

Biomechanical measures of short-term maximal cycling on an ergometer: a test-retest study

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1 **Biomechanical measures of short-term maximal cycling on an** 2 **ergometer: a test-retest study**

3 An understanding of test-retest reliability is important for biomechanists, such
4 as when assessing the longitudinal effect of training or equipment interventions.
5 Our aim was to quantify the test-retest reliability of biomechanical variables
6 measured during short-term maximal cycling. Fourteen track sprint cyclists
7 performed 3 x 4 s seated sprints at 135 rpm on an isokinetic ergometer,
8 repeating the session 7.6 ± 2.5 days later. Joint moments were calculated via
9 inverse dynamics, using pedal forces and limb kinematics. EMG activity was
10 measured for 9 lower limb muscles. Reliability was explored by quantifying
11 systematic and random differences within- and between-session. Within-session
12 reliability was better than between-sessions reliability. The test-retest reliability
13 level was typically moderate to excellent for the biomechanical variables that
14 describe maximal cycling. However, some variables, such as peak knee flexion
15 moment and maximum hip joint power, demonstrated lower reliability,
16 indicating that care needs to be taken when using these variables to evaluate
17 biomechanical changes. Although measurement error (instrumentation error,
18 anatomical marker misplacement, soft tissue artefacts) can explain some of our
19 reliability observations, we speculate that biological variability may also be a
20 contributor to the lower repeatability observed in several variables including
21 ineffective crank force, ankle kinematics and hamstring muscles' activation
22 patterns.

23 **Keywords:** sprint cycling, kinematics, kinetics, emg, maximal power.

24 **Introduction**

25 The reliability of a clinical or sports science test is defined as the consistency or
26 reproducibility of a performance when a test is performed repeatedly (Hopkins, Schabort, &
27 Hawley, 2001). This is an important consideration for researchers, clinicians and applied

28 sports scientists as the better the reliability of the measurement the easier it is to detect a real
29 change in outcome (Hopkins, 2000). If the reliability of a test is low, then the outcome of a
30 test may conceal the true effect of an intervention. Conversely, if the reliability of a test is not
31 known then small random deviations may be misinterpreted as a meaningful change in
32 performance (Yavuzer, Öken, Elhan, & Stam, 2008).

33 Applied biomechanics researchers are often interested in assessing the short- or long-term
34 effects of interventions that aim to improve clinical or sports performance outcomes. In
35 clinical gait analysis, for example, the results of biomechanical assessments are used to
36 inform clinical decision making, by evaluating the effectiveness of interventions such as
37 surgery, physical therapy, medication or orthotics on gait biomechanics (Kadaba et al., 1989;
38 McGinley, Baker, Wolfe, & Morris, 2009; Yavuzer et al., 2008). Test-retest reliability studies
39 of clinical gait have found that the sagittal plane kinematics and kinetics have high values of
40 reliability in comparison to the data collected in the transverse and coronal planes (McGinley
41 et al., 2009). Furthermore, knee abduction/adduction and hip, knee and foot rotation joint
42 angles demonstrate the lowest reliability (McGinley et al., 2009), with the size of the
43 measurement error the same order of magnitude as the real joint motion in these planes. In
44 the context of clinical gait therefore, reliability studies have proved valuable by identifying
45 those variables that need to be interpreted with particular caution in order to effectively
46 inform clinical decision making (McGinley et al., 2009).

47 An understanding of test-retest reliability has similar relevance when assessing sporting
48 movements, as biomechanical measures are often used to evaluate the effectiveness of
49 longitudinal interventions such as changes to training programmes or equipment modification
50 (Costa, Bragada, Marinho, Silva, & Barbosa, 2012; Milner, Westlake, & Tate, 2011). Cycling
51 is a commonly used sporting movement for this purpose, as it is a relatively constrained

52 movement that can be accurately manipulated (Neptune, Kautz, & Hull, 1997; Neptune &
53 Kautz, 2001). Whilst the reliability of submaximal or “endurance” cycling is well reported
54 (Bini & Hume, 2013; Hopkins et al., 2001; Jobson, Hopker, Arkesteijn, & Passfield, 2013;
55 Laplaud, Hug, & Grélot, 2006), only a small amount by comparison is known about the
56 reliability of short-term maximal cycling. This comparative deficit exists despite maximal
57 cycling being an important paradigm for studying physiological capacity (Coso & Mora-
58 Rodríguez, 2006), muscle coordination and motor control strategies, as well as having direct
59 relevance to a range of competitive cycling performance environments (Martin, Davidson, &
60 Pardyjak, 2007). Therefore, quantifying test-retest reliability in maximal cycling
61 biomechanics is important. Test-retest reliability has been quantified for overall net crank
62 power output on an inertial load cycling ergometer within- and between-session (Coso &
63 Mora-Rodríguez, 2006; Hopkins et al., 2001; Mendez-Villanueva, Bishop, & Hamer, 2007),
64 with trained cyclists producing reliable power within the first testing session (Martin,
65 Diedrich, & Coyle, 2000). These studies demonstrated within-session reliability was better
66 than between-sessions reliability for overall net crank power output (Coso & Mora-
67 Rodríguez, 2006; Martin et al., 2000). There have been no studies quantifying the within- and
68 between-session reliability of biomechanical variables (crank power and forces, joint angles,
69 angular velocities, moments and powers and EMG activity) for short-term maximal cycling
70 despite these measures being important descriptors of the outcome, technique and
71 intermuscular coordination of a movement (Brochner Nielsen et al., 2018; Jacobs & van
72 Ingen Schenau, 1992; Wakeling, Blake, & Chan, 2010). EMG activity can be used to
73 determine muscle activation onset and offset times and level of activation (Dorel, Guilhem,
74 Couturier, & Hug, 2012; Hug & Dorel, 2009). This is important when investigating
75 intermuscular coordination in cycling as the timing and magnitude of muscle activation has to
76 be coordinated appropriately to allow an efficient energy transfer from the muscles though

77 the body segments to the pedal (Neptune & Kautz, 2001; Raasch, Zajac, Ma, & Levine,
78 1997). Joint kinetic measures (moments and powers) at the hip, knee and ankle throughout
79 the pedal revolution describe the action and contribution of the joints to pedal power and can
80 be used to identify different coordination strategies between cyclists (Elmer, Barratt, Korff, &
81 Martin, 2011; Martin & Brown, 2009; McDaniel, Behjani, Brown, & Martin, 2014).
82 Combining information on muscle activation from EMG and joint kinetics from inverse
83 dynamics analysis provides a deeper understanding of the joint and muscle actions that
84 produce the movement, and hence both are required to describe intermuscular coordination in
85 maximal cycling and were chosen for measurement and analysis during maximal cycling
86 (Brochner Nielsen et al., 2018; Dorel, 2018).

87 The aim of this study was to quantify the test-retest reliability of kinematic, kinetic, and
88 muscle activation variables during maximal sprint cycling. We hypothesise that within-
89 session reliability would be better than between-sessions reliability.

90 **Methods**

91 *Participants*

92 Fourteen track sprint cyclists participated in the study. Participants regularly competed at
93 track cycling competitions at either Master's international and national level (10), or Junior
94 national level (4). Although the participants were varied in their anthropometrics (7 males
95 and 7 females, age: 40.5 ± 17.7 yr, body mass: 72.5 ± 8.5 kg, height: 1.71 ± 0.06 m,), they
96 were similar with respect to cycling performance level (flying 200 m personal best: $11.98 \pm$
97 0.90 s). Participants were provided with study details and gave written informed consent. The
98 study was approved by the Sheffield Hallam University Faculty of Health and Wellbeing
99 Research Ethics Sub-Committee.

100 ***Experimental protocol***

101 An isokinetic ergometer was set up to replicate each participant's track bicycle position. All
102 participants' crank lengths were set to 165 mm, which was what they rode on their track
103 bicycles. Riders undertook their typical warm-up on the ergometer at self-selected pedalling
104 rate and resistance for at least 10 minutes, followed by one familiarisation sprint (4 s at 135
105 rpm). Martin and colleagues demonstrated that trained cyclists can produce valid and reliable
106 results for maximal cycling power from the first testing session (Martin et al., 2000),
107 therefore one familiarisation sprint was deemed appropriate. Riders then conducted 3 x 4 s
108 seated sprints at a pedalling rate of 135 rpm on the isokinetic ergometer with 4 minutes
109 recovery between efforts. Participants undertook an identical session 7.6 ± 2.5 days apart, at
110 approximately the same time of day (0.11 ± 2.18 h). A pedalling rate of 135 rpm was chosen
111 as this is a typical pedalling rate during the flying 200 m event in track cycling and within the
112 optimal pedalling rate range for track sprint cyclists (Dorel et al., 2005). The competitive
113 level and typical training volume of our participants meant that it was not feasible to ask them
114 to stop exercising 24 hours prior to the testing sessions, so instead they were instructed to
115 undertake the same training in the preceding 24 hours before both sessions.

116 ***Isokinetic ergometer***

117 A SRM Ergometer (Julich, Germany) cycle ergometer frame and flywheel were used to
118 construct an isokinetic ergometer. The modified ergometer flywheel was driven by a 2.2-kW
119 AC induction motor (ABB Ltd, Warrington, UK). The motor was controlled by a frequency
120 inverter equipped with a braking resistor (Model: Altivar ATV312 HU22, Schneider Electric
121 Ltd, London, UK). This set-up enabled the participants to start their bouts at the target
122 pedalling rate, rather than expending energy in accelerating the flywheel. The ergometer was
123 fitted with Sensix force pedals (Model ICS4, Sensix, Poitiers, France) and a crank encoder

124 (Model LM13, RLS, Komenda, Slovenia), sampling data at 200 Hz. Normal and tangential
125 pedal forces were resolved using the crank and pedal angles into the effective (propulsive)
126 and ineffective (applied along the crank) crank forces (Figure 1).

