difference was the reduction in
460 HHb+Mb (and concomitant increase in $\dot{V}O_2$) baseline for the heavy intensity transition in
461 females, but not males. This could reflect a sex difference in how O₂ utilisation is altered by
462 prolonged or intermittent exercise (i.e., the preceding three bouts of moderate intensity
463 cycling); however, the present study was not configured in a manner appropriate to answer
464 that research question. Females did demonstrate lower HbO₂+MbO₂ levels during heavy
465 intensity cycling, perhaps indicating a lesser O₂ availability. This would fit with the notion that
466 O₂ carrying capacity is inferior in females during high-intensity exercise (Diaz-Canestro *et al.*,
467 *et al.*, 2022); however, the lack of difference in the speed and amplitude of HHb+Mb onset
468 kinetics (and pulmonary $\dot{V}O_2$) implies that oxygen extraction is not negatively affected by this
469 in the moderate and heavy intensity domains.

470

471 Although not measured in the present study, the sex difference in muscle fibre type, whereby
472 females demonstrate a greater proportional area of type I fibres (Staron *et al.*, 2000;
473 Roepstorff *et al.*, 2006), appears to have not influenced either $\dot{V}O_2$ or HHb+Mb onset kinetics
474 in the present study. This would concur with data from Barstow *et al.* (1996), who
475 demonstrated no relationship between type I fibre percentage of the *vastus lateralis* and the
476 time constant of phase II $\dot{V}O_2$ kinetics. In contrast, Pringle *et al.* (2003) observed a negative
477 correlation between type I fibre percentage and the phase II time constant in the heavy
478 intensity domain only. Of note is that Pringle *et al.* included a wide range of participants with
479 ~27 – 85% type I fibres, and when groups were split into discrete groups of low and high
480 fibre type percentages (mean difference: 25%), the high percentage group had faster phase
481 II kinetics. The present study was not able to quantify the sex difference in muscle fibre
482 typology, however, previous literature has observed a 5-13% difference in type I fibre
483 percentage of the *vastus lateralis* (Simoneau & Bouchard, 1989; Staron *et al.*, 2000;
484 Roepstorff *et al.*, 2006). Therefore, it could be the case that the sex difference in muscle
485 fibre typology is not large enough to affect the phase II $\dot{V}O_2$ or HHb+Mb kinetics. Indeed,
486 recent evidence suggests that during exercise normalised to metabolic thresholds, sex
487 differences in muscle fibre typology do not influence fatigability (McDougall *et al.*, 2023).

488

489 Muscle fibre typology has previously been demonstrated to affect the amplitude of the slow
490 component within the heavy intensity domain, as individuals with a lower type I fibre
491 percentage experience larger rises in $\dot{V}O_2$ during constant-load exercise (Barstow *et al.*,
492 1996; Pringle *et al.*, 2003). It is suggested that the slow component is mechanistically
493 underpinned by factors such as additional motor unit recruitment (Poole *et al.*, 1994; Burnley
494 *et al.*, 2002) to compensate for fatigue-related changes in muscle metabolism. For instance,
495 muscle PCr stores demonstrate a similar slow component in depletion during heavy intensity
496 exercise (Rossiter *et al.*, 2002). Given that female knee-extensors appears more fatigue-
497 resistant and demonstrate lesser rises in the amplitude of surface electromyography during
498 constant-load exercise (Ansdell *et al.*, 2017; Ansdell *et al.*, 2019; Ansdell *et al.*, 2020a), we
499 hypothesised that the relative amplitude of the slow component would be greater in males to
500 reflect a greater rate of metabolic disturbance. However, as is evident in Tables 2 and 3, no
501 sex difference was observed in the relative slow component amplitude, implying that there
502 was no difference in the metabolic response to constant-load exercise.

503

504 The lack of sex differences in either the phase II kinetics or slow component amplitude
505 collectively suggest that the oxidative response to exercise was not different between males
506 and females. Data on this topic is sparse, and due to the nature of methods such as

507 magnetic resonance spectroscopy (MRS), limited to single-joint, isometric muscle
508 contractions. Previous literature using this technique to study muscle metabolic changes
509 during a 60 s contraction of the dorsiflexors showed no sex difference in changes in PCr, Pi,
510 or pH (Russ *et al.*, 2005). Data from muscle biopsies of the *vastus lateralis* taken before and
511 after repeated 30s cycling sprints suggested a greater preservation of ATP concentrations in
512 females across a ~60 minute protocol (Esbjörnsson-Liljedahl *et al.*, 2002); however the
513 authors suggested that this was likely a result of sex differences in the 20 minute recovery
514 periods, rather than metabolic differences during exercise. Accordingly, the same group
515 observed no sex differences in the metabolic response to a single 30-second cycling sprint
516 (Esbjörnsson-Liljedahl *et al.*, 1999). Collectively, across multiple tasks and methodologies
517 (MRS, biopsy and $\dot{V}O_2$ kinetics), the data suggest that there is no sex difference in the
518 bioenergetic response to high-intensity exercise. This information provides mechanistic
519 insight into the sex differences in the integrative response to exercise. For instance, sex
520 differences in fatigability have partly been attributed to a lesser accumulation of fatiguing
521 metabolites (Hunter, 2014; Ansdell *et al.*, 2020b). It is perhaps more accurate to suggest that
522 previously observed sex differences in fatigue during intensity-matched exercise (Ansdell *et al.*,
523 *et al.*, 2020a; Azevedo *et al.*, 2021) are more likely due to a greater fatigue-resistance of
524 female muscle contractile apparatus, which experience similar degrees of metabolic stress
525 as males. It is established that males and females differ in contractile properties such as
526 calcium (Ca^{2+}) kinetics of the sarcoplasmic reticulum (Harmer *et al.*, 2014), with lower
527 Ca^{2+} ATPase activity thought to permit a more fatigue-resistant skeletal muscle profile during
528 equivalent exercise tasks (Hunter, 2014). Therefore, the present study advances the
529 contemporary understanding of sex differences in the integrative response to exercise and
530 provides mechanistic insight into previously observed phenomena.

