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# 1 **Theobroma Genus: Exploring the Therapeutic Potential of *T. grandiflorum* and *T. bicolor* in biomedicine**

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## 11 **Abstract**

12 The Amazon rainforest hosts a plethora of fruit-bearing plants, yet many remain untapped for commercial  
13 purposes. Among these, *Theobroma* genus stands out for its unique characteristics deeply rooted in culinary and  
14 traditional medicinal practices, significantly contributing to Amazonian biodiversity and cultural heritage.  
15 Particularly, *T. cacao*, the most renowned species, exhibits versatile applications owing to its health benefits, with  
16 distinct groups influencing cocoa quality. Similarly, *T. bicolor*, thriving in humid regions, has undergone  
17 domestication to yield pulp and seeds valuable in food and cosmetic industries. Meanwhile, *T. grandiflorum*,  
18 found across tropical regions of Central and South America, presents unique sensory profiles and fruit  
19 characteristics, making it a significant player in Amazonian agriculture. This review primarily aims to offer  
20 insights into the therapeutic potential of *T. grandiflorum* and *T. bicolor*, with comparisons to *T. cacao*, revealing  
21 a notable increase in publications concerning the physico-chemical and biological properties of these species in  
22 recent years. Specifically, the review examines their chemical composition, bioactive compounds, and  
23 methodologies for determination, with a focus on biological evaluations encompassing enzymatic, cellular, and  
24 animal tests, thereby shedding light on the medicinal properties of these species. Finally, future research  
25 perspectives, emphasising the utilisation of waste biomass and further exploration of these invaluable Amazonian  
26 resources, have been discussed.

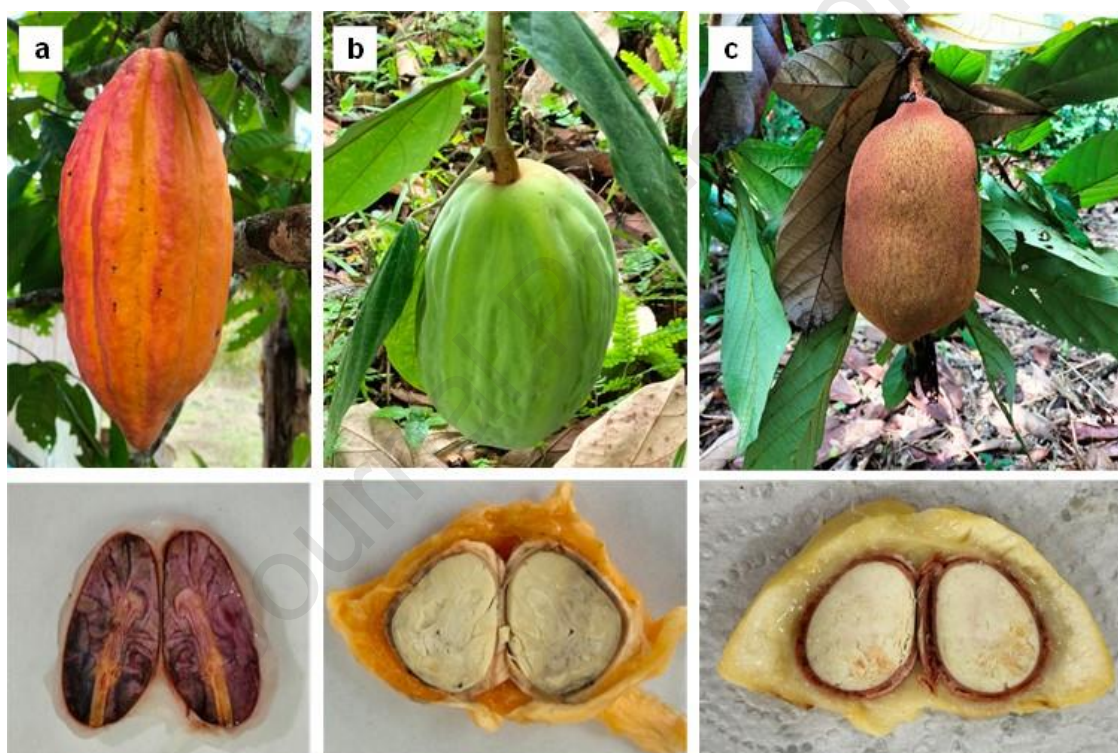
27 **Keywords:** Amazonian fruits, *Theobroma* genus, phenolic compounds, antioxidant capacity, therapeutic  
28 applications.

## 29 Introduction

30 Within the diverse fruit production of the Amazon, numerous plants with potential for agro-industrial exploitation  
31 have been identified; however, they have not undergone widespread commercial development. Examples of such  
32 species include *E. oleracea*, *M. flexuosa*, *A. aculeatum*, *B. gasipaes*, *P. cupana*, *U. tomentosa*, etc. (Amorim et al.,  
33 2024; Souza et al., 2023). Among these promising species, the genus *Theobroma* is recognised for its distinctive  
34 characteristics, which are recognised and valued in culinary and traditional medicinal practices. Indeed, its species  
35 produce fruits with unique attributes and applications, playing a significant role in both the biodiversity and  
36 cultural heritage of the Amazon. Additionally, they hold substantial economic potential, particularly the main  
37 representative species, *Theobroma cacao*, due to its health benefits (González-Orozco et al., 2020), accounting 22  
38 species native to the tropical forests of Central and South America (Huda-Shakirah et al., 2022). Certain species  
39 within this genus, such as *Theobroma grandiflorum* and *Theobroma bicolor*, are recognised as non-timber forest  
40 species that predominantly thrive in humid soils, native from the Amazon basin, sharing similarities with *T. cacao*  
41 (Abdullah et al., 2020). All these three species produce fruits with unique characteristics that make them special  
42 and appreciated in the gastronomy and traditional medicine of Amazonian culture. **Figure 1** shows the anatomical  
43 features of fruit and seed of *T. cacao*, *T. bicolor*, and *T. grandiflorum*. Particularly, *T. cacao* is native to South  
44 America and has successfully adapted to regions between 10° south and north of the equator worldwide. It is the  
45 smallest plant among these three species, and can grow up to 8 meters in height, with a trunk diameter of about  
46 30 cm, featuring lanceolate leaves and white or pink flowers. Additionally, it is characterised by an elongated and  
47 winged fruit with a hard and rough shell, and it produces large, elongated seeds, wrapped in white and sweet pulp  
48 but not very abundant (**Figure 1a**). However, the bean is highly profitable due to its diverse applications in various  
49 industries, thanks to its fat profile, flavour, aroma, and antioxidant capacity derived from bioactive compounds  
50 such as polyphenols, which are linked to potential health benefits (Barrios-Rodríguez et al., 2022). There are three  
51 varieties of *T. cacao* (Criollo, Amazonian Forastero, and Trinitario) associated with specific fermentation  
52 processes, influencing the quality of cocoa derivatives: powder, liquor, and butter (Fanning et al., 2023).

53 Then, *T. bicolor* exhibits growth or cultivation proclivity within humid regions, not surpassing an elevation of  
54 1000 meters above sea level, encompassing territories within Bolivia, Brazil, Ecuador, Colombia, and Peru, with  
55 some cultivation present in Mexico (Ponce-Sánchez et al., 2021). This plant has undergone domestication and  
56 technological refinement to produce pulp and seeds, presenting applications in the food and cosmetic industries  
57 (Mar et al., 2024). In its natural habitat, the tree can attain heights around 10 m, yet under cultivation, it undergoes  
58 pruning measures to sustain more manageable dimensions. The mature plants of *T. bicolor* can yield up to 40

59 fruits per harvest, occurring in February-March and September-October, necessitating daily harvesting to prevent  
 60 fruit rot. The flowers are small and white; however, they are converted in the largest fruit within the genus  
 61 *Theobroma*, characterised by an oval morphology, dimensions ranging from 25 to 35 cm in length, 12 to 15 cm in  
 62 diameter, and a weight of up to ~3 kg. The fruit peel exhibits a woody and robust composition that falls to the  
 63 ground when it reaches its optimum ripeness, it possesses a pod thickness of 12 mm, and features fissures (**Figure**  
 64 **1b**). Its colouration transitions to a yellow hue upon reaching maturity. The pulp presents as whitish or yellowish,  
 65 with a strong odour and a sweet and sour taste. It is rich in proteins, both insoluble and soluble fibres, and fats,  
 66 while its seeds produce butter with a fat content like cocoa, suggesting its potential use as a substitute or blend  
 67 with cocoa butter, particularly in the chocolate industry (González et al., 2016).



68  
 69 **Figure 1.** Images of fruit and seed of *T. cacao* (a), *T. bicolor* (b), and *T. grandiflorum* (c) taken in North-West  
 70 area of the Amazon close to Florencia, Caquetá - Colombia.

71 Finally, *Theobroma grandiflorum* is grown in tropical regions of Central America (Costa Rica), South America  
 72 (Ecuador, Peru, Venezuela, Colombia, Guyana, Suriname), and some Caribbean islands (Martinique, Trinidad and  
 73 Tobago) (Duarte & Paull, 2015). This species has contributed to Amazonian agriculture, especially in Peru and  
 74 Brazil, where most of the research mentioned in this document has been conducted. *T. grandiflorum* exhibits  
 75 various varieties with differences in sensory profile, peel, and seed size (Febrianto & Zhu, 2022). It differs  
 76 anatomically from *T. cacao* and *T. bicolor*, being notably larger, reaching heights of up to 15 m with a trunk  
 77 diameter of up to 70 cm, and featuring a distinct internal structure and design of the kernel. Regarding its flowers,

78 they are the largest ones of the three species, presenting a white or pink colour. The fruit is an oval or elongated  
79 berry with a tough outer shell covered in hairs that become green when gently rubbed, signalling ripeness (**Figure**  
80 **1c**). The pulp yield is approximately 40%, with a colour ranging from white to cream or creamy yellow and a  
81 sweet and sour taste. The seeds, located within the pulp, account for ~18% of the fresh fruit weight. Butter and  
82 liquor extracted from the seeds have a lipid content that is appealing to the cosmetic industry, while the highly  
83 aromatic pulp is of interest to the food industry (Pereira et al., 2018).