127 ***Kinematic and Kinetic Data Acquisition***

128 Two-dimensional kinematic data of the participants' left side were recorded at 100 Hz using
129 one high speed camera with infra-red ring lights (Model: UI-522xRE-M, IDS, Obersulm,
130 Germany). The camera was perpendicular to the participant, centred on the ergometer and set
131 about 3 m from the ergometer. The camera was in a very similar position for both sessions.
132 Reflective markers were placed on the pedal spindle, lateral malleolus, lateral femoral
133 condyle, greater trochanter and iliac crest. The same researcher attached the markers for all
134 sessions. Kinematics and kinetics on the ergometer were recorded by CrankCam software
135 (Centre for Sports Engineering Research, SHU, Sheffield, UK), which synchronised the
136 camera and pedal force data (down sampled to 100 Hz to match the camera data) and was
137 used for data processing, including auto-tracking of the marker positions.

138 ***EMG Data Acquisition***

139 EMG signals were recorded continuously from nine muscles of the left leg: vastus lateralis
140 (VL), rectus femoris (RF), vastus medialis (VM), tibialis anterior (TA), long head of biceps
141 femoris (BF), semitendinosus (ST), lateralis gastrocnemius (GL), soleus (SO), and gluteus
142 maximus (GMAX) with Delsys Trigno wireless surface EMG sensors (Delsys Inc, Boston,
143 MA). The skin at electrode placement sites was prepared by shaving the area then cleaning it
144 with an alcohol wipe. The EMG sensors were then placed in the centre of the muscle belly -
145 with the bar electrodes perpendicular to the muscle fibre orientation, using the guidelines in
146 (Konrad, 2005) and secured using wraps to reduce motion artefacts during pedalling. The

147 same researcher attached the EMG sensors for all sessions. A Delsys wireless sensor
148 containing an accelerometer (148 Hz sampling rate) was attached to the left crank arm to
149 obtain a measure of crank angle synchronised with the EMG signals. The EMG system was
150 operated and recorded in EMGworks Acquisition software (Delsys Inc, Boston, MA),
151 sampling data at 1926 Hz. The Delsys trigno EMG system automatically applied a bandwidth
152 filter of 20 ± 5 Hz to 450 ± 50 Hz (>80 dB/dec) to the raw signals.

153 *Data Processing*

154 All kinetic and kinematic data were filtered using a Butterworth fourth order (zero-lag) low
155 pass filter with a cut off frequency of 14 Hz selected using residual analysis (Winter, 2009).
156 The same cut off frequency was chosen for the kinematic and kinetic data as recommended
157 by Bezodis and colleagues to avoid data processing artefacts in the calculated joint moments
158 (Bezodis, Salo, & Trewartha, 2013). Instantaneous crank power was calculated from the
159 product of the left crank torque and the crank angular velocity. The average left side crank
160 power was calculated by averaging the instantaneous crank power over a complete pedal
161 revolution. Owing to a technical fault with the force measurement in the right pedal, it was
162 not possible to calculate total average crank power per revolution (sum of left and right crank
163 powers). Joint angles were calculated using the convention shown in Figure 1. Joint moments
164 were calculated via inverse dynamics (Elftman, 1939), using pedal forces, limb kinematics,
165 and body segment parameters (de Leva, 1996). Joint extension moments were defined as
166 positive and joint flexion moments as negative. The joint moments are presented from the
167 internal perspective (Derrick et al., 2020). Joint powers at the ankle, knee and hip were
168 determined by taking the product of the net joint moment and joint angular velocity.

169 Insert Figure 1

170 Data were analysed using a custom Matlab (R2017a, MathWorks, Cambridge, UK) script.
171 Each sprint lasted for 4 s providing six complete crank revolutions which were resampled to
172 100 data points around the crank cycle. Crank forces and powers, joint angles, angular
173 velocities, moments and powers were averaged over these revolutions to obtain a single
174 ensemble-averaged time series for each trial.

175 The accelerometer data for the crank arm was filtered using a Butterworth fourth order low
176 pass filter with a cut off frequency of 10 Hz. The minimum value of the acceleration of the
177 sensor in the direction of the crank arm corresponded to top dead centre (TDC) crank
178 position. To synchronise the EMG data with the kinematic and kinetic data, the TDC
179 locations from the accelerometer on the crank arm were matched to the corresponding TDC
180 measured by the crank encoder.

181 The raw EMG signals for the sprint efforts were high pass filtered (Butterworth second order,
182 cut off frequency 30 Hz) to diminish motion artefacts (De Luca, Gilmore, Kuznetsov, & Roy,
183 2010), root mean squared (RMS, 25 ms window) and then low pass filtered (Butterworth
184 second order, cut off frequency 24 Hz) (Brochner Nielsen et al., 2018). The data were then
185 interpolated to 100 data points around the crank cycle and then averaged over 6 crank
186 revolutions to create a linear envelope for each muscle. The EMG signals were normalised to
187 the mean value in the linear envelope across the crank cycle for each muscle.

188 *Statistical Analysis*

189 In order to test for any systematic change in performance between-sessions (for example, due
190 to learning or fatigue effects) paired *t*-tests were used to compare differences between
191 discrete values. Paired *t*-tests only test if there is a statistically significant bias between-
192 sessions (systematic change) but provide no indication of the random error due to biological

193 or mechanical variation between-sessions (Atkinson & Nevill, 1998). Similarly, differences
194 in time series data (instantaneous crank powers, crank forces, joint angles, angular velocities,
195 moments, powers and normalised EMG linear envelopes) between-sessions were assessed
196 using Statistical Parametric Mapping (SPM); paired *t*-tests were used for all variables except
197 crank forces where Hotelling's paired T^2 test was used (Pataky, 2010). Crank force consists
198 of two vector components (effective and ineffective crank force), therefore a multivariate
199 statistical test was required (Pataky, 2010). The level of statistical significance was set to $p <$
200 0.05 for all tests.

201 The reliability of the discrete variables between sessions was assessed using intra-class
202 correlation coefficient (ICC) tests. ICC's were calculated using IBM SPSS Statistics Version
203 24 (IBM UK Ltd, Portsmouth, UK), based on average measures, absolute agreement, two-
204 way mixed effects model (ICC (3,*k*) - where *k* is equal to the number of trials in a session
205 which in this study is three). The ICCs were interpreted using Koo and Li's guidelines: values
206 less than 0.50 are indicative of poor reliability, between 0.50 and 0.75 indicates moderate
207 reliability, 0.75 to 0.90 indicates good reliability and > 0.90 indicates excellent reliability
208 (Koo & Li, 2016). For a variable to be considered as having excellent reliability, both upper
209 and lower bounds of the 95% confidence intervals must fall within the excellent range (i.e. $>$
210 0.9) (Koo & Li, 2016).

211 Standard error of measurement (SEM) for between sessions was calculated using the formula
212 (Weir, 2005), where SD is standard deviation of the mean difference:

$$213 \quad SEM = SD\sqrt{1 - ICC}$$

214 Minimal detectable difference (MDD) was calculated for between sessions using the formula
215 (Weir, 2005):

$$MDD = SEM \times 1.96 \times \sqrt{2}$$

216 The coefficient of variation (CV) was calculated for the average crank power over a complete
217 revolution (Hopkins, 2000).

218 The standard error of measurement (SEM) was calculated for the kinematic and kinetic time
219 series data to evaluate the reliability of these waveforms within- and between-session using
220 the methods described in Pini, Markström, & Schelin, 2019. The mean and SD SEM for a
221 complete revolution was calculated for each variable. The EMG data were visually inspected
222 for signal quality and the frequency spectrum of the raw and filtered EMG signal calculated.
223 EMG signals with a high frequency content below 20 Hz, indicates low frequency noise due
224 to movement artefact (De Luca et al., 2010) and therefore, these trials were discarded. The
225 SEM for within- and between-session for the EMG linear envelopes of the VL, VM, ST, and
226 GMAX muscles were calculated using 13 participants. At least 2 trials for each muscle per
227 session per participant were required to calculate SEM. The calculated reliability of the EMG
228 data is therefore the upper bound, as very noisy trials were discarded.

229 The cross-correlation coefficient (R) was calculated to compare the temporal effects of
230 within- and between-session EMG linear envelopes (Wren, Do, Rethlefsen, & Healy, 2006).

231 The between-sessions cross-correlation coefficient was calculated comparing the session
232 mean EMG linear envelope, and within-session the cross-correlation coefficient was
233 calculated comparing the EMG linear envelope for two trials.

234 **Results**

235 *Discrete variables*

236 Discrete crank level variables demonstrated good to excellent between-sessions reliability
237 $ICC(3,k) > 0.756$ (Table 1). Average crank power for a complete revolution for the left side
238 only was 445.3 ± 95.7 and 438.8 ± 111.5 W for session 1 and 2 respectively (Table 1), which
239 gives an indicative total power for a complete revolution, for both cranks, of 891 and 878 W.
240 MDD between-sessions for peak crank power and forces was 21 W and between 9 to 72 N
241 respectively (Table 1). Peak joint angle values typically demonstrated moderate to excellent
242 reliability, with MDD between-sessions from 1.1 to 4.4° (Table 1). Peak joint angular
243 velocity between-sessions reliability was typically moderate to excellent, except for peak
244 knee flexion and hip extension angular velocity which had poor to good reliability (Table 1).
245 MDD between-sessions for peak joint angular velocities ranged from 14 to $59^\circ/s$ (Table 1).
246 Peak joint moments demonstrated moderate to excellent between-sessions reliability, except
247 for peak knee flexion moment which demonstrated poor to moderate reliability (Table 1).
248 Maximum ankle and knee joint powers demonstrated good to excellent reliability between-
249 sessions whereas, maximum hip power showed poor to good reliability (Table 1). MDD
250 between-sessions for peak joint moments ranged from 2 to 26 N.m and for maximum joint
251 powers 30 to 144 W.