531

532 These data have applications across the spectrum of health and disease. For example,
533 those prescribing steady state exercise to improve skeletal muscle performance in athletes
534 or patients might not need to account for the sex of their participants (Gloeckl *et al.*, 2022;
535 Furrer *et al.*, 2023). This statement should however, be caveated by the fact that evidence
536 regarding the influence of sex on long-term adaptation to exercise is sparse (Ansdell *et al.*,
537 2020b). Indeed, one area for further exploration is the bioenergetic response to exercise
538 within the severe intensity domain, where sex differences in fatigability have previously been
539 observed (Ansdell *et al.*, 2020a; Azevedo *et al.*, 2021). The employment of complementary
540 techniques to quantify O_2 delivery (for instance, the present study did not quantify total
541 Hb+Mb) and muscle fibre typology could also provide greater insight into the influence of sex
542 on the O_2 cascade in a variety of tasks.

543

544 Previous evidence from pre-pubertal children and adolescents suggests that boys have a
545 lower $\tau\dot{V}O_2$ and $\dot{V}O_2$ slow component than girls (Armstrong & Barker, 2009), which
546 contradicts the present findings in adults. Previous studies comparing children and adults
547 also found that intramuscular PCr kinetics were similar in males and females regardless of
548 age (Willcocks *et al.*, 2010), implying similar oxidative capacity. Interestingly, the same study
549 found a greater 'PCr cost' ($\text{mM}\cdot\text{W}^{-1}$) in females compared to males suggesting a greater
550 inefficiency in females, which contradicts the present findings. Caution is urged when
551 comparing data from Willcocks *et al.* (2010) and the present study, however, given the
552 nature of exercise (single joint vs. whole body), the lack of matching for aerobic fitness, and
553 the low sample size (6 males vs. 5 females).

554

555 Finally, hormonal status was not an exclusion criterion or controlled for within female
556 participants in the present study. We based this decision on evidence from Mattu *et al.*
557 (2020) who demonstrated that $\dot{V}O_2$ kinetics did not differ between the follicular and luteal
558 phases of the eumenorrheic menstrual cycle, or with oral contraceptive usage. However, we
559 do acknowledge that the aforementioned study only investigated the moderate intensity
560 domain, and therefore might not apply to $\dot{V}O_2$ kinetics in the heavy intensity domain. In the
561 heavy intensity domain, contributing factors to the $\dot{V}O_2$ slow component (e.g., substrate
562 utilisation) might be affected by hormonal status, although the available evidence is
563 conflicting (Oosthuysen *et al.*, 2023). Therefore, further research is required to determine
564 whether endogenous and exogenous hormones influence the oxidative response to high-
565 intensity exercise.

566

567 **Conclusion**

568 The present study aimed to compare the oxygen extraction and uptake kinetics during
569 moderate and heavy intensity cycling exercise. Contrary to our hypotheses, no sex
570 differences were observed in either the phase II or slow component kinetics for $\dot{V}O_2$ or
571 HHb+Mb. The lack of sex difference implies that males and females do not experience
572 different oxidative responses to exercise, which provides mechanistic insight into previously
573 observed phenomena such as the sex difference in fatigability. Furthermore, based on these
574 data and others demonstrating no hormonal influences (Mattu *et al.*, 2020), we suggest that
575 there is no rationale for the exclusion of female participants in research investigating
576 cardiopulmonary responses to exercise.

577

578 **Acknowledgements**

579 The authors wish to thank the participants for their time and effort, as well as the technical
580 staff within the Department of Sport, Exercise and Rehabilitation for their support.

581

582 **Funding**

583 This project was supported by a Physiological Society Research Springboard Studentship
584 awarded to MSP, as well as Erasmus+ funding awarded to LB (2020-1-DE01-KA103-
585 005569). PA is supported by the UK Office for Veteran's Affairs (G2-SCH-2022-11-12245).

586

587 **Disclosures**

588 There are no conflicts of interest, financial or otherwise.

589

590 **Author Contributions**

591 MSP, ZW, MB, and PA conceived and designed the research. MSP, LB, EH, LH, and PA
592 performed the experiments. LB, EH, LH, ZW, and PA analysed data. MSP, LB, EH, LH, ZW,
593 MB, and PA interpreted results of the experiments. PA drafted the manuscript. MSP, LB, EH,
594 LH, ZW, MB, and PA edited and revised the manuscript. All authors approved the final
595 version of the manuscript.

596

597

598 **Reference List**

599 Allen DG, Lamb GD & Westerblad H. (2008). Skeletal muscle fatigue: cellular mechanisms. *Physiol*
600 *Rev* **88**, 287-332.

601

602 Ansdell P, Brownstein CG, Škarabot J, Hicks KM, Howatson G, Thomas K, Hunter SK & Goodall S.
603 (2019). Sex differences in fatigability and recovery relative to the intensity–duration
604 relationship. *The Journal of Physiology* **597**, 5577-5595.

605

606 Ansdell P, Škarabot J, Atkinson E, Corden S, Tygart A, Hicks KM, Thomas K, Hunter SK, Howatson G &
607 Goodall S. (2020a). Sex differences in fatigability following exercise normalised to the
608 power–duration relationship. *The Journal of Physiology* **598**, 5717-5737.

609

610 Ansdell P, Thomas K, Hicks KM, Hunter SK, Howatson G & Goodall S. (2020b). Physiological sex
611 differences affect the integrative response to exercise: acute and chronic implications.
612 *Experimental Physiology* **105**, 2007-2021.

613

614 Ansdell P, Thomas K, Howatson G, Hunter S & Goodall S. (2017). Contraction intensity and sex
615 differences in knee-extensor fatigability. *Journal of Electromyography and Kinesiology* **37**,
616 68-74.