84 In this review, we proposed an overview on the therapeutic potential of *Theobroma grandiflorum* and *Theobroma*  
85 *bicolor*, by including studies of *T. cacao* (although reviews on this species are present in literature) to allows  
86 comparisons with the other two species. Specifically, this review presents information gathered from several  
87 documents in scientific literature. By a simple key-word search on ISI Web of Science database of topic terms  
88 which includes the scientific names of the three species, alongside terms such as "proximal composition,"  
89 "biomedicine," "therapeutic," "antioxidant capacity," "biological test", it is possible to appreciate that the number  
90 of publications on the physico-chemical and biological properties of these species increased about 57% since  
91 2020. In this information collection, works focused on the characterisation of volatile compounds of the three  
92 species are excluded, as our focus primarily lies on their chemical composition and bioactive compounds profile,  
93 with a focus on the main methodologies for their determination, with the undertaking of a unit conversion task for  
94 both the proximal characterisation and antioxidant profile of the three species to facilitate their comparison.  
95 Furthermore, the review describes the biological evaluation of the *Theobroma grandiflorum* and *Theobroma*  
96 *bicolor*, including enzymatic, cellular, and animal tests. Finally, the review provides in the last section a visionary  
97 perspective on possible areas for further exploration of the analysed species, with the aim of leveraging these raw  
98 materials, including the use of their waste biomass.

#### 99 **Chemical constituents of *T. grandiflorum*, *T. cacao* and *T. bicolor***

100 Literature reports several works on the investigation of the chemical composition of *T. grandiflorum*, *T. cacao* and  
101 *T. bicolor*, reporting mainly records of their proximal composition as well as information on proteomic analysis.  
102 Additionally, different studies are focused on the antioxidant activity of polyphenols and flavonoids, contained in  
103 these *Theobroma* species. Interestingly, these antioxidants have been linked to various health benefits, including  
104 digestive health, anti-inflammatory properties and potential anticancer effects. Thus, studying the chemical  
105 composition of these bioactive compounds cab allow to design and manufacture products that can be used for  
106 biomedical applications, contributing to our well-being.

107 **Table 1** shows the average values of the proximal composition of the three *Theobroma* species. Concerning *T.*  
 108 *cacao*, the reviewed articles do not assess moisture in the pulp due to its close adherence to the seed and limited  
 109 fleshy characteristics, while when evaluating moisture content in the pulp of other species, it was found that *T.*  
 110 *bicolor* exhibits a higher content compared to *T. grandiflorum*. Moreover, the seeds, constituting the organ with  
 111 the highest economic value in these species owing to their fat content, exhibit values within a range of 52 to 61%.

112 **Table 1.** Proximal composition of *Theobroma* species. Some original values have been converted to percentages  
 113 to facilitate comparison.

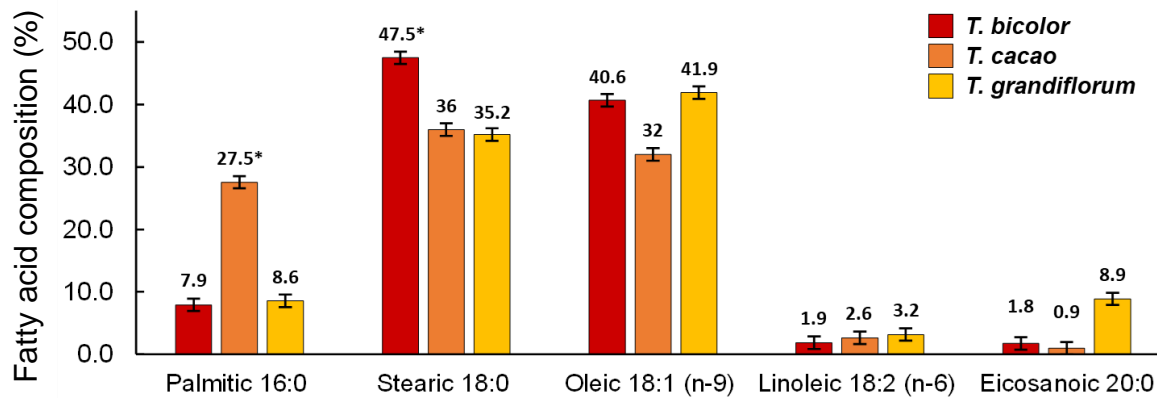
Sample	Moisture (%)	Ashes (%)	Lipids (%)	Protein (%)	Carbs (%)	Fibre (%)	References
<i>Theobroma bicolor</i>							
Pulp fresh	92.1	0.7	0.29	1.3	4.8	0.8	(Sotelo & Alvarez, 1991)
Pulp + seeds	35.6	0.8	33	13.3	5.1	12.2	(Torres et al., 2002)
Seeds fresh	52.3	1.7	17	11.4	8.8	8.8	(Sotelo & Alvarez, 1991)
Seeds fermented-dried	4.4	3.5	38-41.3	22-23.8			(Sotelo & Alvarez, 1991), (Jee, 1984), (Vázquez-Ovando et al., 2015)
Shell	50.6	1.1	0.37	3.25	12.6	32	(Sotelo & Alvarez, 1991)
<i>Theobroma cacao</i>							
Seeds fresh	52.5-61	1.4-2.1	19.6-23.9	6.3-7.9	3.6-10.5	3.1-3.2	(Sotelo & Alvarez, 1991)
Seeds fermented-dried	5.41	5.23	46.1	13.6			(Vázquez-Ovando et al., 2015)
Liquor	26.4	3	54	10.5		2.7	(de Oliveira & Genovese, 2013)
Shell	90.1-94.7	0.5-1	0.01-0.1	0.4-1	2.8-4.4	1.5-3.4	(Sotelo & Alvarez, 1991)
<i>Theobroma grandiflorum</i>							
Pulp fresh or frozen	83-89.7	0.5-3.3	0.4-2.1	0.7-2.5	5.6-10.6		(Pugliese et al., 2013), (Zagmignan et al., 2023)
Seeds fresh	52.7-54.9	1.5	21.7	4.3	15.8		(Pugliese et al., 2013), (De Souza Schmidt Goncalves et al., 2010)
Liquor	22.6	7.3	52	15.2		3.2	(de Oliveira & Genovese, 2013)

114

115 To be noted, ash contents were higher in the seeds than in the pulps of *Theobroma* species. Indeed, the  
 116 endosperm's mineral storage supports vital chlorophyll formation for cotyledon-initiated photosynthesis,  
 117 impacting overall ash content (Sciammaro et al., 2016), which, upon incineration, leads to an elevation of this  
 118 value. However, in the case of *T. grandiflorum*, observed ranges include slightly higher ash values in the pulp.

119 Moreover, a study employing inductively coupled plasma-atomic emission spectroscopy (de Oliveira &  
120 Genovese, 2013) analysed the mineral composition of *T. cocoa* and *T. grandiflorum* liquors, revealing the  
121 following results: nitrogen predominates in both species at 23.3 and 25.9 g/kg dry weight (DW), with potassium  
122 (K) following at 7.5 and 6.8 g/kg DW, phosphorus (P) at 2 and 3.6 g/kg DW, and calcium (Ca) at 0.3 and 0.5  
123 g/kg DW respectively.

124 Regarding the presence of lipids in fresh seeds, all the reviewed documents adhered to the methodology proposed  
125 by the AOAC, where the highest content is reported for *T. cacao* (19.6-23.9%) with comparable values for *T.*  
126 *grandiflorum* (21.7%) (Sotelo & Alvarez, 1991), while the lowest content is observed for *T. bicolor* (17%)  
127 (Pugliese et al., 2013). Moreover, several authors have assessed the chemical composition of the fats in the three  
128 species (Barbalho et al., 2022), (Torres et al., 2002), (Gilbert-Escrivá et al., 2002), (Jee, 1984), (Vázquez-Ovando  
129 et al., 2015), primarily aiming to identify materials that can function as substitutes or be blended with cocoa fat.  
130 The prevailing methodology involves the initial extraction of fats using an organic solvent, such as hexane, diethyl  
131 ether, or petroleum ether, over a time range of 2 to 8 hours. Subsequently, the lipid profile has been determined  
132 using gas chromatography coupled with different detectors: specifically, an ionisation detector for identifying the  
133 sample composition through fatty acid mixture standards (Torres et al., 2002) (Gilbert-Escrivá et al., 2002), or a  
134 quadrupole mass spectrometer detector for identification based on comparing retention indices and mass spectra  
135 (Barbalho et al., 2022) (Vázquez-Ovando et al., 2015). In both methodologies, silica capillary columns are  
136 employed. **Figure 2** presents the relative abundance of major fatty acids reported in three species. Stearic acid  
137 18:0 is notably higher in *T. bicolor* (~47%), surpassing 10% in comparison to the other reviewed species. In  
138 contrast, oleic acid (18:1, n-9), known for its anti-inflammatory properties through the modulation of immune  
139 responses and reduction of inflammatory markers (Santamarina et al., 2021), showed similar content between *T.*  
140 *grandiflorum* and *T. bicolor*, being superior to that found in *T. cacao*. Palmitic acid 16:0 has also been identified  
141 as a prominent component, especially in *T. cacao*, contributing to the distinctive hardness of cocoa butter.  
142 Particularly, some authors have studied the fatty acids in *T. bicolor*, reporting that its high melting point (~36°C)  
143 offers potential for applications in food and could contribute to mitigating the overexploitation of *T. cacao*, whose  
144 high demand and cultivation costs impact market prices (Gilbert-Escrivá et al., 2002). However, other studies  
145 have presented opposing arguments, suggesting that mixing cocoa butter with fat from *T. bicolor* could result in  
146 an undesirably soft product (Jee, 1984).



147  
 148 **Figure 2.** Fatty acid composition (%) of *T. bicolor*, *T. cacao*, and *T. grandiflorum*. Determined via Gas  
 149 Chromatography analysis, \*Significant differences reported by the authors. Data for construction sourced from  
 150 (Barbalho et al., 2022), (Torres et al., 2002), (Gilabert-Escrivá et al., 2002), (Jee, 1984), (Vázquez-Ovando et al.,  
 151 2015).

152 Additionally, the presence of eicosanoic acid is significantly more pronounced in *T. grandiflorum*, reducing its  
 153 melting point (~32 °C) (Gilabert-Escrivá et al., 2002). Lastly, the occurrence of other fatty acids, such as  
 154 palmitoleic (16:1) <0.35% and docosanoic (22:0) <2%, has been reported in modest proportions across all the  
 155 three species.