252 Insert Table 1

253 CV for average crank power over a revolution was $3.0 \pm 1.5\%$ and $4.6 \pm 1.9\%$ for within- and
254 between-session respectively.

255 ***Time Series Variables***

256 Crank power demonstrated excellent within- and between-session reliability, with a mean
257 SEM between-sessions over a complete revolution of 46.6 ± 9.4 W (Figure 2, Figure 3).

258 Crank power was significantly different ($p < 0.05$) between sessions one and two, between
259 crank angles 340 to 6° (7.2% of crank cycle) (Figure 2). The ineffective crank force was less
260 repeatable (mean SEM = 31.6 ± 18.2 N) than effective crank force (mean SEM = 19.8 ± 4.0
261 N) within- and between-session, which was associated with a large SEM for ineffective crank
262 force between crank cycles of 140° and 210° (Figure 4, Figure 5). The crank forces were
263 significantly different ($p < 0.05$) between sessions one and two, between crank angles 191 to
264 199° (2.2% of crank cycle), and 347 and 1° (3.9% of crank cycle) (Figure 4).

265 Joint angles and angular velocities demonstrated excellent within- and between-session
266 reliability (mean SEM $\geq 2.4^\circ$ and $34.1^\circ/s$) (Figure 6). Ankle joint angles and angular
267 velocities were less repeatable than those at the knee and hip joints. Ankle joint angular
268 velocity was significantly different ($p < 0.05$) between sessions one and two, between crank
269 angles 152 to 170° (5.0% of crank cycle) (Figure 6).

270 Joint moments and powers demonstrated reasonable within- and between-session reliability
271 (mean SEM ≥ 15.5 N.m and 62.6 W) (Figure 6, Figure 7). Hip joint moments and powers
272 were less repeatable than those at the knee and ankle joints, particularly around the location
273 of maximum hip extension moment and power (Figure 7). Ankle joint moment was
274 significantly different ($p < 0.05$) between sessions one and two, between crank angles 340 to
275 6° (7.2% of crank cycle) (Figure 6). Hip joint power was significantly different ($p < 0.05$)
276 between session one and two between crank angles 340 to 2° (6.1% of crank cycle) (Figure
277 6).

278 EMG linear envelope normalised to the mean value in the signal demonstrated high within-
279 and between-session reliability (Figure 8). Mean SEM values for EMG linear envelopes
280 ranged between 0.14 to 0.16, and 0.16 to 0.20 proportion of the mean EMG signal, for
281 within- and between-session respectively. The GMAX, TA, and BF muscles demonstrated
282 the lowest reliability for EMG activity, and the VL and VM muscles the highest reliability
283 (Figure 8). The cross-correlation coefficient (R) which compares timing of EMG linear
284 envelopes between-sessions ranged from 0.976 to 0.990 (Figure 8).

285 Insert Figure 2, Figure 3, Figure 4, Figure 5, Figure 6, Figure 7, Figure 8

286 **Discussion and implications**

287 The purpose of this study was to quantify the test-retest reliability of kinematic, kinetic, and
288 EMG muscle activation variables measured during short-term maximal sprint cycling. Our
289 main findings were that between-sessions test-retest reliability level was typically moderate
290 to excellent for the biomechanical variables that describe maximal cycling, and furthermore
291 that within-session reliability was better than between-sessions reliability. However, some
292 variables, such as peak knee flexion moment and maximum hip joint power demonstrated
293 lower reliability, indicating that care needs to be taken when using these variables to evaluate
294 changes in maximal cycling biomechanics.

295 Within- and between-session values of SEM for joint angles and angular velocities
296 demonstrated high reliability (Figure 6). We found that ankle joint kinematics (angle and
297 angular velocity) were less repeatable than knee and hip joint kinematics, evidenced by the
298 larger mean SEM values for the ankle joint kinematics. The source of the lower reliability in
299 our ankle joint kinematics data is not clear, although it seems unlikely to be a measurement
300 error, given that anatomical landmark marker placement errors for the lower limb are greatest

301 at the hip, rather than the ankle joint (intra-examiner precision for the greater trochanter
302 marker is 12.2 mm along the long axis of the femur, and 11.1 mm in the anterior-posterior
303 direction, compared to lateral malleolus - 2.6 mm along the long axis fibula, 2.4 mm anterior-
304 posterior direction) (Della Croce, Cappozzo, & Kerrigan, 1999; Della Croce, Leardini,
305 Chiari, & Cappozzo, 2005). Furthermore, the soft tissue artefact (STA) of the lower limb
306 markers in cycling is also largest for the hip rather than the ankle joint (greater trochanter
307 marker displacement at 30 rpm submaximal cycling, 37.3 mm anterior-posterior and 10.3 mm
308 proximal-distal, compared to the lateral malleolus 15.8 mm anterior-posterior and 8.6 mm
309 proximal-distal) (Li et al., 2017). By comparison there are potential biological explanations
310 for the lower reliability of the ankle joint kinematics. Martin and Nichols, for example,
311 demonstrated that the ankle has a different role to the knee and hip joints in maximal cycling
312 and acts to transfer - instead of maximise power (Martin & Nichols, 2018). More specifically,
313 the ankle works in synergy with the hip joint to transfer power produced by the muscles
314 surrounding the hip joint to the crank (Fregly & Zajac, 1996). Our results support this notion
315 by suggesting that cyclists may regulate their ankle angle as part of this hip-ankle synergy, in
316 order to maintain a stable effective crank force. A specially designed experiment would be
317 required to test this hypothesis.

318 In terms of joint kinetics, joint moments and powers demonstrated lower reliability at more
319 proximal compared to distal joints – with the largest values of SEM for the hip joint moment
320 (Figure 6, Figure 7). This observation may be due to the STA and skin marker misplacement
321 errors being largest at the hip joint, as discussed above (Della Croce et al., 1999; Li et al.,
322 2017). It may also be due to the fact that measurement errors in general (STA, marker
323 misplacement, force pedal measurement precision) will propagate through the inverse
324 dynamics calculations (Myers, Laz, Shelburne, & Davidson, 2015). In either scenario, this

325 indicates that the observed differences in proximal to distal joint reliability are likely to be
326 due to measurement error, rather than biological variability.

327 The peak knee flexion moment showed poor to moderate between-sessions reliability, with
328 the largest MDD of all joint moments (26 N.m). Error due to knee marker misplacement is
329 dependent on knee flexion angle, with previous studies demonstrating that the greater the
330 knee flexion, the larger error in the joint angle (Della Croce et al., 1999). Marker
331 displacement could therefore explain the poor reliability of our peak knee flexion angular
332 velocity and moment data. Further work is required, using more detailed marker sets and
333 models of STA, to reduce the influence of STA and skin marker misplacement on the
334 calculated kinematics and kinetic variables, which may improve the reliability of the
335 calculated knee flexion and hip joint variables.

336 Average crank power output over a complete revolution was highly reliable both within- and
337 between-session, supporting the findings of Martin and colleagues that trained cyclists are
338 able to reproduce reliable maximal crank power within one testing session (Martin et al.,
339 2000). Effective crank force exhibited similar reliability to crank power, whereas ineffective
340 crank force demonstrated lower within- and between-session reliability which was associated
341 with the large intra-participant variability and SEM in ineffective crank force between crank
342 angles of 140° and 210° (Figure 4, Figure 5). It is unlikely that force pedals' measurement
343 precision would provide an explanation for these observed differences in reliability between
344 the effective and ineffective crank forces, given that the measurement precision values are the
345 same for all components of force for the instrumented pedals we used (combined error -
346 linearity and hysteresis 1% measuring range (MR) and crosstalk between the components
347 (<1.5% MR) (Sensix, Poitiers, France)). Therefore, it seems probable that the reliability

348 difference between effective and ineffective force may have a biological basis, a notion
349 which can be expanded upon using our EMG results.

350 EMG linear envelopes generally demonstrated excellent reliability (Figure 8). However, the
351 GMAX, BF and the TA muscles demonstrated the lowest reliability for EMG activity. Lower
352 reliability of the EMG activity for the GMAX and TA muscles have been demonstrated in
353 submaximal cycling (Jobson et al., 2013). The between-sessions reliability of the EMG
354 activity of the GMAX muscle has been shown to decrease with increasing workload
355 (between-sessions CV = 43.1% at 265 W compared to CV = 23.0 at 135 W) (Jobson et al.,
356 2013) which might suggest greater biological variation in the GMAX muscle activity with
357 increased workload, potentially explaining the lower reliability of the GMAX EMG activity.
358 Jobson and colleagues suggested the lower reliability of the EMG activity for the TA muscle
359 might be owing to the fact some cyclists have two bursts of muscle activity per crank
360 revolution which may introduce more between crank revolution variability (Jobson et al.,
361 2013). Measurement error could also be a potential source of the lower reliability of the EMG
362 activity for the TA, as the location of the EMG sensor can strongly influence the pattern of
363 EMG activity recorded owing to crosstalk from the peroneus longus muscle during dynamic
364 movements (Campanini et al., 2007; Hug, 2011). Therefore, small changes in positioning of
365 the EMG sensor between sessions could influence the EMG activity measured. Wren and
366 colleagues suggested the lower reliability of the hamstrings may be due to measurement error
367 reflecting the increased sensitivity of these muscles to electrode placement owing to muscle
368 length and overlying fat mass (Wren et al., 2006). The lower reliability of EMG activity in
369 the BF hamstring muscle may also have a biological basis however, given that our findings
370 are consistent with other studies who suggest that this is related to their bi-articular function
371 (Ryan & Gregor, 1992). Van Ingen Schenau and colleagues for example demonstrated that

372 the bi-articular muscles are important for controlling the direction of the external force on the
373 pedal (van Ingen Schenau, Boots, De Groot, Snackers, & Van Woensel, 1992). They
374 identified that the paradoxical coactivation of the mono-articular agonists (vastii) with bi-
375 articular antagonists (hamstrings) emerges so the bi-articular muscles can help control the
376 desired direction of the force applied to the pedal by adjusting the relative distribution of net
377 moments over the joints (van Ingen Schenau et al., 1992).