- 617
618 Armstrong N & Barker AR. (2009). Oxygen uptake kinetics in children and adolescents: a review.
619 *Pediatr Exerc Sci* **21**, 130-147.
- 620
621 Azevedo RdA, Forot J, Iannetta D, MacInnis MJ, Millet GY & Murias JM. (2021). Slight power output
622 manipulations around the maximal lactate steady state have a similar impact on fatigue in
623 females and males. *Journal of Applied Physiology* **130**, 1879-1892.
- 624
625 Barstow TJ. (2019). Understanding near infrared spectroscopy and its application to skeletal muscle
626 research. *Journal of Applied Physiology* **126**, 1360-1376.
- 627
628 Barstow TJ, Jones AM, Nguyen PH & Casaburi R. (1996). Influence of muscle fiber type and pedal
629 frequency on oxygen uptake kinetics of heavy exercise. *Journal of Applied Physiology* **81**,
630 1642-1650.
- 631
632 Beltrame T, Villar R & Hughson RL. (2017). Sex differences in the oxygen delivery, extraction, and
633 uptake during moderate-walking exercise transition. *Applied Physiology, Nutrition, and*
634 *Metabolism* **42**, 994-1000.
- 635
636 Burnley M, Doust JH, Ball D & Jones AM. (2002). Effects of prior heavy exercise on $\dot{V}O_2$ kinetics
637 during heavy exercise are related to changes in muscle activity. *Journal of Applied Physiology*
638 **93**, 167-174.
- 639
640 Burnley M, Doust JH & Jones AM. (2006). Time required for the restoration of normal heavy exercise
641 $\dot{V}O_2$ kinetics following prior heavy exercise. *Journal of Applied Physiology* **101**, 1320-1327.
- 642
643 Burnley M & Jones AM. (2007). Oxygen uptake kinetics as a determinant of sports performance.
644 *European Journal of Sport Science* **7**, 63-79.
- 645
646 Cano A, Ventura L, Martinez G, Cugusi L, Caria M, Deriu F & Manca A. (2022). Analysis of sex-based
647 differences in energy substrate utilization during moderate-intensity aerobic exercise. *Eur J*
648 *Appl Physiol* **122**, 29-70.
- 649
650 Cardinale DA, Larsen FJ, Schiffer TA, Morales-Alamo D, Ekblom B, Calbet JAL, Holmberg HC & Boushel
651 R. (2018). Superior Intrinsic Mitochondrial Respiration in Women Than in Men. *Front Physiol*
652 **9**, 1133.
- 653
654 DeLorey DS, Kowalchuk JM & Paterson DH. (2003). Relationship between pulmonary O_2 uptake
655 kinetics and muscle deoxygenation during moderate-intensity exercise. *Journal of Applied*
656 *Physiology* **95**, 113-120.
- 657
658 Diaz-Canestro C, Pentz B, Sehgal A & Montero D. (2022). Differences in Cardiac Output and Aerobic
659 Capacity Between Sexes Are Explained by Blood Volume and Oxygen Carrying Capacity. *Front*
660 *Physiol* **13**, 747903.

661
662 do Nascimento Salvador PC, Schäfer L, Grassi B, Guglielmo LGA & Denadai BS. (2019). Changes in
663 VO₂ Kinetics After Elevated Baseline Do Not Necessarily Reflect Alterations in Muscle Force
664 Production in Both Sexes. *Frontiers in Physiology* **10**.

665
666 Ducrocq GP, Hureau TJ, Bøgseth T, Meste O & Blain GM. (2021). Recovery from Fatigue after Cycling
667 Time Trials in Elite Endurance Athletes. *Med Sci Sports Exerc* **53**, 904-917.

668
669 Esbjörnsson-Liljedahl M, Bodin K & Jansson E. (2002). Smaller muscle ATP reduction in women than
670 in men by repeated bouts of sprint exercise. *Journal of Applied Physiology* **93**, 1075-1083.

671
672 Esbjörnsson-Liljedahl M, Sundberg CJ, Norman B & Jansson E. (1999). Metabolic response in type I
673 and type II muscle fibers during a 30-s cycle sprint in men and women. *Journal of Applied
674 Physiology* **87**, 1326-1332.

675
676 Faude O, Kindermann W & Meyer T. (2009). Lactate Threshold Concepts. *Sports Medicine* **39**, 469-
677 490.

678
679 Furrer R, Hawley JA & Handschin C. (2023). The molecular athlete: exercise physiology from
680 mechanisms to medals. *Physiological Reviews* **103**, 1693-1787.

681
682 Gloeckl R, Zwick RH, Furlinger U, Jarosch I, Schneeberger T, Leitl D, Koczulla AR, Vonbank K, Alexiou
683 C, Vogiatzis I & Spruit MA. (2022). Prescribing and adjusting exercise training in chronic
684 respiratory diseases – Expert-based practical recommendations. *Pulmonology*.

685
686 Goulding RP & Marwood S. (2023). Interaction of Factors Determining Critical Power. *Sports
687 Medicine* **53**, 595-613.

688
689 Goulding RP, Rossiter HB, Marwood S & Ferguson C. (2021). Bioenergetic Mechanisms Linking V'O₂
690 Kinetics and Exercise Tolerance. *Exercise and Sport Sciences Reviews* **49**.

691
692 Grassi B. (2001). Regulation of Oxygen Consumption at Exercise Onset: Is It Really Controversial?
693 *Exercise and Sport Sciences Reviews* **29**, 134-138.

694
695 Grassi B, Poole DC, Richardson RS, Knight DR, Erickson BK & Wagner PD. (1996). Muscle O₂ uptake
696 kinetics in humans: implications for metabolic control. *Journal of Applied Physiology* **80**, 988-
697 998.

698
699 Grassi B, Porcelli S, Salvadego D & Zoladz JA. (2011). Slow VO₂ kinetics during moderate-intensity
700 exercise as markers of lower metabolic stability and lower exercise tolerance. *Eur J Appl
701 Physiol* **111**, 345-355.

702
703 Grassi B, Rossiter HB & Zoladz JA. (2015). Skeletal muscle fatigue and decreased efficiency: two sides
704 of the same coin? *Exerc Sport Sci Rev* **43**, 75-83.

705
706 Harmer AR, Ruell PA, Hunter SK, McKenna MJ, Thom JM, Chisholm DJ & Flack JR. (2014). Effects of
707 type 1 diabetes, sprint training and sex on skeletal muscle sarcoplasmic reticulum Ca²⁺
708 uptake and Ca²⁺-ATPase activity. *The Journal of Physiology* **592**, 523-535.