156 Works in literature report as well the protein content, primarily utilising the Kjeldahl technique, observing that  
 157 fresh seeds of *T. bicolor* exhibit the highest values (Sotelo & Alvarez, 1991), (Pugliese et al., 2013), (De Souza  
 158 Schmidt Goncalves et al., 2010). This trend is confirmed by experiments conducted by (Vázquez-Ovando et al.,  
 159 2015), who identified a similar pattern in fermented-dehydrated seeds. In contrast, *T. grandiflorum* liquor  
 160 demonstrates higher protein values compared to *T. cacao* liquor in the study conducted by (de Oliveira &  
 161 Genovese, 2013). However, (Pérez-Mora et al., 2018) et al. reported average protein values of *T. bicolor* (31.3),  
 162 *T. grandiflorum* (20.6), and *T. cacao* (25.1) in seeds expressed on a DW (mg/g). These values were determined  
 163 using the Bradford assay, which is based on total protein content and selectively quantifies solubilised proteins.  
 164 The same authors conducted a proteomic analysis of seeds and pulp juice of the three species, employing two  
 165 separation techniques: First Dimension - Isoelectric Focusing and Second Dimension (sodium dodecyl sulphate  
 166 polyacrylamide Gel Electrophoresis). This combined approach provided a high level of resolution to separate and  
 167 visualise proteins in the samples. Following separation, the proteins underwent further analysis by mass  
 168 spectrometry. The results indicated that *T. grandiflorum* presents the greatest number of protein species (122) in  
 169 seeds, followed by *T. bicolor* (94) and *T. cacao* (62). Additionally, the three Theobroma species exhibit common



170 species-specific spots, with 4 in their pulps and 10 in seeds. Some of the identified proteins include vicilin, a  
171 protease inhibitor, and a flavonol synthase/flavanone 3-hydroxylase.

172 Finally, regarding carbohydrate content, higher levels are reported in the pulp of *T. grandiflorum* compared to *T.*  
173 *bicolor* (Pugliese et al., 2013), (Zagmignan et al., 2023), and (Sotelo & Alvarez, 1991). The latter also presents a  
174 higher fibre content (8.8%) in fresh seeds compared to *T. cacao* (3.2%) (Sotelo & Alvarez, 1991). When evaluating  
175 the content of simple sugars, expressed in g/100g DW, sucrose is the predominant sugar in *T. bicolor* (7.7) and *T.*  
176 *grandiflorum* (10.8), while fructose is prevalent in *T. cacao* (7.3). In a study conducted by (Pereira et al., 2017),  
177 it was observed that after a fermentation process with the addition of *L. casei* in *T. grandiflorum* juice, aiming to  
178 generate a probiotic beverage, the initial sugars content in the juice reduced as follows: glucose from 4 to 2.5,  
179 fructose from 10.8 to 2.9, and sucrose from 5.3 to 3.8 (in g/100 mL of beverage).

### 180 **Bioactive compounds profile and their role in antioxidant activities**

181 After describing the chemical composition of these Theobroma species, the presence of bioactive compounds  
182 represents a fascinating add-value for the exploitation of these raw materials in different fields, including the  
183 biomedical one. **Table 2** provides an overview of the different antioxidant compounds and values, extracted from  
184 the three Theobroma species using different processing parameters. For the characterisation and quantification of  
185 these antioxidant compounds, the assessment has predominantly focused on the overall evaluation of phenolic  
186 compounds, that could include phenolic acids, flavonoids, using the Folin–Ciocalteu’s reagent (FCR). This is a  
187 colorimetric method based on an electron-transfer reaction, where the antioxidant species, rich in phenolic  
188 functional groups, serves as the electron donor, and the FCR reagent functions as the oxidant (Pérez et al., 2023).  
189 This assay is widely accepted by the scientific community and applied to different plant materials and foods (Chun  
190 & Kim, 2004). Moreover, numerous studies quantify flavonoids using the aluminium chloride (AlCl<sub>3</sub>) method,  
191 where the flavonoids, with their abundant oxo and hydroxyl groups, exhibit a strong tendency to bind metal ions  
192 like Al(III), typically in a 1:1 ratio. This affinity is subject to experimental variables, notably pH (Matić et al.,  
193 2017) (Shraim et al., 2021), it has not been compared with *T. cacao*. Then, proanthocyanidins, a specific group of  
194 condensed tannins such as catechin and epicatechin, have been evaluated using various methods (Pugliese et al.,  
195 2013), (Pinent et al., 2016), (de Oliveira & Genovese, 2013). One method is represented by the Butanol-HCl  
196 assay, where proanthocyanidins react with butanol-HCl under acidic conditions to produce a pink-coloured  
197 complex, which is then measured spectrophotometrically at around 550 nm (Yu et al., 2023). Another method  
198 involves the use of 4-dimethylaminocinnamaldehyde (DMACA), which results in the formation of a blue-coloured  
199 complex under acidic conditions and is measured in the range of 640-650 nm (Chowdhury et al., 2023). This

200 determination has been reported for *T. grandiflorum* and *T. cacao*. However, comparing results from different  
201 methodologies can be challenging due to specific aspects employed in the determinations, such as variations in  
202 the type of the sample, the conducted extraction procedure, or the chemical used as equivalents, among other  
203 factors.

204 Based on the data found in literature (**Table 2**) the phenolic content was higher in seeds than in pulp. Specifically,  
205 in the study conducted by (Vázquez-Ovando et al., 2015), using a two-stage aqueous extraction for 24 hours,  
206 higher content of phenolics are observed for *T. cacao*, ~40 mg GAE/g, compared to *T. bicolor* (~7 mg GAE/g).  
207 Similar values are reported for *T. grandiflorum* (~8 mg GAE/g), when conducting an aqueous extraction and ~3  
208 mg GAE/g in methanol extraction The optimization of ultrasonic-assisted extraction of *Centaurea* sp. antioxidative  
209 phenolic compounds using response surface methodology. When comparing the phenolic content of fresh pulp  
210 between *T. bicolor* and *T. grandiflorum*, the former exhibits higher content of 1.62 mg/g FW in water extraction  
211 (González et al., 2016) compared to 0.66 mg/g FW in acetone extraction for the latter (Carmona-Hernandez et al.,  
212 2021).

213 **Table 2.** Antioxidant profile, compound extraction parameters, values, and assessment methods for *T. bicolor*, *T. cacao*, and *T. grandiflorum*.

Sample	Compound Extraction parameters	Method	Value (mg/g)	Ref
<b><i>Theobroma bicolor</i></b>				
Pulp lyophilised	Phenols. Acetone 80%, ratio 1:6.6 (w/v), T 0.5 h	FCR (6 min), Na <sub>2</sub> CO <sub>3</sub> 7%, React. T 90 min Abs 760 nm	0.17 TAE DW	(Pérez-Mora et al., 2018)
Pulp lyophilised Aril + seeds	Phenols. H <sub>2</sub> O, concentration of 16-80µg/mL	FCR (10 min), Na <sub>2</sub> CO <sub>3</sub> 12%, React. T 120 min Abs 760 nm	0.10 GAE ext 0.10 GAE ext	(Tauchen et al., 2016)
Seeds fermented defatted	Phenols. MetOH 50%, pH 2, ratio 1:20 (w/v), T 48h	FCR (15 min), Na <sub>2</sub> CO <sub>3</sub> 1M, React. T 15 min Abs 760	6.81 GAE DW	(Vázquez-Ovando et al., 2015)
<b><i>Theobroma cacao</i></b>				
Pulp lyophilised	Phenols. MetOH 70%, ratio 1:20 (w/v), T 2 h	FCR (6 min), Na <sub>2</sub> CO <sub>3</sub> 7%, React. T 90 min Abs 760 nm	0.34 TAE DW	(Pérez-Mora et al., 2018)
Pulp lyophilised Aril + seeds	Phenols. H <sub>2</sub> O, concentration of 16-80µg/mL	FCR (10 min), Na <sub>2</sub> CO <sub>3</sub> 12%, React. T 120 min Abs 760 nm	0.05 GAE ext 0.22 GAE ext	(Tauchen et al., 2016)
Seeds	Phenols. MetOH 50%, pH 2, ratio 1:20 (w/v), T 48h	FCR (15 min), Na <sub>2</sub> CO <sub>3</sub> 1M, React. T 15 min Abs 760	41.41 GAE DW	(Vázquez-Ovando et al., 2015)
Liquor	Phenols. MetOH 70%, ratio 1:20 (w/v), T 2 h	FCR. Abs 750nm	28.45 CE DW	(de Oliveira & Genovese, 2013)
	Proanthocyanidins. MetOH	NH <sub>4</sub> Fe (SO <sub>4</sub> ) <sub>2</sub> ·12H <sub>2</sub> O 2% (w/v), BuOH-HCl 95:5 (v/v) React. T 40 min (95 °C). Abs 520 & 580 nm	10.59 QTE DW	
	Proanthocyanidins, Acetone 70% + acetic acid 0.5%, ratio 1:10 (w/v), T 10 min (USB)	4-dimethylamin cinnamaldehyde-HCl (alcoholic solution). Abs 640 nm	22.01 QTE DW	
Leaves	Phenols. H <sub>2</sub> O, concentration of 16-80µg/mL	FCR (10 min), Na <sub>2</sub> CO <sub>3</sub> 12%, React. T 120 min. Abs 760 nm	0.15 GAE ext	(Tauchen et al., 2016)
<b><i>Theobroma grandiflorum</i></b>				
Pulp lyophilised	Phenols. Acetone 80%, ratio 1:6.6 (w/v), T 0.5 h	FCR (6 min), Na <sub>2</sub> CO <sub>3</sub> 7%, React. T 90 min Abs 760 nm	0.23 TAE DW	(Pérez-Mora et al., 2018)
	Phenols. H <sub>2</sub> O, concentration of (16-80µg/mL)	FCR (10 min), Na <sub>2</sub> CO <sub>3</sub> 12%, React. T 120 min Abs 760 nm	0.16 GAE ext	(Tauchen et al., 2016)