378 On a mechanical basis, the goal of maximal cycling is to maximise the effective crank force
379 as this maximises the propulsive power and thus the speed of the bicycle. Taking our crank
380 force and EMG data together therefore, our results allow us to speculate that cyclists may
381 regulate bi-articular muscles activation to control the direction of the pedal force, with the
382 aim of maximising effective crank force and maintaining a stable outcome at the expense of
383 the ineffective force which does not directly affect the task outcome. The bi-articular muscles
384 (BF, ST and GL) are active in the region of the crank cycle where the ineffective crank is
385 more variable which could explain the biological mechanism underlying this finding. This
386 principle has been observed in walking (Kadaba et al., 1989; Giakas & Baltzopoulos, 1997)
387 and running (Kinoshita, Bates, & DeVita, 1985), where the propulsion and braking ground
388 reaction forces (anterior-posterior and vertical direction) have been shown to have lower
389 between-stride variability than the medio-lateral force. However, further, purposefully
390 designed experiments are required to confirm or refute these speculations.

391 SPM indicated a significant between-session difference for small regions of the crank cycle,
392 for crank power, crank forces, ankle angular velocity and moment, and hip power. These
393 differences are unlikely to be meaningful changes as these are less than 7.2% of the crank
394 cycle, and typically occur in regions of low magnitude in these variables.

395 The experimental protocol could have introduced some variability to the kinematics, as
396 although the participants were instructed to remain seated during the sprints on the ergometer,
397 they tended to hover slightly over the saddle (potentially with the aim to increase crank
398 power), which increases pelvis movement. Also, the ergometer was set-up to match each
399 participant's track bike. Therefore, saddle height was not standardised to percentage of inside
400 leg length, which is often recommended (de Vey Mestdagh, 1998). Some of the participants
401 had a relatively low saddle height compared to their leg length, which resulted in relatively
402 large pelvis obliquity (rocking) and transverse rotation when they sprinted. This strategy may
403 have introduced more within- and between-trial variability, particularly at the hip joint. We
404 acknowledge that we measured 2D kinematics using a high-speed video camera, which is not
405 considered the 'gold standard' for measuring kinematics which is 3D motion capture systems
406 (Fonda, Sarabon, & Li, 2014). However, these methods were utilised because during cycling
407 the movement is predominantly in the sagittal plane (Umberger & Martin, 2001; van Ingen
408 Schenau, Van Woensel, Boots, Snackers, & De Groot, 1990) and therefore previous studies
409 that have investigated maximal cycling have just considered the sagittal plane actions, as this
410 is the plane where muscles produce power to generate effective crank force (Barratt, Korff,
411 Elmer, & Martin, 2011; Elmer et al., 2011; Martin & Brown, 2009; McDaniel et al., 2014).
412 Therefore, we measured 2D kinematics in the sagittal plane using a simple marker set which
413 has the added benefit of reducing time required for data collection sessions which is an
414 important ethical consideration when working with elite athletes.

415 **Conclusion**

416 Typically, the biomechanical variables that describe maximal cycling are reliable. However,
417 some variables have lower reliability indicating that care needs to be taken when using these
418 variables to evaluate changes in maximal cycling biomechanics. Our results allow us to

419 speculate that biological variability is the source of the lower reliability of the ineffective
420 crank force, ankle kinematics and hamstring muscles activation while measurement error is
421 the source of the lower reliability in hip and knee joint kinetics. Further research using
422 purposefully designed experiments is required to confirm or refute these speculations. We
423 recognise that there were some data collection problems (noisy EMG data and no right force
424 pedal data) which might indicate potentially lower reliability of our data collection method.
425 These reliability data can be used to help understand the practical relevance of a longitudinal
426 intervention on athletes' maximal cycling performance.

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429 implementing statistical parametric mapping.

430 **References**

- 431 Atkinson, G., & Nevill, A. M. (1998). Statistical methods for assessing measurement error
432 (reliability) in variables relevant to sports medicine. *Sports Medicine*, 26, 217-238.
- 433 Barratt, P., Korff, T., Elmer, S. J., & Martin, J. C. (2011). Effect of crank length on joint-
434 specific power during maximal cycling. *Medicine and Science in Sports and Exercise*, 43,
435 1689-1697.
- 436 Bezodis, N. E., Salo, A. I. T., & Trewartha, G. (2013). Excessive fluctuations in knee joint
437 moments during early stance in sprinting are caused by digital filtering procedures. *Gait &*
438 *Posture*, 38, 653-657.

439 Bini, R. R., & Hume, P. A. (2013). Between-day reliability of pedal forces for cyclists during
440 an incremental cycling test to exhaustion. *Isokinetics and Exercise Science*, *21*, 203-209.

441 Brochner Nielsen, N. P., Hug, F., Guevel, A., Colloud, F., Lardy, J., & Dorel, S. (2018).
442 Changes in motor coordination induced by local fatigue during a sprint cycling task.
443 *Medicine and Science in Sports and Exercise*, *50*, 1394-1404.

444 Campanini, I., Merlo, A., Degola, P., Merletti, R., Vezzosi, G., & Farina, D. (2007). Effect of
445 electrode location on EMG signal envelope in leg muscles during gait. *Journal of*
446 *Electromyography and Kinesiology*, *17*, 515-526.

447 Coso, J. D., & Mora-Rodríguez, R. (2006). Validity of cycling peak power as measured by a
448 short-sprint test versus the wingate anaerobic test. *Applied Physiology, Nutrition, and*
449 *Metabolism*, *31*, 186-189.

450 Costa, M. J., Bragada, J. A., Marinho, D. A., Silva, A. J., & Barbosa, T. M. (2012).
451 Longitudinal interventions in elite swimming: A systematic review based on energetics,
452 biomechanics, and performance. *The Journal of Strength & Conditioning Research*, *26*,
453 2006-2016.

454 de Leva, P. (1996). Adjustments to zatsiorsky-seluyanov's segment inertia parameters.
455 *Journal of Biomechanics*, *29*, 1223-1230.

456 De Luca, C. J., Gilmore, L. D., Kuznetsov, M., & Roy, S. H. (2010). Filtering the surface
457 EMG signal: Movement artifact and baseline noise contamination. *Journal of Biomechanics*,
458 *43*, 1573-1579.

459 de Vey Mestdagh, K. (1998). Personal perspective: In search of an optimum cycling posture.
460 *Applied Ergonomics*, 29, 325-334.

461 Della Croce, U., Cappozzo, A., & Kerrigan, D. C. (1999). Pelvis and lower limb anatomical
462 landmark calibration precision and its propagation to bone geometry and joint angles.
463 *Medical & Biological Engineering & Computing*, 37, 155-161.

464 Della Croce, U., Leardini, A., Chiari, L., & Cappozzo, A. (2005). Human movement analysis
465 using stereophotogrammetry: Part 4: Assessment of anatomical landmark misplacement and
466 its effects on joint kinematics. *Gait & Posture*, 21, 226-237.

467 Derrick, T. R., van den Bogert, Antonie J, Cereatti, A., Dumas, R., Fantozzi, S., & Leardini,
468 A. (2020). ISB recommendations on the reporting of intersegmental forces and moments
469 during human motion analysis. *Journal of Biomechanics*, 99, doi:
470 <https://doi.org/10.1016/j.jbiomech.2019.109533>

471 Dorel, S. (2018). Mechanical effectiveness and coordination: New insights into sprint cycling
472 performance. In J. Morin, & P. Samozino (Eds.), *Biomechanics of training and testing* (pp.
473 33-62). Switzerland: Springer.

474 Dorel, S., Guilhem, G., Couturier, A., & Hug, F. (2012). Adjustment of muscle coordination
475 during an all-out sprint cycling task. *Medicine and Science in Sports and Exercise*, 44, 2154-
476 2164.

477 Dorel, S., Hautier, C. A., Rambaud, O., Rouffet, D., Praagh, E. V., Lacour, J. R., & Bourdin,
478 M. (2005). Torque and power-velocity relationships in cycling: Relevance to track sprint
479 performance in world-class cyclists. *International Journal of Sports Medicine*, 26, 739-746.

480 Elftman, H. (1939). Forces and energy changes in the leg during walking. *American Journal*
481 *of Physiology*, 125, 339-356.

482 Elmer, S. J., Barratt, P., Korff, T., & Martin, J. C. (2011). Joint-specific power production
483 during submaximal and maximal cycling. *Medicine and Science in Sports and Exercise*, 43,
484 1940-1947.

485 Fonda, B., Sarabon, N., & Li, F. (2014). Validity and reliability of different kinematics
486 methods used for bike fitting. *Journal of Sports Sciences*, 32, 940-946.

487 Fregly, B. J., & Zajac, F. E. (1996). A state-space analysis of mechanical energy generation,
488 absorption, and transfer during pedaling. *Journal of Biomechanics*, 29, 81-90.

489 Giakas, G., & Baltzopoulos, V. (1997). Time and frequency domain analysis of ground
490 reaction forces during walking: An investigation of variability and symmetry. *Gait &*
491 *Posture*, 5, 189-197.

492 Hopkins, W. G. (2000). Measures of reliability in sports medicine and science. *Sports*
493 *Medicine*, 30, 1-15.

494 Hopkins, W. G., Schabert, E. J., & Hawley, J. A. (2001). Reliability of power in physical
495 performance tests. *Sports Medicine*, 31, 211-234.

496 Hug, F. (2011). Can muscle coordination be precisely studied by surface electromyography?
497 *Journal of Electromyography and Kinesiology*, 21, 1-12.

498 Hug, F., & Dorel, S. (2009). Electromyographic analysis of pedaling: A review. *Journal of*
499 *Electromyography and Kinesiology*, 19, 182-198.