709
710 Harms CA, McClaran SR, Nickele GA, Pegelow DF, Nelson WB & Dempsey JA. (1998). Exercise-
711 induced arterial hypoxaemia in healthy young women. *J Physiol* **507 (Pt 2)**, 619-628.

712
713 Hughson RL, Tschakovsky ME & Houston ME. (2001). Regulation of Oxygen Consumption at the
714 Onset of Exercise. *Exercise and Sport Sciences Reviews* **29**, 129-133.

715
716 Hunter SK. (2014). Sex differences in human fatigability: mechanisms and insight to physiological
717 responses. *Acta Physiologica* **210**, 768-789.

718
719 James JJ, Klevenow EA, Atkinson MA, Vosters EE, Bueckers EP, Quinn ME, Kindy SL, Mason AP,
720 Nelson SK, Rainwater KAH, Taylor PV, Zippel EP & Hunter SK. (2023). Underrepresentation of
721 women in exercise science and physiology research is associated with authorship gender.
722 *Journal of Applied Physiology* **135**, 932-942.

723
724 Koppo K, Bouckaert J & Jones AM. (2004). Effects of Training Status and Exercise Intensity on Phase II
725 $\dot{V}O_2$ Kinetics. *Medicine & Science in Sports & Exercise* **36**, 225-232.

726
727 Korzeniewski B & Rossiter HB. (2015). Each-step activation of oxidative phosphorylation is necessary
728 to explain muscle metabolic kinetic responses to exercise and recovery in humans. *The*
729 *Journal of Physiology* **593**, 5255-5268.

730
731 Mattu AT, Iannetta D, MacInnis MJ, Doyle-Baker PK & Murias JM. (2020). Menstrual and oral
732 contraceptive cycle phases do not affect submaximal and maximal exercise responses.
733 *Scandinavian Journal of Medicine & Science in Sports* **30**, 472-484.

734
735 McDougall RM, Tripp TR, Frankish BP, Doyle-Baker PK, Lun V, Wiley JP, Aboodarda SJ & MacInnis MJ.
736 (2023). The influence of skeletal muscle mitochondria and sex on critical torque and
737 performance fatigability in humans. *The Journal of Physiology* **601**, 5295-5316.

738
739 Miotto PM, McGlory C, Holloway TM, Phillips SM & Holloway GP. (2018). Sex differences in
740 mitochondrial respiratory function in human skeletal muscle. *American Journal of*
741 *Physiology-Regulatory, Integrative and Comparative Physiology* **314**, R909-R915.

742
743 Murias JM, Keir DA, Spencer MD & Paterson DH. (2013). Sex-related differences in muscle
744 deoxygenation during ramp incremental exercise. *Respiratory Physiology & Neurobiology*
745 **189**, 530-536.

746
747 Murias JM, Kowalchuk JM & Paterson DH. (2010). Speeding of $\dot{V}O_2$ kinetics with endurance training
748 in old and young men is associated with improved matching of local O₂ delivery to muscle
749 O₂ utilization. *Journal of Applied Physiology* **108**, 913-922.

750
751 Murphy WG. (2014). The sex difference in haemoglobin levels in adults — Mechanisms, causes, and
752 consequences. *Blood Reviews* **28**, 41-47.

753
754 Oosthuysen T, Strauss JA & Hackney AC. (2023). Understanding the female athlete: molecular
755 mechanisms underpinning menstrual phase differences in exercise metabolism. *Eur J Appl*
756 *Physiol* **123**, 423-450.

757
758 Parker BA, Smithmyer SL, Pelberg JA, Mishkin AD, Herr MD & Proctor DN. (2007). Sex differences in
759 leg vasodilation during graded knee extensor exercise in young adults. *J Appl Physiol (1985)*
760 **103**, 1583-1591.

761
762 Poole DC, Barstow TJ, Gaesser GA, Willis WT & Whipp BJ. (1994). VO₂ slow component: physiological
763 and functional significance. *Med Sci Sports Exerc* **26**, 1354-1358.

764
765 Poole DC & Jones AM. (2012). Oxygen Uptake Kinetics. *Compr Physiol* **2**, 933-996.

766
767 Porcelli S, Marzorati M, Morandi L & Grassi B. (2016). Home-based aerobic exercise training
768 improves skeletal muscle oxidative metabolism in patients with metabolic myopathies.
769 *Journal of Applied Physiology* **121**, 699-708.

770
771 Pringle JSM, Doust JH, Carter H, Tolfrey K, Campbell IT & Jones AM. (2003). Oxygen uptake kinetics
772 during moderate, heavy and severe intensity 'submaximal' exercise in humans: the influence
773 of muscle fibre type and capillarisation. *European Journal of Applied Physiology* **89**, 289-300.

774
775 Roepstorff C, Thiele M, Hillig T, Pilegaard H, Richter EA, Wojtaszewski JF & Kiens B. (2006). Higher
776 skeletal muscle alpha2AMPK activation and lower energy charge and fat oxidation in men
777 than in women during submaximal exercise. *J Physiol* **574**, 125-138.

778
779 Rossiter HB, Ward SA, Howe FA, Kowalchuk JM, Griffiths JR & Whipp BJ. (2002). Dynamics of
780 intramuscular 31P-MRS Pi peak splitting and the slow components of PCr and O₂ uptake
781 during exercise. *Journal of Applied Physiology* **93**, 2059-2069.

782
783 Rossiter HB, Ward SA, Kowalchuk JM, Howe FA, Griffiths JR & Whipp BJ. (2001). Effects of prior
784 exercise on oxygen uptake and phosphocreatine kinetics during high-intensity knee-
785 extension exercise in humans. *J Physiol* **537**, 291-303.

786
787 Russ DW, Lanza IR, Rothman D & Kent-Braun JA. (2005). Sex differences in glycolysis during brief,
788 intense isometric contractions. *Muscle & Nerve* **32**, 647-655.

789
790 Ryan TE, Erickson ML, Brizendine JT, Young H-J & McCully KK. (2012). Noninvasive evaluation of
791 skeletal muscle mitochondrial capacity with near-infrared spectroscopy: correcting for blood
792 volume changes. *Journal of Applied Physiology* **113**, 175-183.