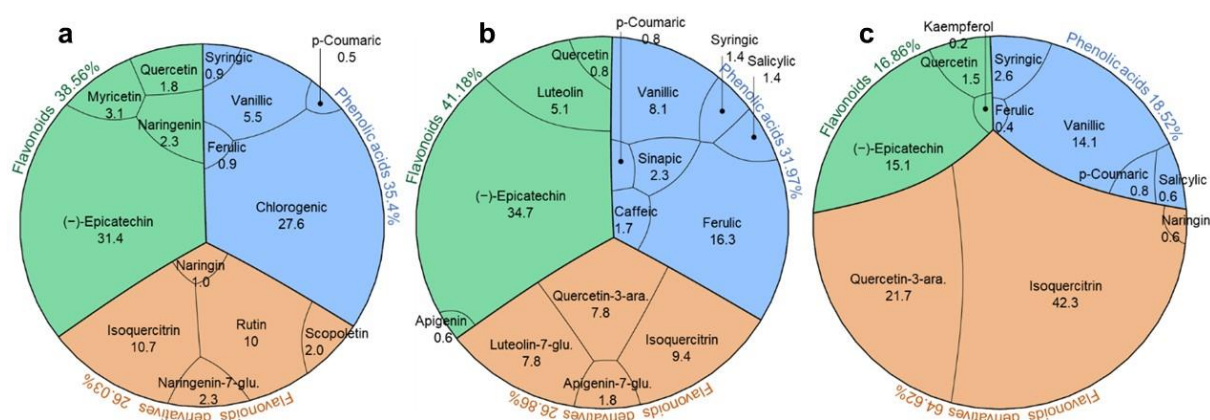
Pulp	Phenols. EtOH 80%, ratio 1:5 (w/v), T 0.5 h (mixer)		0.52 GAE FW	(Carmona-Hernandez et al., 2021)
	Phenols. MetOH 70%, ratio 1:5 (w/v), T 0.5 h (mixer)	FCR (2 min), Na <sub>2</sub> CO <sub>3</sub> 3.5%, React. T 90 min Abs 655 nm	0.47 GAE FW	
	Phenols. Acetone 70%, ratio 1:5 (w/v), T 0.5 h (mixer)		0.66 GAE FW	
	Phenols (EC50). MetOH 70%. T 24 h. From a concentrated extract, dilutions 1:40 (v/v)	FCR React. T 30 min Abs 760 nm	683 GAE ext	(Curimbaba et al., 2020)
Pulp lyophilised	Phenols. H <sub>2</sub> O/ pulp, ratio 2:5 (w/v), T 2 min (mixer), filtration	FCR (1 min), Na <sub>2</sub> CO <sub>3</sub> 15%, React. T 120 min Abs 750 nm	0.02 GAE DW	(Costa et al., 2022)
Pulp dried 40 °C	Phenols. H <sub>2</sub> O, ratio 1:10 (w/v), T 1 h (60 °C, USB)	FCR (8 min), Na <sub>2</sub> CO <sub>3</sub> 20%, React. T 5 h Abs 765 nm	2.01 GAE DW	(Andrade et al., 2022)
	Phenols. EtOH 12%, ratio 1:10 (w/v), T 1 h (USB)		2.64 GAE DW	
Pulp	Phenols. H <sub>2</sub> O, diluted 34%, supernatant filtered,	FCR Na <sub>2</sub> CO <sub>3</sub> 20%, React. T 30 min Abs 765 nm	0.11 GAE DW	(Pereira et al., 2017)
Pulp	Phenols. MetOH 70%, ratio 1:40 (w/v), T 1 min (mixer)	FCR. Abs 750nm	0.06 CE DW	(Pugliese et al., 2013)
Pulp frozen			0.04 CE DW	
Aril	Phenols. H <sub>2</sub> O, concentration of 16-80µg/mL	FCR (10 min), Na <sub>2</sub> CO <sub>3</sub> 12%, React. T 120 min Abs 760 nm	0.06 GAE ext	(Tauchen et al., 2016)
Pulp dried 40 °C	Flavonoids. H <sub>2</sub> O, ratio 1:10 (w/v), T 1 h (60 °C, USB)	NaNO <sub>2</sub> 5% (5 min), AlCl <sub>3</sub> 10% (1 min), NaOH (1 M). Abs 415 nm	2.05 QE DW	(Andrade et al., 2022)
	Flavonoids. EtOH 12%, ratio 1:10 (w/v). T 1 h (USB)		2.47 QE DW	
Pulp fresh	Flavonoids. EtOH 80%, ratio 1:10 (w/v), T 0.5 h (USB)	NaNO <sub>2</sub> 5% (5 min), AlCl <sub>3</sub> 10% (1 M), NaOH (1 min). Abs 415 nm (excitation filter), 520 (reads)	0.04 QE FW	(Carmona-Hernandez et al., 2021)
	Flavonoids. MetOH 70%, ratio 1:10 (w/v), T 0.5 h (USB)		0.01 QE FW	
	Flavonoids. Acetone 70%, ratio 1:10 (w/v), T 0.5 h (USB)		0.63 QE FW	
Pulp	Proanthocyanidins. MetOH: acetic acid (99%).	FeSO <sub>4</sub> ·7H <sub>2</sub> O 154 mg /L of BuOH-HCl (3:2) React. T 15 min (90 °C), Abs 540 nm	5.71 QTE DW	(Pugliese et al., 2013)
Pulp frozen			4.72 QTE DW	
Seeds dried 40 °C	Phenols. H <sub>2</sub> O, ratio 1:10 (w/v),	FCR (8 min), Na <sub>2</sub> CO <sub>3</sub> 20%,	7.89	(Andrade et al., 2022)

	T 1 h (60 °C, USB) Phenols. EtOH 12%, ratio 1:10 (w/v), T 1 h (USB)	React. T 5 h. Abs 765 nm	GAE DW 2.62	
Seeds	Phenols. MetOH 70%, ratio 1:40 (w/v), T 1 min (mixer)	FCR. Abs 750nm	0.22 CE DW	(Pugliese et al., 2013)
Seeds	Phenols	Not reported	7.64 GAE DW	(Pinent et al., 2016)
Seeds lyophilised	Phenols. MetOH 70%, acetic acid 5%. Filtered	FCR (3 min), Na <sub>2</sub> CO <sub>3</sub> saturated, React. T 30 min (37 °C). Abs 750 nm	0.05 CE DW	(De Souza Schmidt Goncalves et al., 2010)
Seeds dried 40 °C	Flavonoids. H <sub>2</sub> O, ratio 1:10 (w/v), T 1 h (60 °C, USB)	NaNO <sub>2</sub> 5% (5 min), AlCl <sub>3</sub> 10% (1 min), NaOH (1 M). Abs 415 nm	5.39 QE DW	(Andrade et al., 2022)
	Flavonoids. EtOH 12%, ratio 1:10 (w/v), T 1 h (USB)		5.06 QE DW	
Seeds	Proanthocyanidins. MetOH: acetic acid (99%)	FeSO <sub>4</sub> ·7H <sub>2</sub> O 154 mg /L of BuOH-HCl (3:2) React. T 15 min (90 °C). Abs 540 nm	0.32 QTE DW	(Pugliese et al., 2013)
	Proanthocyanidins	Not reported	1.98 QTE DW	(Pinent et al., 2016)
	Phenols. MetOH 70%, ratio 1:20 (w/v), T 2 h	FCR. Abs 750nm	7.84 CE DW	
Liquor	Proanthocyanidins. MetOH.	NH <sub>4</sub> Fe (SO <sub>4</sub> ) <sub>2</sub> ·12H <sub>2</sub> O 2% (w/v), BuOH-HCl 95:5 (v/v). React. T 40 min (95 °C). Abs 520 & 580 nm	5.85 QTE DW	(de Oliveira & Genovese, 2013)
	Proanthocyanidins. Acetone 70% + acetic acid 0.5%, ratio 1:10 (w/v), T 10 min (USB)	4-dimethylamin cinnamaldehyde- HCl (alcoholic solution). Abs 640 nm	2.18 QTE DW	
Leaves	Phenols. H <sub>2</sub> O, concentration of 16-80µg/mL	FCR (10 min), Na <sub>2</sub> CO <sub>3</sub> 12%, React. T: 120 min. Abs 760 nm	0.40 GAE ext	(Tauchen et al., 2016)

214 EtOH: ethanol, MetOH: methanol, BuOH: butanol, Folin-Ciocalteu reagent: FCR, TAE: tannic acid equivalents, GAE: gallic acid equivalents,  
215 CE: catechin equivalents, QTE: quebracho tannin equivalents, QE: quercetin equivalents, DW: Dry weight, FW: Fresh weight, T: time, USB: ultrasonic bath

216 Regarding the phenolic compound content in pulps, *T. grandiflorum* presents values in the range of 0.02 to 2.64  
217 mg/g, where lower values are observed using aqueous extractions; however, assisting the extraction with  
218 ultrasounds helps to get the highest value. Indeed, ultrasound technique is effective in recovering bioactive  
219 compounds from plant materials, thereby enhancing the processing (Bouafia et al., 2021). This effectiveness  
220 extends not only to the extraction of secondary metabolites but also to the extraction of pectin from *T. cacao* pod  
221 husk, which presents a rich content of phenolics when compared with commercial pectin (Girón-Hernández et al.,  
222 2024). Furthermore, the influence of the solvent has also been assessed in *T. grandiflorum*, revealing that a 12%  
223 ethanol concentration in the extraction solution increased the flavonoid and phenolic content when conducting  
224 extractions on dehydrated pulp (Andrade et al., 2022). However, the same study demonstrated that water  
225 represents the most effective solvent when evaluating these compounds in seeds. Furthermore, an investigation of  
226 different solvents (acetone, methanol and ethanol) on the *T. grandiflorum* has indicated that acetone is capable to  
227 enhance the yield of phenolic compounds, while ethanol has been proved to be the optimal solvent for quantifying  
228 flavonoids from fresh samples (Carmona-Hernandez et al., 2021). Certainly, an increase in ethanol concentration  
229 up to 40% in the extraction solution resulted in improved recovery of both phenolic and flavonoid contents, as  
230 evaluated in powdered leaf samples of *O. stamineus* by Chew (Chew et al., 2011). The quantification of the total  
231 flavonoids in both the pulp and seeds of *T. grandiflorum* has revealed values for the pulp ranging from 0.01 to  
232 0.63 mg QE/g on a fresh weight, with higher results obtained from extractions using acetone (Carmona-Hernandez  
233 et al., 2021). Ethanol also exhibited higher values compared to aqueous extraction, with 2.05 and 2.47 mg QE/g  
234 on a dry weight basis respectively (Andrade et al., 2022). Indeed, flavonoid solubility is relatively low in water,  
235 prompting some authors to recommend the use of water-organic solvent mixtures to enhance the extraction of  
236 these compounds (Chebil et al., 2007). However, in studies conducted by (Andrade et al., 2022), the flavonoid  
237 content doubled in aqueous extraction from the seeds compared to the pulp of *T. grandiflorum*, particularly  
238 yielding higher amounts compared to extraction with a water-ethanol mixture. Additionally, the total flavonoid  
239 values reported in *T. grandiflorum* seeds are lower compared to those reported for defatted *T. cacao* seeds (~8 mg  
240 epicatechin equivalents/g) using comparable methodologies (Zzaman et al., 2014). Proanthocyanidins, another  
241 family of compounds, are also assessed, demonstrating similar values (~6 mg QTE/g DW) in the pulp and liquor  
242 of *T. grandiflorum*, while the evaluation of these compounds in *T. cacao* liquor is ~11 mg QTE/g DW when using  
243 methanol extractions (Pugliese et al., 2013) (de Oliveira & Genovese, 2013). The total proanthocyanidins content  
244 in *T. grandiflorum* seeds ranges from 0.3 to 2 mg QTE/g DW with comparable methodologies (Pugliese et al.,  
245 2013), (Pinent et al., 2016).