500 Jacobs, R., & van Ingen Schenau, G. J. (1992). Intermuscular coordination in a sprint push-
501 off. *Journal of Biomechanics*, 25, 953-965.

502 Jobson, S. A., Hopker, J., Arkesteijn, M., & Passfield, L. (2013). Inter-and intra-session
503 reliability of muscle activity patterns during cycling. *Journal of Electromyography and*
504 *Kinesiology*, 23, 230-237.

505 Kadaba, M., Ramakrishnan, H., Wootten, M., Gaine, J., Gorton, G., & Cochran, G. (1989).
506 Repeatability of kinematic, kinetic, and electromyographic data in normal adult gait. *Journal*
507 *of Orthopaedic Research*, 7, 849-860.

508 Kinoshita, H., Bates, B., & DeVita, P. (1985). Intertrial variability for selected running gait
509 parameters. *Biomechanics IX-A* (pp. 499-502). Champaign, IL: Human Kinetics Publishers.

510 Konrad, P. (2005). *The ABC of EMG*. (No.1.4). Scottsdale: Noraxon INC.

511 Koo, T. K., & Li, M. Y. (2016). A guideline of selecting and reporting intraclass correlation
512 coefficients for reliability research. *Journal of Chiropractic Medicine*, 15, 155-163.

513 Laplaud, D., Hug, F., & Grélot, L. (2006). Reproducibility of eight lower limb muscles
514 activity level in the course of an incremental pedaling exercise. *Journal of Electromyography*
515 *and Kinesiology*, 16, 158-166.

516 Li, J., Lu, T., Lin, C., Kuo, M., Hsu, H., & Shen, W. (2017). Soft tissue artefacts of skin
517 markers on the lower limb during cycling: Effects of joint angles and pedal resistance.
518 *Journal of Biomechanics*, 62, 27-38.

519 Martin, J. C., Diedrich, D., & Coyle, E. F. (2000). Time course of learning to produce
520 maximum cycling power. *International Journal of Sports Medicine*, 21, 485-487.

521 Martin, J. C., & Brown, N. A. T. (2009). Joint-specific power production and fatigue during
522 maximal cycling. *Journal of Biomechanics*, *42*, 474-479.

523 Martin, J. C., Davidson, C., & Pardyjak, E. (2007). Understanding sprint-cycling
524 performance: The integration of muscle power, resistance, and modeling. *International*
525 *Journal of Sports Physiology and Performance*, *2*, 5-21.

526 Martin, J. C., & Nichols, J. (2018). Simulated work loops predict maximal human cycling
527 power. *The Journal of Experimental Biology*, *221*, doi: 10.1242/jeb.180109

528 McDaniel, J., Behjani, N. S. E., S.J., Brown, N. A. T., & Martin, J. C. (2014). Joint-specific
529 power-pedaling rate relationships during maximal cycling. *Journal of Applied Biomechanics*,
530 *30*, 423-430.

531 McGinley, J. L., Baker, R., Wolfe, R., & Morris, M. E. (2009). The reliability of three-
532 dimensional kinematic gait measurements: A systematic review. *Gait & Posture*, *29*, 360-
533 369.

534 Mendez-Villanueva, A., Bishop, D., & Hamer, P. (2007). Reproducibility of a 6-s maximal
535 cycling sprint test. *Journal of Science and Medicine in Sport*, *10*, 323-326.

536 Milner, C. E., Westlake, C. G., & Tate, J. J. (2011). Test–retest reliability of knee
537 biomechanics during stop jump landings. *Journal of Biomechanics*, *44*, 1814-1816.

538 Myers, C. A., Laz, P. J., Shelburne, K. B., & Davidson, B. S. (2015). A probabilistic
539 approach to quantify the impact of uncertainty propagation in musculoskeletal simulations.
540 *Annals of Biomedical Engineering*, *43*, 1098-1111.

541 Neptune, R. R., & Kautz, S. A. (2001). Muscle activation and deactivation dynamics: The
542 governing properties in fast cyclical human movement performance? *Exercise and Sport*
543 *Sciences Reviews*, 29, 76-81.

544 Neptune, R. R., Kautz, S. A., & Hull, M. L. (1997). The effect of pedaling rate on
545 coordination in cycling. *Journal of Biomechanics*, 30, 1051-1058.

546 Pataky, T. C. (2010). Generalized n-dimensional biomechanical field analysis using statistical
547 parametric mapping. *Journal of Biomechanics*, 43, 1976-1982.

548 Pini, A., Markström, J. L., & Schelin, L. (2019). Test–retest reliability measures for curve
549 data: An overview with recommendations and supplementary code. *Sports Biomechanics*,
550 doi: 10.1080/14763141.2019.1655089

551 Raasch, C. C., Zajac, F. E., Ma, B., & Levine, W. S. (1997). Muscle coordination of
552 maximum-speed pedaling. *Journal of Biomechanics*, 30, 595-602.

553 Ryan, M. M., & Gregor, R. J. (1992). EMG profiles of lower extremity muscles during
554 cycling at constant workload and cadence. *Journal of Electromyography and Kinesiology*, 2,
555 69-80.

556 Umberger, B. R., & Martin, P. E. (2001). Testing the planar assumption during ergometer
557 cycling. *Journal of Applied Biomechanics*, 17, 55-62.

558 van Ingen Schenau, G. J., Boots, P. J. M., De Groot, G., Snackers, R. J., & Van Woensel, W.
559 W. L. M. (1992). The constrained control of force and position in multi-joint movements.
560 *Neuroscience*, 46, 197-207.

561 van Ingen Schenau, G. J., Van Woensel, W. W. L. M., Boots, P. J. M., Snackers, R. W., & De
562 Groot, G. (1990). Determination and interpretation of mechanical power in human
563 movement: Application to ergometer cycling. *European Journal of Applied Physiology and*
564 *Occupational Physiology*, 61, 11-19.

565 Wakeling, J. M., Blake, O. M., & Chan, H. K. (2010). Muscle coordination is key to the
566 power output and mechanical efficiency of limb movements. *The Journal of Experimental*
567 *Biology*, 213, 487-492.

568 Weir, J. P. (2005). Quantifying test-retest reliability using the intraclass correlation
569 coefficient and the SEM. *The Journal of Strength & Conditioning Research*, 19, 231-240.

570 Winter, D. A. (2009). Biomechanics and motor control of human movement. (4th ed., pp. 70-
571 73). Hoboken, NJ: Wiley.

572 Wren, T. A., Do, K. P., Rethlefsen, S. A., & Healy, B. (2006). Cross-correlation as a method
573 for comparing dynamic electromyography signals during gait. *Journal of Biomechanics*, 39,
574 2714-2718.

575 Yavuzer, G., Öken, Ö, Elhan, A., & Stam, H. J. (2008). Repeatability of lower limb three-
576 dimensional kinematics in patients with stroke. *Gait & Posture*, 27, 31-35.

577

Table 1: Between-sessions reliability for kinematic and kinetic variables, * indicates significant difference between sessions ($p < 0.05$), ICC(3,k) = Between-sessions intraclass correlation with lower (LB) and upper (UB) bound confidence intervals, SEM = standard error of measurement, MDD = minimal detectable difference

Variable	Units	Mean(SD)	Mean difference	<i>p</i>	ICC	95%	95%	SEM	MDD
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		Session 1	Session 2		(3,k)	LB	UB			
Power (average for left crank)	W	445.3 ± 95.7	438.8 ± 111.5	-6.5	0.429	0.979	0.938	0.993	4.3	12
Peddalling rate	rpm	134.8 ± 1.3	134.7 ± 1.4	-0.2	0.021*	0.986	0.935	0.996	0.0	0.1
Max effective crank force	N	593.3 ± 126.2	579.0 ± 130.9	-14.4	0.072	0.986	0.952	0.996	3.2	9
Max ineffective crank force	N	603.5 ± 172.1	605.3 ± 165.4	1.8	0.944	0.923	0.756	0.975	25.9	72
Min ineffective crank force	N	-192.7 ± 65.2	-207.3 ± 82.3	-14.7	0.136	0.937	0.805	0.980	8.7	24
Max instantaneous crank power	W	1387.2 ± 309.2	1348.4 ± 316.5	-38.7	0.043*	0.986	0.946	0.996	7.7	21
Peak ankle plantarflexion angle	°	141.7 ± 11.3	142.3 ± 11.5	0.6	0.446	0.983	0.948	0.994	0.4	1.1
Peak ankle dorsiflexion angle	°	113.1 ± 5.0	113.8 ± 5.8	0.7	0.281	0.955	0.863	0.985	0.5	1.3
Peak knee extension angle	°	142.7 ± 6.4	143.5 ± 5.7	0.8	0.489	0.864	0.580	0.956	1.6	4.4
Peak knee flexion angle	°	70.0 ± 3.6	70.2 ± 3.4	0.2	0.715	0.857	0.550	0.954	1.0	2.6
Peak hip extension angle	°	68.1 ± 5.0	68.4 ± 4.6	0.3	0.720	0.893	0.665	0.966	1.0	2.8
Peak hip flexion angle	°	26.1 ± 4.3	25.6 ± 4.2	-0.5	0.447	0.916	0.746	0.973	0.7	1.9
Peak ankle plantarflexion angular velocity	°/s	236.6 ± 65.7	247.1 ± 65.0	10.4	0.441	0.839	0.509	0.948	19.7	55
Peak ankle dorsiflexion angular velocity	°/s	-262.0 ± 91.2	-268.5 ± 107.2	-6.6	0.561	0.957	0.868	0.986	8.6	24
Peak knee extension angular velocity	°/s	472.8 ± 43.2	479.1 ± 33.8	6.3	0.434	0.838	0.504	0.948	11.8	33
Peak knee flexion angular velocity	°/s	-507.5 ± 57.6	-513.3 ± 43.6	-5.8	0.635	0.772	0.279	0.927	21.4	59
Peak hip extension angular velocity	°/s	265.6 ± 29.1	273.8 ± 21.9	8.2	0.141	0.814	0.447	0.939	8.5	24
Peak hip flexion angular velocity	°/s	-277.6 ± 30.7	-273.4 ± 35.1	4.2	0.390	0.924	0.769	0.975	4.9	14
Peak ankle plantarflexion moment	N.m	78.6 ± 18.6	81.4 ± 20.2	2.8	0.372	0.910	0.729	0.971	3.4	9
Peak ankle dorsiflexion moment	N.m	-14.0 ± 7.0	-12.3 ± 6.0	1.8	0.049*	0.928	0.743	0.978	0.8	2
Peak knee extension moment	N.m	90.0 ± 34.5	82.9 ± 33.5	-7.1	0.028*	0.965	0.852	0.990	2.0	6
Peak knee flexion moment	N.m	-50.7 ± 20.9	-57.7 ± 15.0	-7.0	0.151	0.697	0.127	0.900	9.4	26
Peak hip extension moment	N.m	132.3 ± 30.7	140.4 ± 32.8	8.1	0.086	0.919	0.737	0.974	4.6	13
Peak hip flexion moment	N.m	-47.7 ± 26.1	-41.3 ± 17.0	6.5	0.115	0.870	0.600	0.958	5.1	14
Maximum ankle power	W	259.6 ± 111.7	258.5 ± 107.8	-1.1	0.937	0.951	0.846	0.984	10.9	30
Maximum knee power	W	659.6 ± 321.7	620.4 ± 253.6	-39.2	0.160	0.968	0.901	0.990	17.6	49
Maximum hip power	W	519.8 ± 186.3	578.1 ± 153.0	58.3	0.104	0.826	0.474	0.944	52.1	144