793

794 Schiaffino S & Reggiani C. (2011). Fiber types in mammalian skeletal muscles. *Physiol Rev* **91**, 1447-
795 1531.

796

797 Simoneau JA & Bouchard C. (1989). Human variation in skeletal muscle fiber-type proportion and
798 enzyme activities. *American Journal of Physiology-Endocrinology and Metabolism* **257**, E567-
799 E572.

800

801 Staron RS, Hagerman FC, Hikida RS, Murray TF, Hostler DP, Crill MT, Ragg KE & Toma K. (2000). Fiber
802 type composition of the vastus lateralis muscle of young men and women. *J Histochem*
803 *Cytochem* **48**, 623-629.

804

805 Temesi J, Mattioni Maturana F, Peyrard A, Piucco T, Murias JM & Millet GY. (2017). The relationship
806 between oxygen uptake kinetics and neuromuscular fatigue in high-intensity cycling
807 exercise. *European Journal of Applied Physiology* **117**, 969-978.

808

809 Tschakovsky ME & Hughson RL. (1999). Interaction of factors determining oxygen uptake at the
810 onset of exercise. *Journal of Applied Physiology* **86**, 1101-1113.

811

812 Willcocks RJ, Williams CA, Barker AR, Fulford J & Armstrong N. (2010). Age- and sex-related
813 differences in muscle phosphocreatine and oxygenation kinetics during high-intensity
814 exercise in adolescents and adults. *NMR in Biomedicine* **23**, 569-577.

815

816

817

818 **Figure Legends**

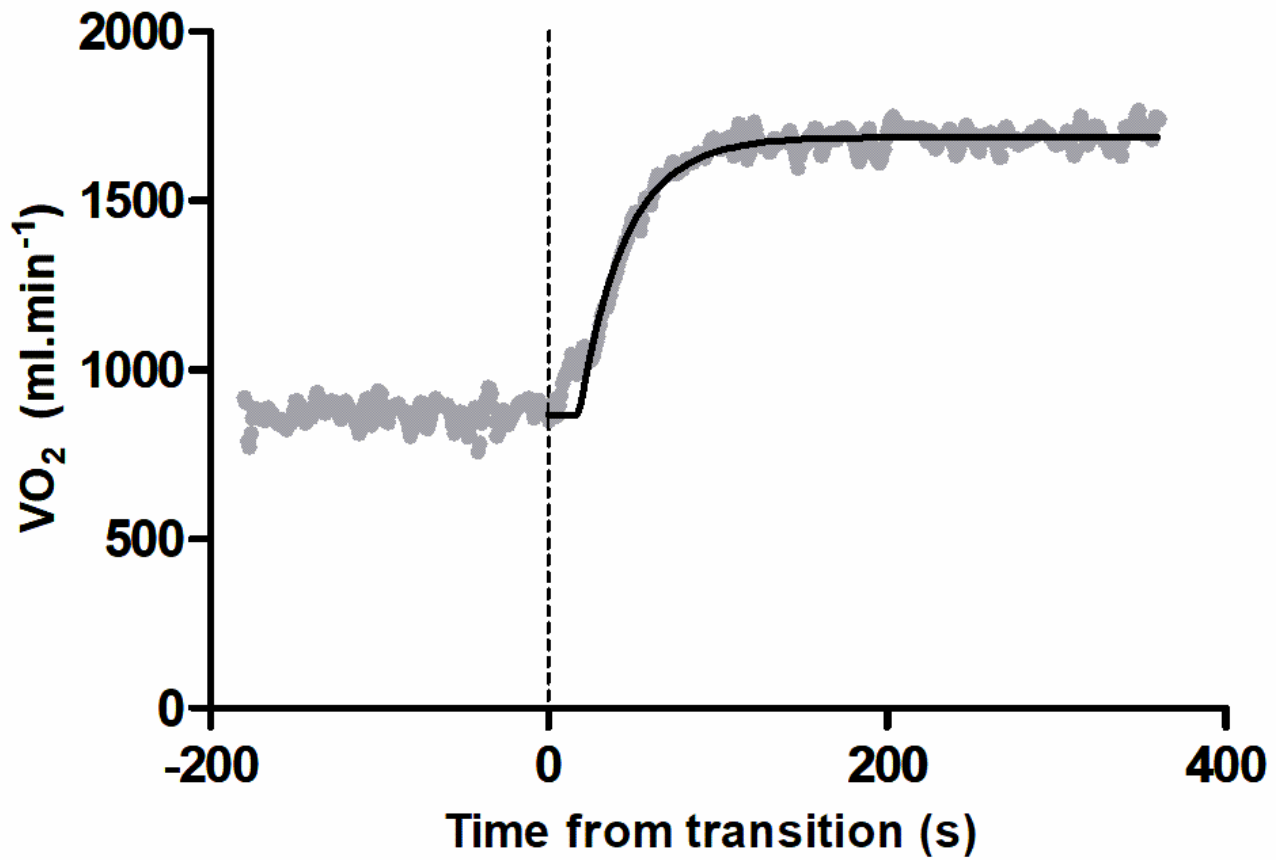
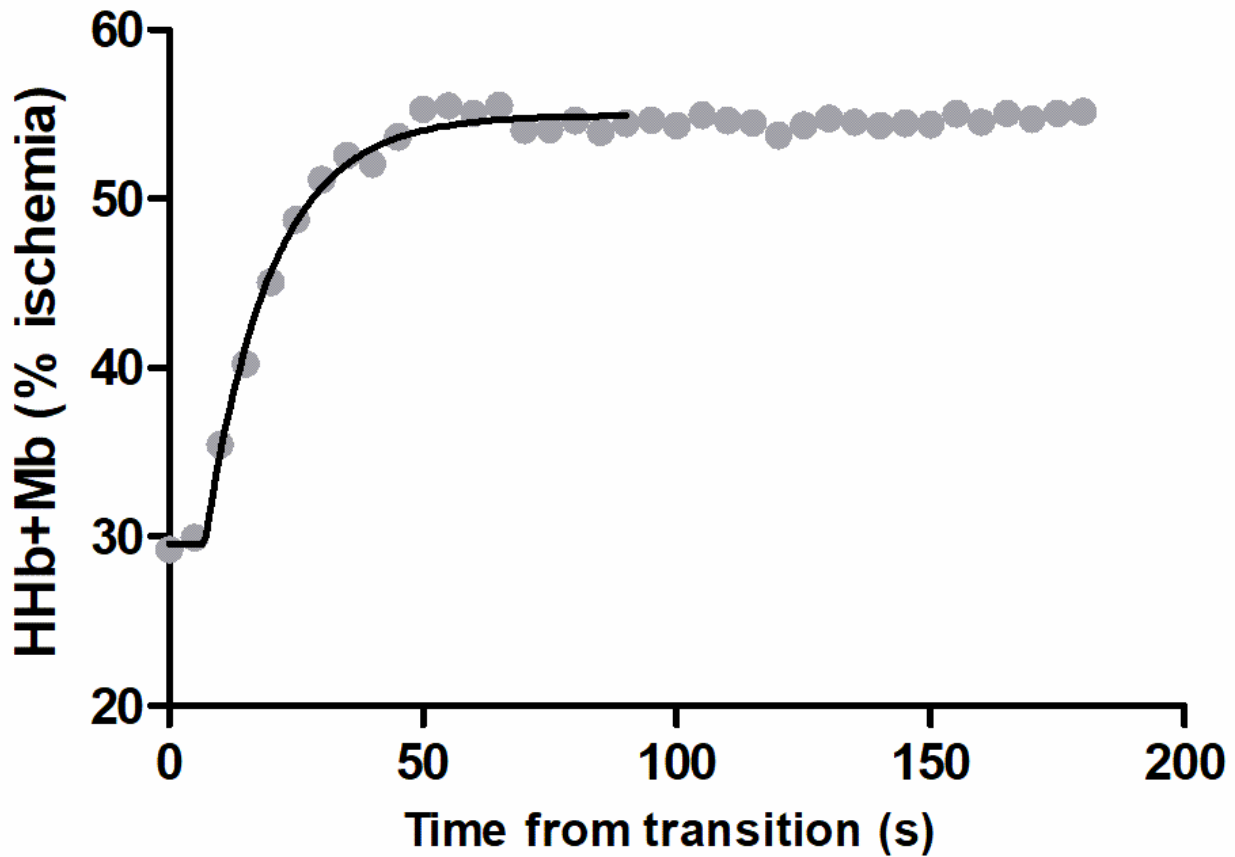
819 Figure 1: Visualisation of the monoexponential curve fitting procedures for a representative
820 participant's data in the moderate intensity domain. Panel A describes the $\dot{V}O_2$ data (1 Hz)
821 and Panel B describes the HHb+Mb data (0.2 Hz).