246 Furthermore, different compounds have been identified in the pulp and seeds of *Theobroma* species, employing  
 247 chromatographic techniques based on their UV spectra or mass spectrometry (MS). Particularly, UV has been  
 248 utilised to quantify the content of hypolaetin, isoscutellarein, catechol, isoscutellarein, quinic acid, chlorogenic  
 249 acid, and theobromine based on their standards (Andrade et al., 2022; Carmona-Hernandez et al., 2021; de Moraes  
 250 Barros et al., 2016; de Oliveira et al., 2015; Pugliese et al., 2013; Sotelo & Alvarez, 1991; Tauchen et al., 2016).  
 251 In these studies, compounds such as catechin and kaempferol are either traced or not detected. The MS technique  
 252 has revealed a greater number of compounds based on their mass-to-charge ratio, facilitating the identification of  
 253 Theograndins, which have shown lower antioxidant capacity compared to other known polyphenols. However,  
 254 their bioavailability might be better due to their water solubility, thus hydrolysing to their corresponding  
 255 flavonoids (Yang et al., 2003).  
 256 Regarding comparative studies between the three species, Tauchen et al. (2016) conducted a metabolomic analysis  
 257 on ethanolic extracts (70%) of edible and medicinal Amazonian plants, including *T. bicolor*, *T. cacao*, and *T.*  
 258 *grandiflorum*, using Ultra-High Performance Liquid Chromatography and Tandem Mass Spectrometry (UPLC-  
 259 MS/MS). **Figure 3** illustrates the major components in the pulp of these species, derived from data obtained from  
 260 Tauchen et al., 2016. Three big groups of compounds have been reported: flavonoids, flavonoids derivatives and  
 261 phenolic acids. Based on the reviewed data, *T. bicolor* shows greater similarity to *T. cacao*, presenting similar  
 262 proportion and species of compounds on the most abundant within the groups. In the case of *T. bicolor*, it is  
 263 particularly interesting due to its usable pulp proportion of ~26% (González et al., 2016), rendering this fruit a  
 264 valuable raw material for flavonoids. Additionally, unlike the other two species, this fruit also contains a notable  
 265 proportion of chlorogenic acids and rutin. *T. cacao* is characterised by the presence of luteolin, and *T.*  
 266 *grandiflorum* by showing the presence of kaempferol in low amounts.



267 **Figure 3.** Treemaps of the relative abundance (%) of major metabolites in the pericarps (pulp) of (a) *T. bicolor*,  
 268 (b) *T. cacao*, and (c) *T. grandiflorum*, determined via UHPLC–MS/MS analysis. Figure created using data sourced  
 269 from (Tauchen et al., 2016)  
 270

271 Moreover, the antioxidant capacity has been assessed mainly on various components (pulp, seed, liquor) of fruits  
272 from the *Theobroma* species using analyses such as 2,2-diphenyl-1-picrylhydrazyl (DPPH), Oxygen Radical  
273 Absorbance Capacity (ORAC), 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) (ABTS<sup>•+</sup>), Ferric  
274 Reducing Ability of Plasma (FRAP), or testing the antioxidant activity of Butylated hydroxytoluene (BHT). After  
275 *T. cacao*, *T. grandiflorum* presents more available information from literature. However, comparing the reported  
276 values is not straightforward due to the diversity of the experiments, but it can generally be expected that the  
277 antioxidant capacity tends to be higher in the materials extracted from the seeds compared to those ones from the  
278 pulp (Pugliese et al., 2013), (Andrade et al., 2022), (Tauchen et al., 2016), and (González et al., 2016). Particularly,  
279 the most used method for evaluating antioxidant activity is DPPH. In this regard, a comparative study on pulp,  
280 showed higher values for *T. grandiflorum* (~188 µg Trolox Equivalent (TE)/mg), followed by *T. bicolor* (~107 µg  
281 TE/mg), while the pulp of *T. cacao* presented significant differences with a value of approximately 52 µg TE/mg  
282 extract (Tauchen et al., 2016). However, other studies evaluated DPPH in the liquor of *T. grandiflorum* and *T.*  
283 *cacao*, presenting values of ~79.6 and 19.1 µg TE/mg per gram of dry weight, respectively (de Oliveira &  
284 Genovese, 2013), showing lower antioxidant capacity compared with the corresponding pulp.

285 Moreover, other studies have reported their finding as IC<sub>50</sub> value, that indicates the concentration of a sample  
286 required to scavenge 50% of DPPH free radicals (Martinez-Morales et al., 2020). For instance, (Yang et al., 2003)  
287 reported values ranging from 40 to 90 µM for the specific fraction of Theograndin I and 120 µM for Theograndin  
288 II. Then, the authors have also analysed the influence of the solvent used for extracting the compounds, revealing  
289 that acetone is the most favourable one, presenting a DPPH IC<sub>50</sub> values of Log ascorbic acid equivalent 2.6  
290 µg/mL, calculated from their reported 44.0 mg AAE/100 g (fresh weight).

291 Furthermore, ORAC was the second most popular method, (Tauchen et al., 2016) evaluated fresh pulps and  
292 observed that *T. grandiflorum* showed higher values (~435 µg TE/mg extract) than *T. bicolor* (243 µg TE/mg  
293 extract) and *T. cacao* (180 µg TE/mg extract). Similarly, it has been observed that the ORAC presents higher  
294 values when measured in fresh pulp in wet basis (~217 µmol TE/g DW) compared to frozen (~109 µmol TE/g  
295 DW) (Pugliese et al., 2013). Values decreased significantly (~17 µmol TE/g DW) when the pulp was dehydrated  
296 using forced air at 40 °C; however, in vitro digestion and fermentation of the dehydrated pulp increased antioxidant  
297 capacity (~94 and ~108 µmol TE/g DW) respectively. Additionally, studies conducted in liquor showed higher  
298 values for *T. cacao* (~451 µmol TE/ g DW) compared to *T. grandiflorum* (~136 µmol TE/ g DW).

299 Moreover, ABTS<sup>•+</sup> values in fresh pulp and seeds of *T. grandiflorum* were similar, ~29 µmol TE/ g of sample, and  
300 increased after in vitro digestion to ~46 µmol TE/ g. However, after fermentation with *Lactobacillus* bacteria



301 (rhamnosus, longum, delbrueckii) this value decreased for the pulp down to ~19  $\mu\text{mol TE/g}$  but not for the seed  
302 (~37  $\mu\text{mol TE/g}$ ) after fermentation (Andrade et al., 2022). A similar trend was observed after fermenting the  
303 pulp in a ratio 34% (w/v) for 18 hours with *L. casei*, resulting in increased ABTS<sup>+</sup> values for the treated pulp  
304 from 0.52 to 0.67  $\mu\text{mol TE/mL}$  (Pereira et al., 2017). Regarding the determination of antioxidant capacity using  
305 the FRAP method, no reports were found for samples of *T. bicolor*, while results ranging between 15.26 and 18.69  
306  $\mu\text{mol TE/g}$  have been reported for non-fermented seeds of *T. grandiflorum* (Pinent et al., 2016), (Andrade et al.,  
307 2022). The antioxidant capacity evaluated by this method was reduced after both *in vitro* digestion and  
308 fermentation of the seeds (Andrade et al., 2022). Furthermore, a study examining flowers of the *Theobroma* genus  
309 evaluated their aqueous extracts, demonstrating antioxidant capacities assessed using DPPH ranging from 199 to  
310 1634  $\mu\text{mol TE/g}$  and from 361 to 1991  $\mu\text{mol TE/g}$  for ABTS<sup>+</sup>. However, the study did not specifically report the  
311 related species (Mar et al., 2021). Finally, information related to leaves showed that *T. grandiflorum* presents a  
312 higher antioxidant capacity compared to *T. cacao* based on DPPH and ORAC assays (**Table 2**).

### 313 **Therapeutic potential**

314 The content of secondary metabolites, particularly the fraction of phenolic extracts from the *Theobroma* species  
315 evaluated in this review, has prompted the develop of various experiments to assess their potential in therapeutic  
316 and biomedical applications. To date, research has primarily focused on the biological evaluation of both seed and  
317 pulp of *T. grandiflorum*, with some studies comparing this species with *T. cacao*. The following information  
318 comprises *in vitro*, including cellular tests, and *in vivo* animal studies. No clinical studies have been reported;  
319 however, both pulp, butter, and liquor extracted from these fruits are frequently consumed without any reported  
320 adverse effects.