Figure captions

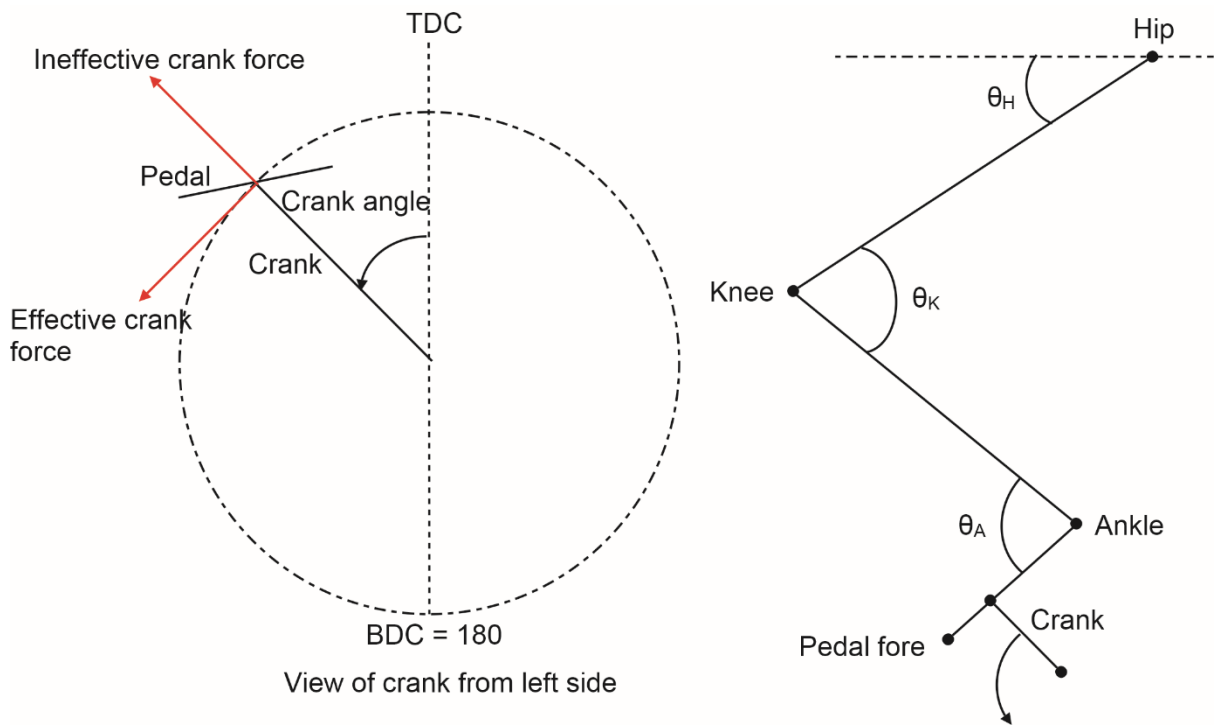


Figure 1: Joint angle and crank forces convention. TDC = top dead centre, BDC = bottom dead centre, θ_H = hip angle, θ_K = knee angle, θ_A = ankle angle

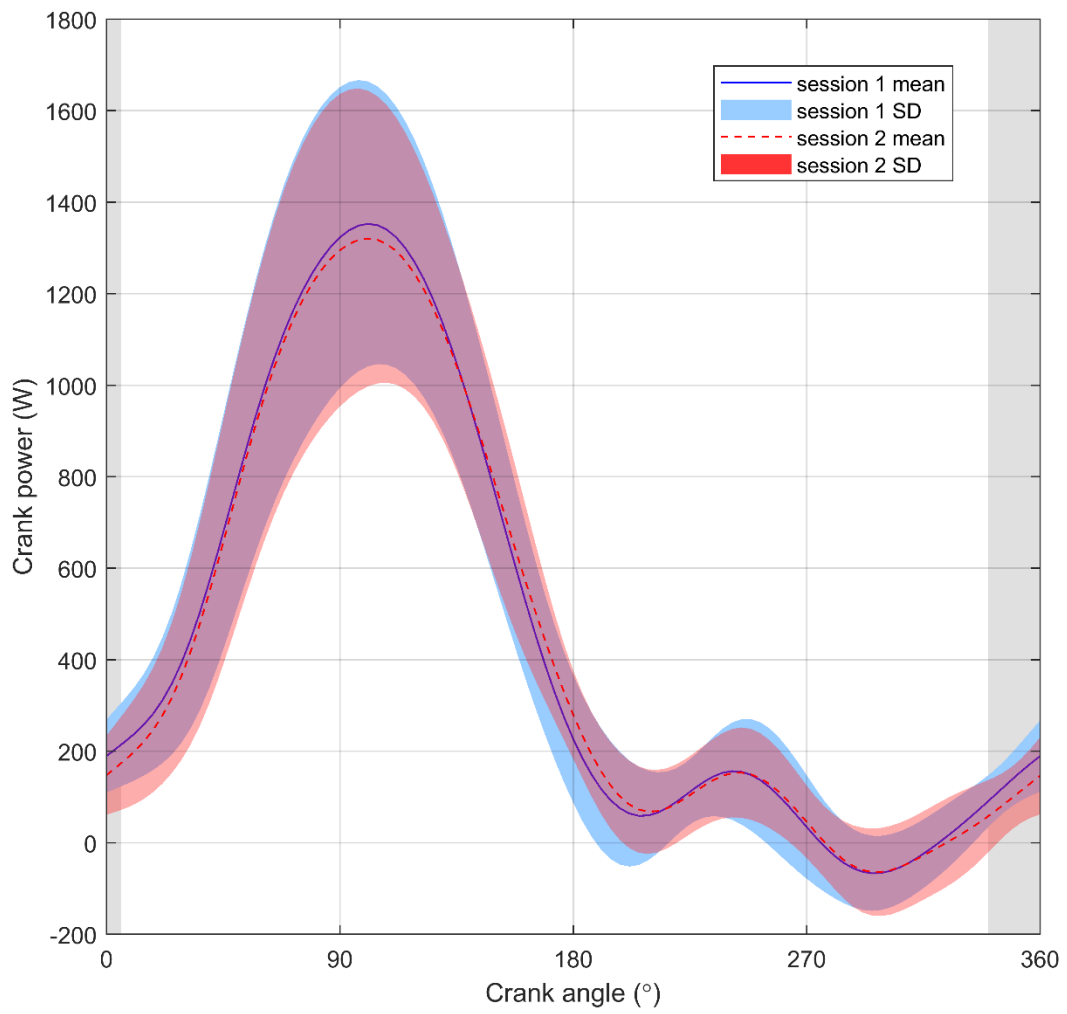


Figure 2: Crank power: group means for session one and two. Areas of the graph shaded grey where the Statistical parametric mapping (SPM) is significant.

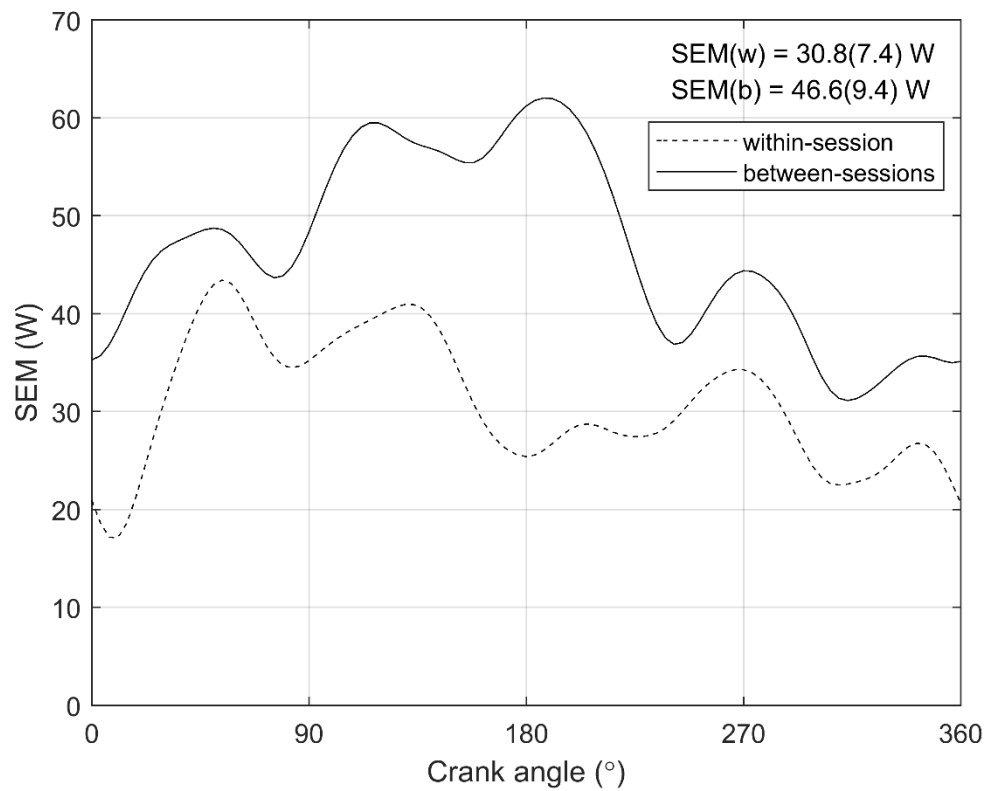


Figure 3: Crank power: standard error of measurement (SEM) within- and between-session. Mean and standard deviation of SEM within-session (w) and between-sessions (b) over complete crank cycle.