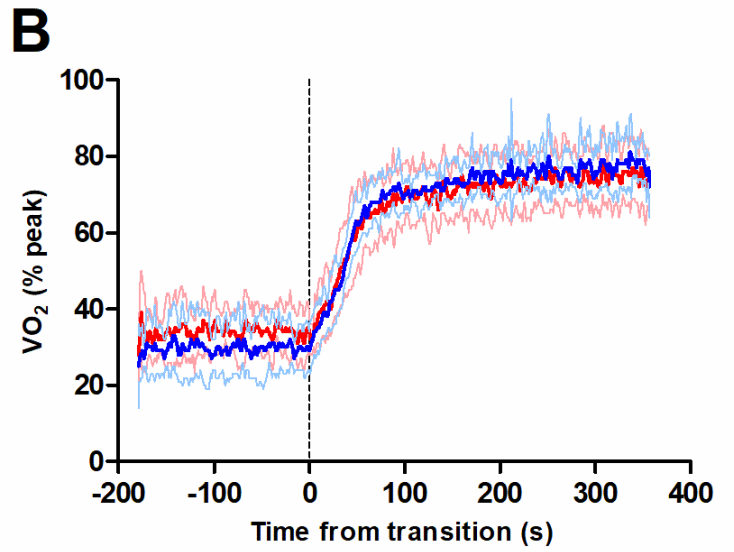
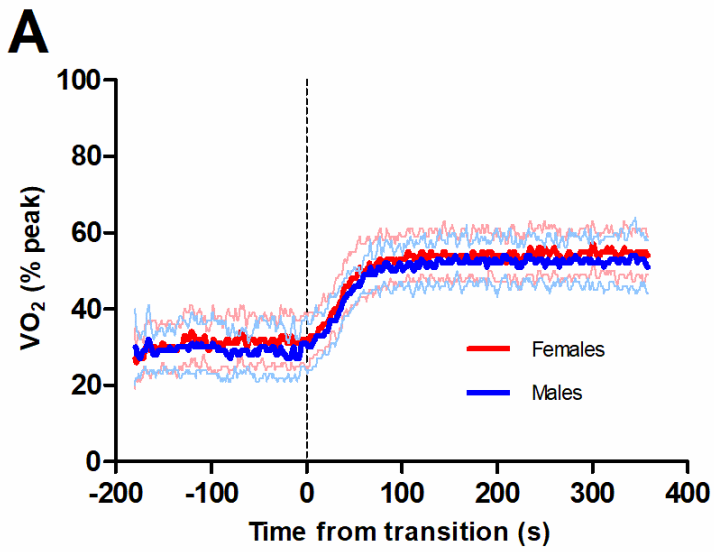
822