#### 321 *In vitro* studies

322 The phenolic content in *T. grandiflorum* has sparked interest in its potential for anti-diabetic effects thanks to its  
323 capacity for  $\alpha$ -amylase inhibition, the enzyme responsible for breaking down carbohydrates, specifically starches,  
324 into smaller sugars such as maltose and glucose (Andrade et al., 2022). In this context, a rich fraction of flavonoids  
325 was obtained through Solid Phase Extraction using a polyamide column and evaluated for its ability to reduce  $\alpha$ -  
326 amylase activity, exhibited a half-maximal inhibitory concentration (IC<sub>50</sub>) at 1.1 mg of dried *T. grandiflorum*  
327 pulp/mL of reaction, expressed in terms of catechin equivalents, totalling 0.3 mg/mL (De Souza Schmidt  
328 Goncalves et al., 2010). Furthermore, another study examined the interaction of ethanolic and aqueous extracts  
329 from pulp and seeds of *T. grandiflorum* with the  $\alpha$ -amylase enzyme, following simulated gastrointestinal digestion  
330 and probiotic fermentation (using *L. delbrueckii*, *L. jhonsoni*, *L. rhamus*, and *B. longum*). As result, the aqueous

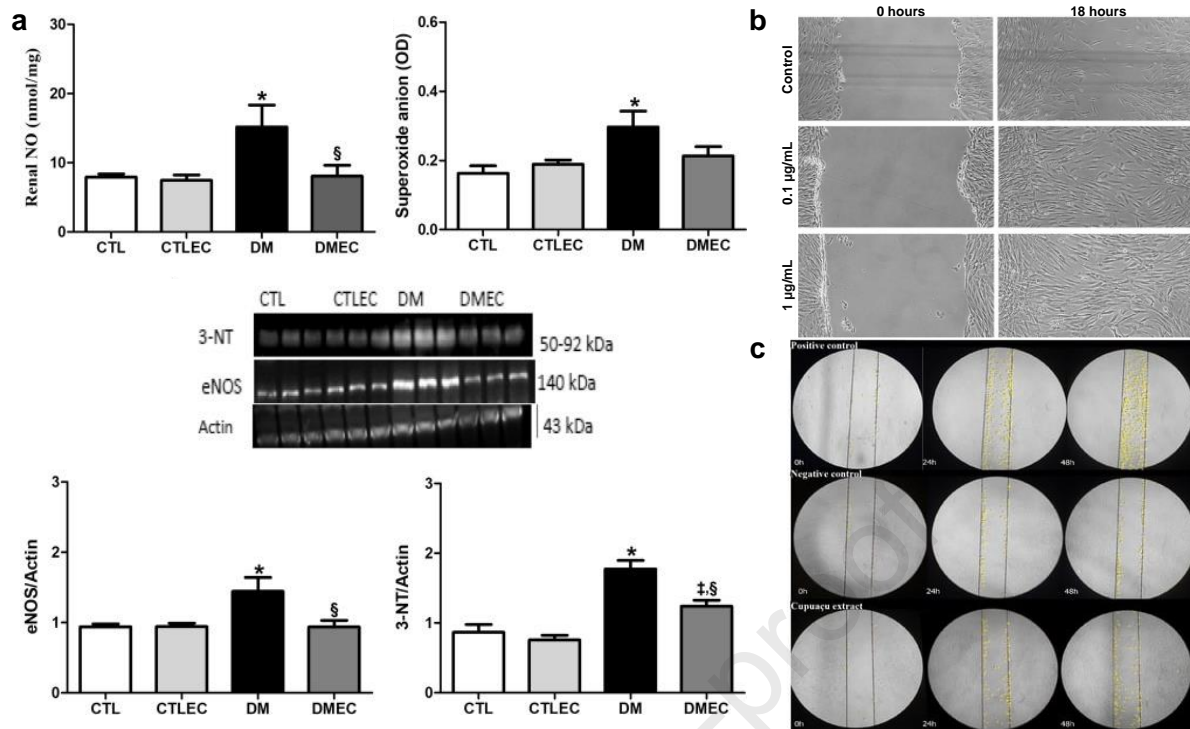
331 extract from *T. grandiflorum* seeds exhibited a higher percentage of  $\alpha$ -amylase inhibition (~97.4%) compared to  
332 the ethanol extracts from the pulp and seeds, which showed moderate inhibition ranging between 57 and 75%  
333 (Andrade et al., 2022). Another finding to consider from this research was that, after simulating gastrointestinal  
334 digestion, the aqueous extract of seeds and pulp exhibited the presence of flavonoids such as epicatechin,  
335 rhamnetin and daidzein with contents of ~112.2, 7.5, and 0.5  $\mu\text{g/g}$  for pulp respectively (Andrade et al., 2022).  
336 Other interesting application is represented by the skincare, where fat extracted from *T. grandiflorum* seeds has  
337 been utilised in the production of emulgels, a combination of emulsions and gels (Erum et al., 2024). Various fat  
338 proportions (5, 10, and 20%), combined with polysorbate 80, dehydroacetic acid benzyl alcohol and water, where  
339 heated to produce an emulsion, and then incorporated into a carbomer gel. The resulting emulgel was studied in  
340 vitro for its UV light filtering ability. Samples at 5, 10, and 20% concentrations, diluted with 0.2 mg/mL ethanol,  
341 were measured for UV absorption (290-320nm). Sun protection factor was calculated using Mansur's equation.  
342 The resulting that the formulation with the highest fat concentration (20%) exhibited the best sun protection factor  
343 (~12), making it suitable for sunscreen product development (Aparicio-Álvarez et al., 2023).

344 Finally, the in vitro inhibitory effect of Amazonian fruit juices on CYP3A4, a crucial enzyme in the metabolism  
345 of many medications, was assessed by incubating liver microsomes containing the CYP3A4 enzyme with  
346 midazolam and the juice extracts. Following the reaction period, samples were analysed using high-performance  
347 liquid chromatography with a diode array detector. *T. grandiflorum* juice presented interesting results, where the  
348 pulps not only reduced the enzyme activity but also exhibited residual activity on 44.3% compared to the positive  
349 control star fruit (*Averrhoa carambola*) (Costa et al., 2022). Therefore, the fruit juice of *T. grandiflorum* could  
350 potentially enhance the assimilation of certain drugs, thereby reducing the amount of drug consumed. Based on  
351 the authors' observation, this activity could be attributed to the proportion of flavonoids contained in the fruit  
352 juice.

353 Certainly, *in vitro* cell tests are considered crucial to proceed with the validation of the efficacy of biocompounds  
354 in animal experimentation. Among the scrutinised *Theobroma* species, attention is once again directed towards  
355 *Theobroma grandiflorum*. Assessments have mainly focused on its cytotoxicity and interaction with intestinal  
356 cells (e.g., Caco-2), oxidative stress with renal mesangial cells, regenerative ability with skin fibroblasts, and  
357 antitumoral capacity against alveolar epithelial cells, which can be the origin of lung cancer. Particularly, some  
358 authors studied the fractionation of a seed extract, leading to the identification of two compounds labelled as  
359 Theograndins I and II (Yang et al., 2003). Despite demonstrating a reduced antioxidant capacity compared to other  
360 flavonoid compounds found in *T. grandiflorum* seeds, such as epicatechin, kaempferol, and quercetin,

361 Theograndins exhibit heightened water solubility owing to their sulphated structure, thereby enhancing their  
362 bioavailability in contrast to other flavonoids. Theograndin I displayed weak cytotoxicity in the HCT-116 and  
363 SW-480 human colon cancer cell lines, with IC50 values of 205  $\mu\text{M}$  and 164  $\mu\text{M}$ , respectively. Meanwhile,  
364 Theograndin II exhibited cytotoxic effects in the HCT-116 and SW-480 human colon cancer cell lines, with IC50  
365 values of 143  $\mu\text{M}$  and 125  $\mu\text{M}$ , respectively (Yang et al., 2003). Interestingly, the unfermented *T. grandiflorum*  
366 seed extract has been compared with grape seed extract (rich in proanthocyanidins) for its antioxidant effect on  
367 Caco-2 and STC-1 cells (Pinent et al., 2016). It was found that *T. grandiflorum* extract prevented tert-  
368 butylhydroperoxide-induced oxidative stress in both cell lines for one hour after application at a lower  
369 concentration than grape seeds. However, no effect was observed for either extract after 20 hours. Regarding to  
370 oxidative stress, Purano et al. (Punaro et al., 2019) assessed the impact of *T. grandiflorum* pulp on nitrosative  
371 stress and inflammatory mediators. The authors cultured mouse immortalised mesangial cells under high glucose  
372 concentrations with and without the extract, at concentrations of up to 500 mg/mL, for a maximum of 72 hours.  
373 Following this period, the cells underwent analysis for proliferation, nitric oxide levels, and reactive oxygen or  
374 nitrogen species (**Figure 4a**). The findings revealed significant antioxidant properties, as the extract was able to  
375 reduce oxidative and nitrosative stress in the presence of high glucose, while also maintaining normal viability  
376 and showing no cytotoxic effects for up to 72 hours on the cells.

377 Furthermore, cytocompatibility of the extracts has been evaluated using fibroblasts cells. Specially, in vitro tests  
378 with L929 cells, derived from the subcutaneous connective tissue of male mice, showed that the aqueous extracts  
379 of the peel and seeds exhibited very high cell viability ranging from 81 to 109% (evaluated by MTT assay), at  
380 concentrations from 10 to 1000  $\mu\text{g/mL}$  (Andrade et al., 2022). Additionally, Barbalho et al (Barbalho et al., 2022)  
381 studied the potential of *T. grandiflorum* seed extract in expediting the wound healing process was evaluated by  
382 incubating primary human dermal fibroblasts cells with the seed extract at a concentration of 100  $\mu\text{g/mL}$ . After  
383 24 hours, the fibroblasts presented high viability (assessed using calcein assay), but they also showed an increase  
384 in their metabolic activity via MTT, particularly evident after 4 and 7 days of incubation. Similar results were  
385 reported by Sano et al. (Sano et al., 2018), who observed that *T. grandiflorum* butter induced fibroblast cell  
386 confluence 18 hours post-wound formation, with more evident results when using 1  $\mu\text{g/ml}$  of *T. grandiflorum*  
387 butter compared with the concentration of 0.1  $\mu\text{g/ml}$  as well as with the control represented by only cells cultured  
388 on tissue culture plate (**Figure 4b**).



389

390 **Figure 4.** *In vitro* biological assessment of *T. grandiflorum* (a) against nitrosative stress: NO, superoxide anion,  
 391 3-NT and eNOS in the kidneys after treatment (NO: nitric oxide; 3-NT: 3-nitrotyrosine; eNOS: endothelial nitric  
 392 oxide synthase; EC: extract of cupuaçu. Control (CTL); control plus EC (CTLEC); diabetic (DM); diabetic plus  
 393 EC (DMEC) (Punaro et al., 2019)) and on improving wound healing after scratch tests as demonstrated by (Sano  
 394 et al., 2018) (b) and (Barbalho et al., 2022) (c).