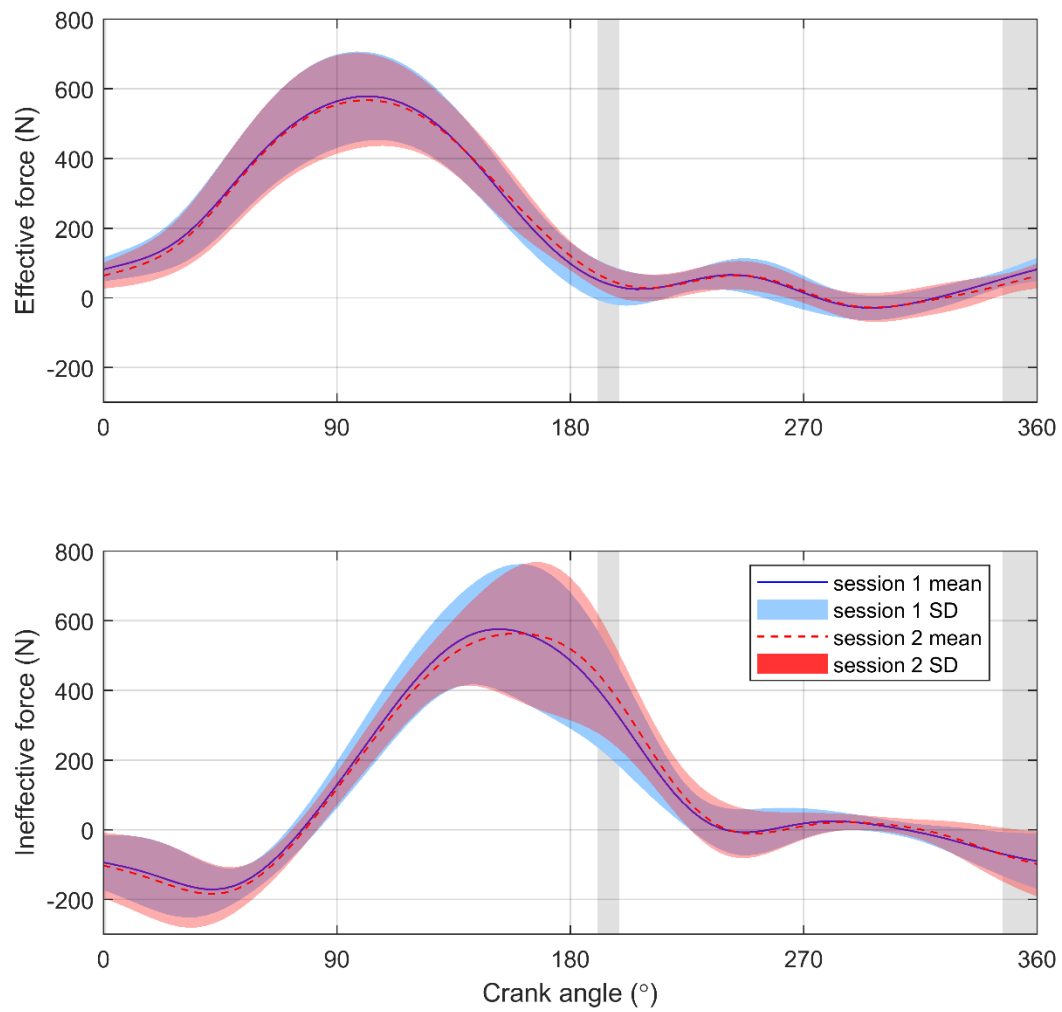


Figure 4: Crank forces: group means for session one and two. Areas of the graph shaded grey where the Statistical parametric mapping (SPM) is significant.

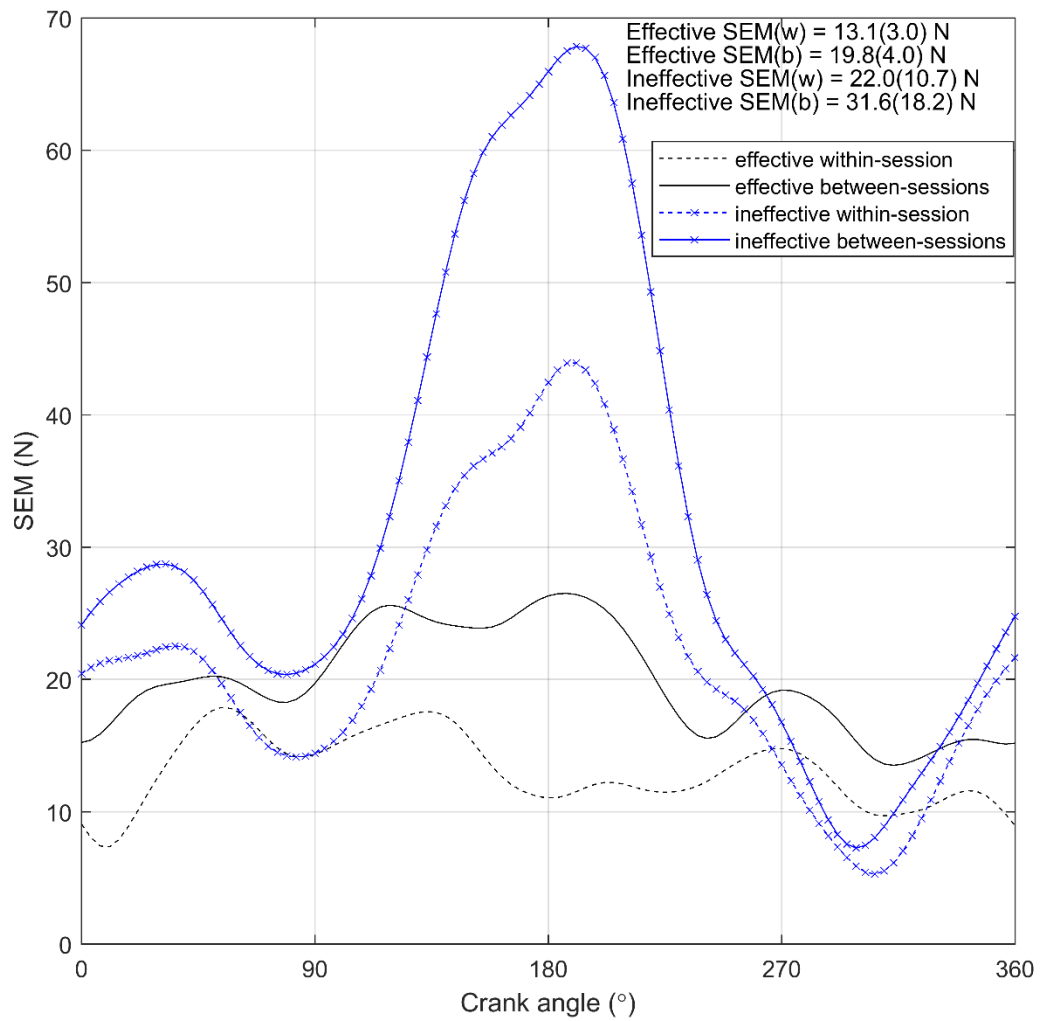


Figure 5: Crank forces: standard error of measurement (SEM) within- and between-session. Mean and standard deviation of SEM within-session (w) and between-sessions (b) over complete crank cycle.