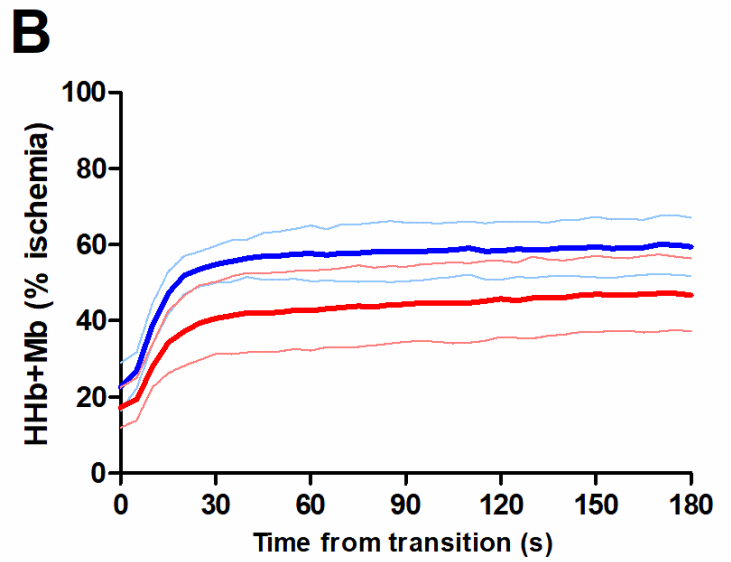
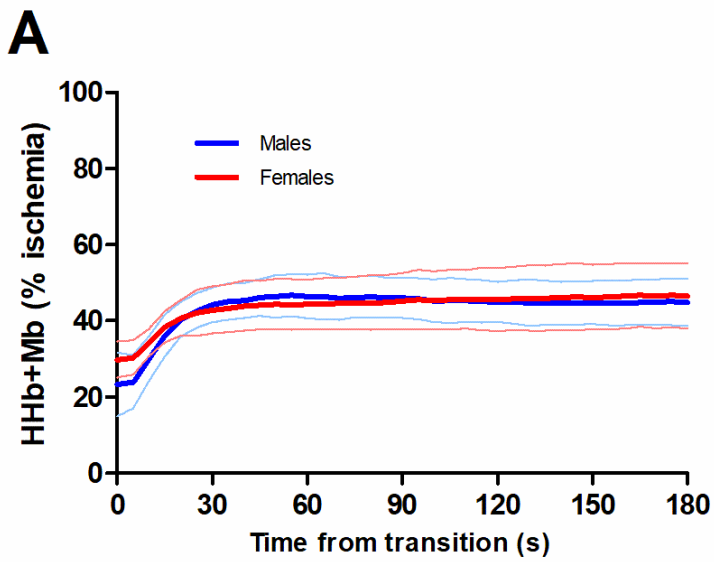
823 Figure 2: Group mean $\dot{V}O_2$ data from males (blue, n=8) and females (red, n=8) during
824 moderate (Panel A) and heavy (Panel B) intensity transitions. The bold lines represent group
825 means and the thin lines represent standard deviation.

826

827 Figure 3: Group mean HHb+Mb data from males (blue, n=8) and females (red, n=7) during
828 moderate (Panel A) and heavy (Panel B) intensity transitions. The bold lines represent group
829 means and the thin lines represent standard deviation.

A**B**





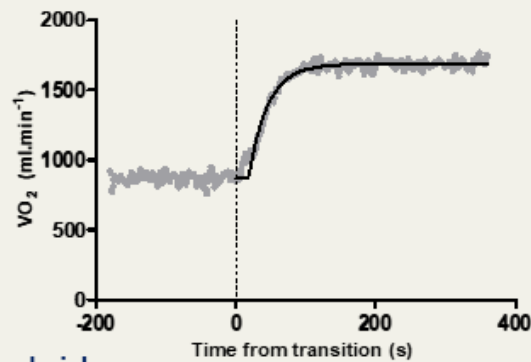
No sex differences in oxygen uptake or extraction kinetics in the moderate or heavy exercise intensity domains

METHODS

8 males, 8 females (27 ± 5 years)

Moderate (80% lactate threshold) and heavy (30% Δ) intensity transitions

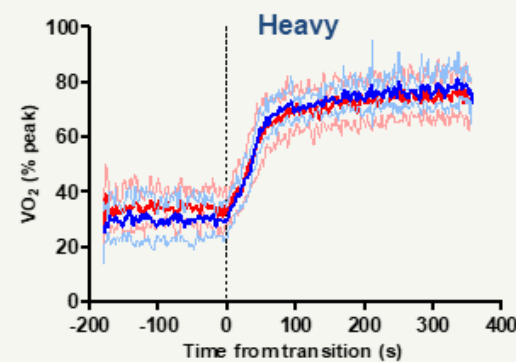
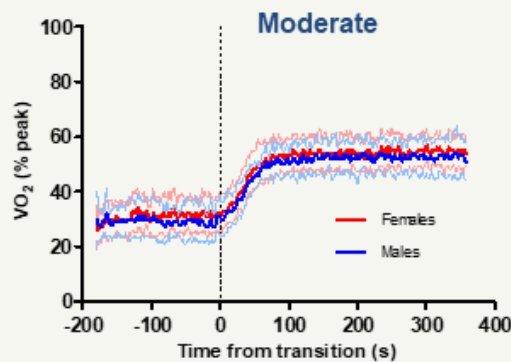
Modelling of VO_2 and HHb+Mb phase II kinetics & slow components



[doi here](#)

RESULTS

No sex differences in the speed or relative amplitude of the phase II or slow component kinetics.



CONCLUSION The dynamic response of oxidative metabolism does not differ between sexes in the moderate and heavy intensity domains.

Table 1: Anthropometric data and outcome variables from incremental exercise testing.

	Males (n = 8)	Females (n = 8)	P value	Cohen's d
Age (years)	27 ± 3	27 ± 7	0.734	0.117
Stature (cm)	182 ± 5	163 ± 4	< 0.001	1.383
Body Mass (kg)	75.3 ± 10.2	61.8 ± 5.9	0.006	0.501
$\dot{V}O_{2peak}$ (L.min ⁻¹)	3.47 ± 0.58	2.50 ± 4.42	0.002	0.627
Relative $\dot{V}O_{2peak}$ (ml.kg ⁻¹ .min ⁻¹)	46.2 ± 6.6	40.5 ± 6.7	0.111	0.326
Pmax (W)	328 ± 54	236 ± 43	0.002	0.646
Pmax (W.kg ⁻¹)	4.4 ± 0.8	3.8 ± 0.5	0.118	0.253
Power at LT (W)	153 ± 25	119 ± 29	0.023	0.528
LT (% Pmax)	47 ± 6	50 ± 7	0.373	0.183
LT (W.kg ⁻¹)	2.0 ± 0.2	1.9 ± 0.4	0.440	0.127
80% LT (W)	123 ± 20	95 ± 23	0.023	0.528
80% LT (W.kg ⁻¹)	1.6 ± 0.2	1.5 ± 0.3	0.440	0.110
30% Δ (W)	206 ± 30	154 ± 32	0.005	0.653
30% Δ (W.kg ⁻¹)	2.7 ± 0.4	2.5 ± 0.4	0.198	0.213

LT: lactate threshold, Pmax: Maximal ramp test power output, $\dot{V}O_{2peak}$: Maximal rate of oxygen consumption.