395 Furthermore, Barbalho et al. observed the regenerative potential of *T. grandiflorum* in supporting wound closure  
 396 immediately after scratch, 24 and 48 h later (Figure 4c) in addition to a significant induction in mRNA expression  
 397 of MKI67 (a cellular proliferation marker) and elastin, without promoting excessive expression of HAS2, which  
 398 is associated with abnormal keratinocyte migration (Barbalho et al., 2022). Additionally, there was a lack of  
 399 expression of matrix metalloproteinases, which are associated with poor lesion healing. Therefore, *T. grandiflorum*  
 400 seed extract can represent suitable biocompatible compounds with regenerative tissues functionalities.

401 Finally, a preliminary study is reported in literature on the additional e.g., antitumoral ability of *T. bicolor* extracts.  
 402 Indeed, a profiling study on the bioactivity of 280 plants in Panama reported that the leaves of *T. bicolor* exhibited  
 403 outstanding antioxidant capacity as evaluated by a non-enzymatic test (Roy et al., 2019). However, the extract  
 404 from *T. bicolor* leaves was not found to preferentially inhibit the growth of the lung cancer cell line A549 alveolar  
 405 epithelial cells (EC50 < 10 µM).

406 *In vivo* animal studies

407 Regarding the *in vivo* validation of *T. bicolor* and *T. grandiflorum* to assess their therapeutic properties, it is noted  
408 that the available information focuses exclusively on *T. grandiflorum*, leaving a gap for research on *T. bicolor*.  
409 Among the gathered literature, studies related to the processing within the intestinal transit have reported the  
410 potential benefits of *T. grandiflorum* extracts to mitigate reactive nitrogen and oxygen species, and its association  
411 with diabetes and inflammatory processes are highlighted. Particularly, a study with C57BL/6 mice provided  
412 insights into how *T. grandiflorum* pulp extracts are processed in the gastrointestinal tract, revealing their  
413 distribution and microbial metabolic conversion (de Moraes Barros et al., 2016). The resulting compounds were  
414 mainly found in the stomach and small intestine, with lesser amounts in the caecum and colon due to microbial  
415 activity. Furthermore, flavonoids bound to sugars and epicatechin metabolites were detected in the caecum, colon  
416 and faeces. Interestingly, glucuronide flavones were hydrolysed into aglycones (the flavonoid itself, devoid of any  
417 attached sugar molecules), while procyanidins were transformed into different metabolites. Finally, this study  
418 mentioned that *T. grandiflorum* flavonoids can influence the intestinal microbiota, affecting the profile of  
419 microbial metabolite.

420 Other works have investigated the impact of *T. grandiflorum* liquor (de Oliveira & Genovese, 2013), (de Oliveira  
421 et al., 2015) and *T. grandiflorum* pulp (Punaro et al., 2019) on oxidative stress and nitrosative stress, respectively,  
422 in diabetic-mimicking conditions, revealing differences in their mechanisms and pathways. Particularly, Belchor  
423 and Genovese (de Oliveira & Genovese, 2013) examined the effects of the daily consumption of *T. grandiflorum*  
424 and *T. cacao* liquors (3.6 and 7.2 g/kg body weight respectively) on oxidative stress and lipid profile in induced  
425 diabetic rats. Both liquors improved body weight gain and liver weight/body weight ratio, while also improving  
426 the lipid profile by reducing triacylglycerol levels and increasing HDL-cholesterol levels, particularly with higher  
427 liquor doses. Additionally, extracts enhanced plasma antioxidant capacity and reduced lipid peroxidation in a  
428 dose-dependent manner. The synergy between fatty acids and phenolic compounds in these liquors may contribute  
429 to mitigating hypertriglyceridemia linked to insulin deficiency. Furthermore, the researchers found that despite *T.*  
430 *grandiflorum* liquor containing fewer phenolic compounds than *T. cacao*, its lower palmitic acid content can  
431 improve lipid profile and antioxidant status in the diabetic rats, underscoring the benefit of *T. grandiflorum*'s  
432 specific fatty acid and phenolic profile over *T. cacao*. This investigation on the liquors of *T. cacao* and *T.*  
433 *grandiflorum* continued when Prof. Genovese's research group decided to explore the impact of their phenolic  
434 content on oxidative stress, potentially associated with metabolic changes induced by a high-fat diet (de Oliveira  
435 et al., 2015). Upon identifying flavonoids unique to *T. grandiflorum*, absent in cocoa liquor, the animals were  
436 divided into groups receiving either a standard or high-fat diet for 16 weeks, with some rats receiving one of the

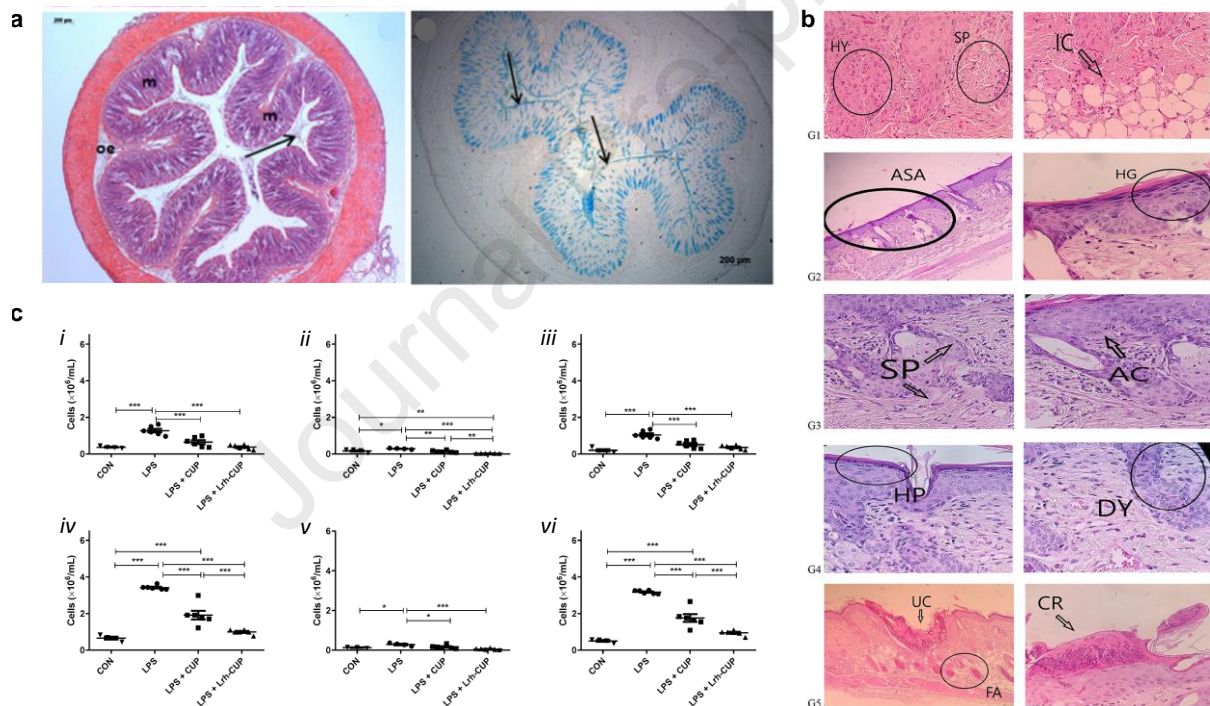
437 liquor extracts during the final 4 weeks. The results showed that both extracts effectively reduced liver damage  
438 from the high-fat diet by lowering lipid peroxidation and boosting antioxidant capacities in plasma and tissues.  
439 Importantly, only phenolics from *T. grandiflorum* liquor improved glucose tolerance by enhancing insulin  
440 sensitivity, attributed to its unique flavonoids.

441 Moreover, concerning specifically the effect of *T. grandiflorum* on oxidative stress, Curimbaba et al. reported that  
442 its antioxidant compounds, released by intestinal bacteria, can offer protection. *T. grandiflorum* pulp flour reduced  
443 diarrhoea by 75% and colon adherence by 50% in animals with induced intestinal damage (Curimbaba et al.,  
444 2020). Authors found that these effects underscored its intestinal antioxidant and anti-inflammatory properties and  
445 its ability to enhance mucin production for barrier function. However, the severity of intestinal damage diminished  
446 this protective effect (**Figure 5a**). Indeed, enzymatically, myeloperoxidase and alkaline phosphatase activities  
447 decreased, but glutathione depletion in colon tissue persisted. Additionally, dietary *T. grandiflorum* lowered IL-6  
448 and IL-1 $\beta$  levels in the colon but had no effect on short-chain fatty acid production, associated with TNF- $\alpha$   
449 reduction. On the other hand, regarding nitrosative stress, Punaro et al. investigated the impact of *T. grandiflorum*  
450 pulp extract on renal complications in induced diabetic rats (Punaro et al., 2019). They administered 1 mL/day of  
451 *T. grandiflorum* (1 g/mL) to adult male rats for 8 weeks, resulting in reduced food intake and nitrosative stress, as  
452 evidenced by decreased levels of O<sub>2</sub><sup>-</sup>, NO, and 3-NT (a marker of nitrosative stress). Additionally, they observed  
453 inhibition of NF- $\kappa$ B, which plays a crucial role in inflammation-related renal damage, and reduced cytokine  
454 production, particularly IL-6. These findings suggest that *T. grandiflorum* extract may help mitigate nitrosative  
455 stress and inflammation, potentially delaying renal complications in diabetic patients.

456 As mentioned before, another area with potential use of the *T. grandiflorum* products is the skincare, where in  
457 their *in vivo* evaluation has been assessed by Aparicio-Alvarez et al. by using photoprotective formulations  
458 containing up to 20% of *T. grandiflorum* butter (producing an emulgel). in Balb/c male mice (Aparicio-Álvarez  
459 et al., 2023). The animals were shaved and topically treated with the emulgel three times over 24 hours.  
460 Subsequently, the mice were irradiated from a UVB lamp (5,000 J m<sup>-2</sup>) for 15 consecutive sessions of 30 minutes  
461 each. Mice treated with the formulation containing *T. grandiflorum* butter at 20%, displayed either minor damage  
462 or reduced severity of the damage, comparable to a commercial standard photoprotector (**Figure 5b**).