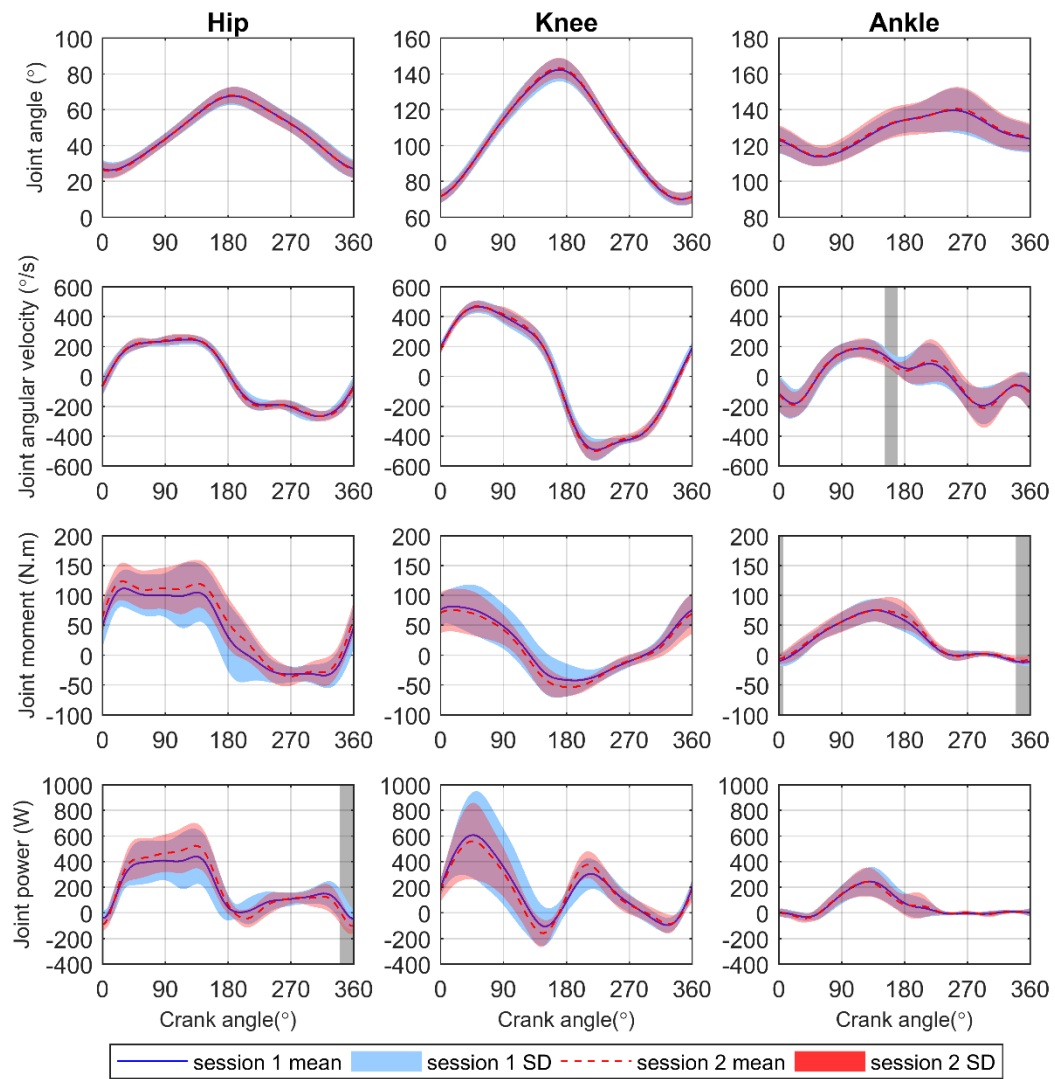


Figure 6: Joint angles, angular velocities, moments and powers: group means for session one and two. Areas of the graph shaded grey where the Statistical parametric mapping (SPM) is significant.

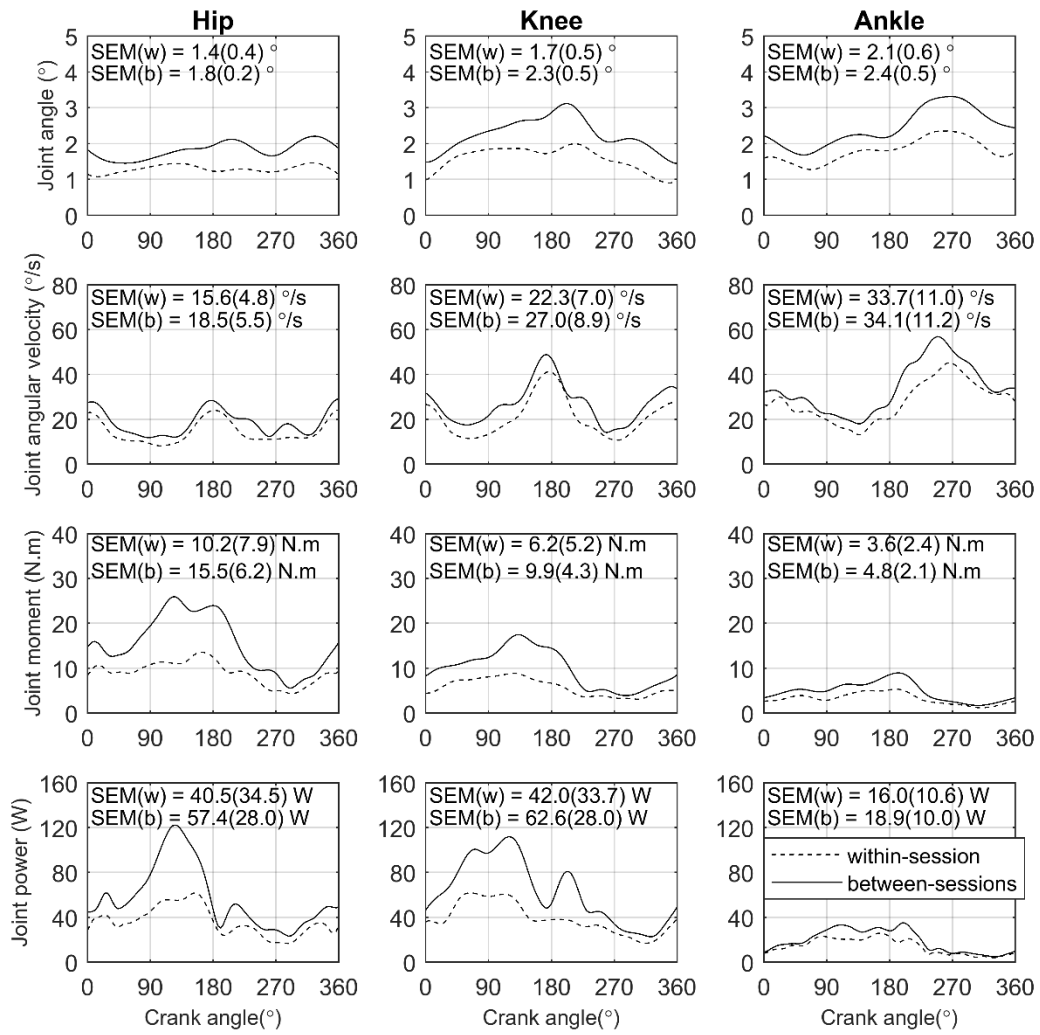


Figure 7: Joint angles, angular velocities, moments and powers: standard error of measurement (SEM) within- and between-session. Mean and standard deviation of SEM within-session (w) and between-sessions (b) over complete crank cycle.

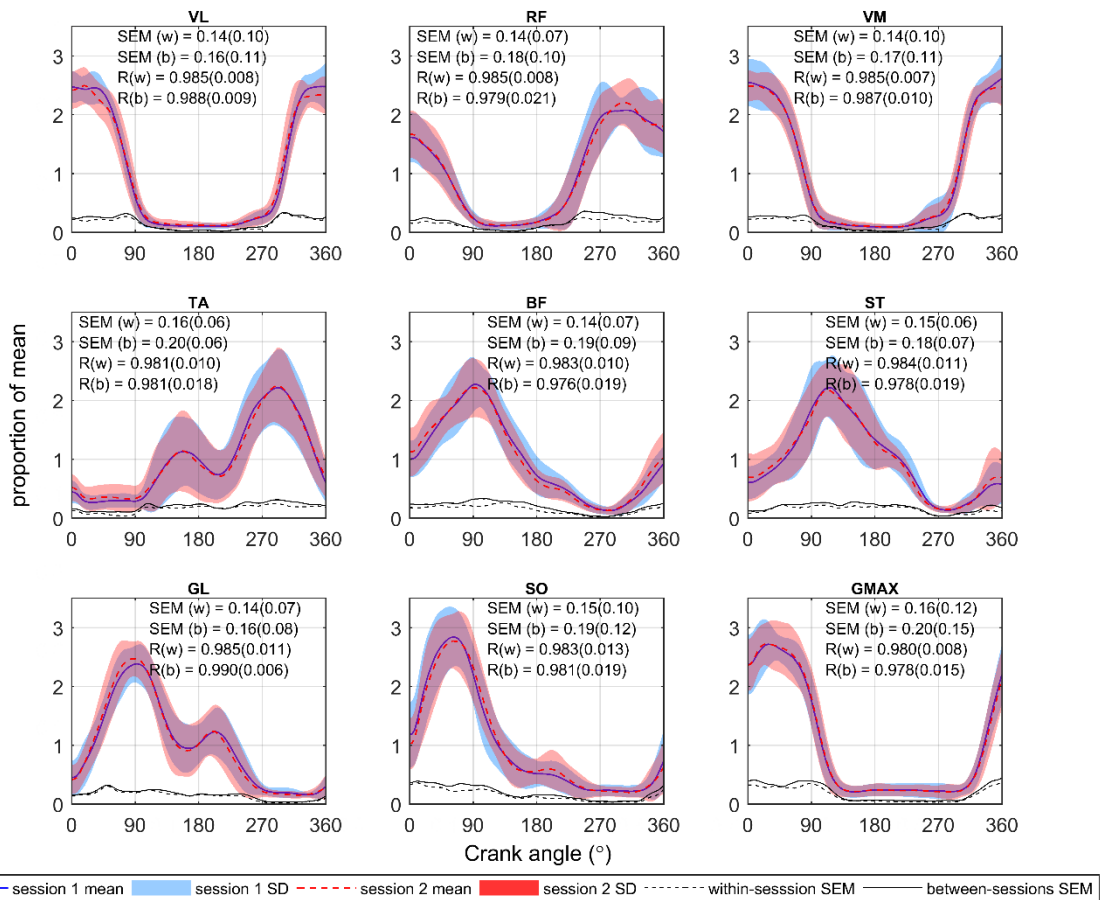


Figure 8: EMG linear envelopes (normalised to mean value in signal) for each muscle: group means for session one and two and standard error of measurement (SEM) within- and between-session. VL = vastus lateralis, RF = rectus femoris, VM = vastus medialis, TA = tibialis anterior, BF=biceps femoris, ST= semitendinosus, GL = gastrocnemius lateralis, SO = soleus, GMAX = gluteus maximus. Mean and standard deviation of SEM within-session (w) and between-sessions (b) over complete crank cycle.