Table 2: Data from the monoexponential modelling of $\dot{V}O_2$ kinetics during moderate and heavy intensity transitions.

	Males (n = 8)	Females (n = 8)	P value	Cohen's d
Moderate Intensity Domain				
TD (s)	16.6 ± 4.1	17.9 ± 4.9	0.575	0.121
Baseline $\dot{V}O_2$ (L.min ⁻¹)	0.97 ± 0.05	0.77 ± 0.09	< 0.001	1.516
Amplitude (L.min ⁻¹)	0.83 ± 0.19	0.60 ± 0.18	0.024	0.477
Amplitude (ml.kg ⁻¹ .min ⁻¹)	11.0 ± 1.6	9.7 ± 2.6	0.230	0.323
Amplitude (% $\dot{V}O_{2peak}$)	24 ± 3	24 ± 5	0.949	0.018
$\dot{V}O_2$ gain (ml.min ⁻¹ .W ⁻¹)	10.2 ± 0.7	10.5 ± 1.1	0.587	0.130
τ (s)	27.9 ± 7.5	24.8 ± 6.6	0.385	0.160
r^2	0.965 ± 0.032	0.947 ± 0.034		
Heavy Intensity Domain				
TD (s)	15.0 ± 4.8	14.5 ± 4.5	0.858	0.033
Baseline $\dot{V}O_2$ (L.min ⁻¹)	1.00 ± 0.06	0.85 ± 0.11	0.005	0.894
Amplitude (L.min ⁻¹)	1.50 ± 0.38	0.96 ± 0.25	0.005	0.541
Amplitude (ml.kg ⁻¹ .min ⁻¹)	19.9 ± 4.7	15.6 ± 3.7	0.060	0.351
Amplitude (% $\dot{V}O_{2peak}$)	43 ± 5	38 ± 7	0.179	0.355
$\dot{V}O_2$ gain (ml.min ⁻¹ .W ⁻¹)	9.4 ± 0.8	9.3 ± 0.9	0.766	0.059
τ (s)	28.8 ± 7.9	27.2 ± 4.4	0.633	0.075
r^2	0.961 ± 0.031	0.927 ± 0.030		
SC Amplitude (L.min ⁻¹)	0.38 ± 0.16	0.28 ± 0.12	0.158	0.250
SC Amplitude (% $\dot{V}O_{2peak}$)	11.9 ± 6.9	10.8 ± 3.2	0.686	0.061
SC Amplitude (% end exercise $\dot{V}O_2$)	13.6 ± 6.5	12.9 ± 3.9	0.822	0.035

SC: slow component, τ : time constant, TD: time delay, $\dot{V}O_2$: rate of oxygen consumption

Table 3: Data from the monoexponential modelling of deoxyhaemoglobin kinetics during moderate and heavy intensity transitions.

	Males (n = 8)	Females (n = 7)	P value	Cohen's d
Moderate Intensity Domain				
TD (s)	8.7 ± 1.5	9.6 ± 2.6	0.447	0.212
Baseline HHb+Mb (% ischemia)	23 ± 8	29 ± 7	0.186	0.251
Amplitude (% ischemia)	21 ± 7	17 ± 5	0.225	0.219
τ (s)	8.1 ± 2.8	10.0 ± 3.8	0.274	0.260
r ²	0.974 ± 0.022	0.956 ± 0.037		
Heavy Intensity Domain				
TD (s)	6.2 ± 2.8	7.0 ± 2.3	0.582	0.101
Baseline HHb+Mb (% ischemia)	23 ± 6	17 ± 7	0.146	0.332
Amplitude (% ischemia)	36 ± 9	30 ± 8	0.193	0.252
τ (s)	11.3 ± 3.7	12.2 ± 4.0	0.665	0.090
r ²	0.983 ± 0.011	0.979 ± 0.023		
SC Amplitude (% ischemia)	14 ± 5	16 ± 4	0.432	0.142

HHb+Mb: deoxyhaemoglobin and myoglobin, SC: slow component, τ: time constant, TD: time delay consumption

Table 4: Data from near-infrared spectroscopy before and during the moderate and heavy intensity exercise transitions in males (n = 8) and females (n = 7).

		Moderate Intensity		Heavy Intensity		
		Unloaded Pedalling	End Stage	Unloaded Pedalling	90 secs	End Stage
HbO₂ + MbO₂ (%ischemia)	Male	57 ± 6	53 ± 7	68 ± 11*	54 ± 10* [#]	47 ± 10 [#]
	Female	41 ± 10	44 ± 9	51 ± 7	41 ± 4 [#]	53 ± 8
TOI (%)	Male	69 ± 3	61 ± 5 [#]	71 ± 4	57 ± 6 [#]	50 ± 8 [#] [§]
	Female	75 ± 3*	71 ± 6* [#]	78 ± 4*	70 ± 8* [#]	69 ± 9* [#]
TOI (% ischemia)	Male	72 ± 4	60 ± 7 [#]	76 ± 6	53 ± 9 [#]	39 ± 10 [#] [§]
	Female	67 ± 9	53 ± 10 [#]	77 ± 6	52 ± 9 [#]	51 ± 17 [#]

HbO₂+MbO₂: oxygenated haemoglobin and myoglobin; TOI: tissue oxygenation index; * = greater than the opposite sex (p < 0.05); [#] = lower than unloaded pedalling (p < 0.05); [§] = lower than 90 secs.