463 Finally, investigations on the potential immunomodulatory effect of *T. grandiflorum* pulp have been proposed in  
464 literature, mainly when interacting with *Lactobacillus*. For instance, Zagnignan et al. examined the impact of  
465 fermenting the *T. grandiflorum* juice with *Lactobacillus rhamnosus* on the immunomodulatory response during  
466 induced short-term endotoxemia in C57BL/6 mice (Zagnignan et al., 2023). The study revealed notable

467 enhancements in various facets of the endotoxemic condition, including reductions in weight loss (observed in  
 468 the spleen, liver, intestine, and kidneys), hypothermia, severity index, and cell migration (**Figure 5c**). These  
 469 improvements were notably superior to those observed in control endotoxemic groups, which were administered  
 470 either juice alone or saline solution. Furthermore, another study (Barros-Santos et al., 2020) focused on the  
 471 isolation of *Lactobacillus plantarum* strains from fermented *T. cacao* and *T. grandiflorum* pulps. Subsequently,  
 472 the cognitive, anxiety, and depressive-like behaviours in male swiss mice were evaluated after supplementation  
 473 with *Theobroma* extracts separately. The findings indicated that *Lactobacillus plantarum* 286, isolated from *T.*  
 474 *cacao*, exhibited notable effects. Conversely, strain 81 isolated from *T. grandiflorum* did not elicit any discernible  
 475 effects on mouse models of anxiety and depressive-like behaviours. These insights underscore the potential  
 476 significance of *Lactobacillus* species isolated from fermented *Theobroma* pulps in modulating physiological  
 477 responses, particularly in the context of mental health.



478

479 **Figure 5.** *In vivo* biological assessment of *T. grandiflorum*. Photomicrographs of rat colons treated with *T.*  
 480 *grandiflorum* diets using hematoxylin and eosin (Left) and PAS/Alcian Blue (Right) (Curimbaba et al., 2020) (a);  
 481 Histopathological evaluation of the skin of mice after 2 weeks. G1: mice exposed to UVB light without any  
 482 treatment (negative control group): (Left) HY, hypergranulosis; SP, spongiosis; (Right) IC, increased collagen;  
 483 G2: mice exposed to UVB light treated with commercial sunscreen product (positive control): (Left) ASA, absence  
 484 of skin attachments; (Right) HG, hypergranulosis; G3, mice exposed to UVB light treated with *T. grandiflorum*  
 485 seed butter emulgel at 5%: (Left) SP, spongiosis; (Right) AC, acanthosis; G4, mice exposed to UVB light treated  
 486 with *T. grandiflorum* seed butter emulgel at 10%: (Left) HP, hyperplasia; (Right) DY, dysplasia; G5, mice exposed  
 487 to UVB light treated with *T. grandiflorum* seed butter emulgel at 20%: (Left) UC, ulcer; FA, focal atrophy of hair

488 follicles; (Right) CR, crust. (Aparicio-Álvarez et al., 2023)(b); Effects of fermented and unfermented *T.*  
489 *grandiflorum* juice in the migration of cells to the peritoneal cavity in mice submitted to LPS-mediated  
490 endotoxemia. (i) Total leukocytes in the peritoneal cavity after 6 h of LPS-mediated endotoxemia; (ii)  
491 Polymorphonuclear leukocytes in the peritoneal cavity after 6 h of LPS-mediated endotoxemia; (iii) Mononuclear  
492 cells in the peritoneal cavity after 6 h of LPS-mediated endotoxemia; (iv) Total leukocytes in the peritoneal cavity  
493 after 120 h of LPS-mediated endotoxemia; (v) Polymorphonuclear leukocytes in the peritoneal cavity after 120 h  
494 of LPS-mediated endotoxemia; (vi) Mononuclear cells in the peritoneal cavity after 120 h of LPS-mediated  
495 endotoxemia (Zagmignan et al., 2023) (c).

#### 496 **Current Research and Future Prospects**

497 Undoubtedly, *T. bicolor* presents an intriguing subject for comprehensive investigation, given the compelling  
498 bioactive profile of its fruits, enriched with phenolic compounds. Employing mass spectrometry techniques, *T.*  
499 *bicolor* has been linked with the presence of noteworthy compounds such as naringenin, scopoletin, myricetin,  
500 alongside a substantial quantity of chlorogenic acids. -

501 Consequently, despite the time and resource-intensive nature of characterising the chemical composition of both  
502 species, undertaking such an endeavour would yield substantial benefits. For instance, identifying and quantifying  
503 their secondary metabolites using chemical standards would furnish more precise insights valuable in tailoring  
504 therapeutic applications. For *T. bicolor* specifically, while existing reports focus on total polyphenol content,  
505 discerning the specific concentrations of flavonoids and proanthocyanidins warrants attention. Further exploration  
506 into its antioxidant capacity, assessed through the FRAP, alongside expanding information on other reported  
507 determinations like ORAC, DPPH, and ABTS<sup>•+</sup>, holds promise. By augmenting data on this species across these  
508 tests, seasonal effects on the chemical composition of its fruits could potentially be elucidated.

509 In terms of biological analyses, a constructive approach would involve mirroring the precedents set by studies on  
510 *T. grandiflorum*. This entails investigating the potential of *T. bicolor*; concerning blood sugar regulation, due to its  
511 chlorogenic acid content. Extracts from this species may exert effects on glucose absorption and production in the  
512 liver, holding implications for managing diabetes. Furthermore, the notable presence of phenolic compounds in  
513 extracts from both *T. bicolor* and *T. grandiflorum* warrants evaluation for their interaction with Caco-2 cells.  
514 Existing research points towards a protective effect of phenolic compounds on the digestive system. Exploring  
515 additional components such as liquor and butter from the fruits, noted for their heightened antioxidant capacity,  
516 could yield further insights.

517 Furthermore, the polyphenols found in *Theobroma* species, primarily in *T. cacao*, such as flavonoids and phenolic  
518 acids, have garnered significant research interest for their potential applications in promoting mental health. These  
519 compounds have shown promise in alleviating symptoms of depression, managing stress, and enhancing



520 resilience. However, further research, including not only *T. cacao* but other *Theobroma* species mentioned in this  
521 review, is necessary to fully elucidate the mechanisms and efficacy of *Theobroma* secondary metabolites in  
522 promoting mental health. Preliminary animal studies suggest promising potential for their use as adjunctive or  
523 complementary therapies in the management of depression, stress, and resilience enhancement. Additionally,  
524 exploring deeper into the interaction with friendly microorganisms such as *Lactobacillus* could provide insights  
525 into this symbiotic association with mental health. Building upon recommendations from studies reviewed here  
526 that present evidence from animal models, future investigations could prolong the assessment period for  
527 interactions of fruit extracts. Additionally, ethical considerations suggest an expansion in the number of animals  
528 to enhance result variability. Given the absence of reported adverse effects from consuming fruits of the studied  
529 *Theobroma* species, coupled with evidence supporting cytocompatibility, clinical interventions employing  
530 extracts from these fruits could offer a more nuanced evaluation of their therapeutic potential.

531 In this regard, nanotechnology has been recognised as a vital tool for creating promising nanostructures in  
532 potential nutraceutical delivery systems (Martínez-Ballesta et al., 2018). These nanocarrier systems can  
533 encapsulate, protect, and/or regulate the release of the polyphenols, thereby enhancing their biostability and  
534 improving absorption and oral bioavailability (Nsairat et al., 2023). Furthermore, a variety of these nano-delivery  
535 systems are classified as biodegradable and biocompatible, and are reported to employ minimal nondrug additives  
536 to enhance the pharmacokinetics of their polyphenols payloads (Charoo et al., 2019). Additionally,  
537 nanotechnology can augment the biological and antioxidant activity of many polyphenols and prolong their half-  
538 lives by shielding them from physical, biological, and chemical destructive factors (Dini, 2022). Moreover,  
539 nanotechnology can be engineered to enable encapsulated polyphenols to evade first-pass metabolism by the  
540 gastrointestinal tract and liver enzymes, thereby increasing their bioavailability. However, encapsulating  
541 polyphenols into a novel delivery system may incur expenses and result in an unusual appearance and unpleasant  
542 odour, potentially impacting consumer demand (Puri et al., 2022).

543 Among all the available nanotechnologies, Layer-by-Layer assembly, an environmentally friendly process based  
544 on the electrostatic interactions between polyelectrolytes (where *Theobroma*-extracted pectin can act as  
545 polyanion), allows to create functionalised multilayered nanocoated carriers that can timely release in a controlled  
546 manner the bioactive compounds (Desmond et al., 2024). Due to the high versatility of the Layer-by-Layer  
547 assembly, the polyphenols can be loaded into the core as well as within the multilayered coating forming the shell  
548 of the nanocarrier.

549 An area ready for exploration lies in valorising the biomass resulting from fruit processing. Notably, the shells of  
550 *T. bicolor* and *T. grandiflorum* represent over 40% of the fresh fruit weight. Drawing from existing literature on  
551 *T. cacao* utilisation, embracing circular economy principles could facilitate the sustainable utilisation of other  
552 *Theobroma* species that normally, after the extraction of beans and pulps, produce a large volume of biowaste rich  
553 in bioactive compounds. Thus, this can serve as a new source suitable for therapeutic applications. Furthermore,  
554 the intensification of the production of promising *Theobroma* species, particularly *T. grandiflorum* and *T. bicolor*,  
555 presents various advantages, including economic benefits. Diversification of income for farmers is a significant  
556 positive outcome, reducing their dependency on traditional crops such as cocoa. In terms of agricultural benefits,  
557 integrating these species into agroforestry systems promotes biodiversity and sustainable land use. However, there  
558 are also environmental limitations associated with this intensification, including the risk of land use changes, such  
559 as deforestation, if not managed sustainably. Finally, social and cultural factors play a crucial role. Smallholder  
560 farmers may be hesitant to adopt new crops due to unfamiliarity and risk aversion. Effective knowledge transfer  
561 regarding cultivation techniques and the benefits of these crops to local communities is essential and can present  
562 a significant barrier.

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### 566 **CRedit authorship contribution statement**

567 **Maria Benloch-Tinoco:** Writing – original draft, Methodology, Investigation, Formal analysis, Data curation,  
568 Conceptualisation. **Jose Manuel Nuñez Ramírez:** Writing – review & editing. **Paola García:** Writing – review  
569 & editing. **Piergiorgio Gentile:** Writing – review & editing, Supervision, Resources. **Joel Girón-Hernández:**  
570 Writing – review & editing, Supervision, Resources.

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572 The authors declare that they have no known competing financial interests or personal relationships that could  
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### 574 **Declaration of generative AI and AI-assisted technologies in the writing process**

575 During the preparation of this work, the author(s) utilised ChatGPT 3.5 for proof-editing purposes. Following the  
576 use of this tool/service, the author(s) thoroughly reviewed and edited the content as necessary and take full  
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**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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