

1 Targeting N-Cadherin (CDH2) and the malignant bone marrow microenvironment in acute
2 leukaemia.

3

4 Parker, Jessica¹, Hockney, Sean¹, Blaschuk, Orest W.², Pal, Deepali*^{1,3}

5 *Corresponding author: deepali.pal@northumbria.ac.uk

6 ¹ Department of Applied Sciences, Northumbria University, Newcastle upon Tyne, NE1
7 8ST. UK.

8 ² Zonula Incorporated, Kirkland QC, H9J 2X2. Canada.

9 ³ Wolfson Childhood Cancer Research Centre, Translational and Clinical Research
10 Institute, Faculty of Medical Sciences, Newcastle University, Herschel Building Level
11 6, Brewery Lane, Newcastle upon Tyne, NE1 7RU. UK.

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30 **Abstract**

31 This review discusses current research on acute paediatric leukaemia, the leukaemic bone
32 marrow microenvironment and recently discovered therapeutic opportunities to target
33 leukaemia-niche interactions. The tumour microenvironment plays an integral role in
34 conferring treatment resistance to leukaemia cells, this poses as a key clinical challenge that
35 hinders management of this disease. Here we focus on the role of the cell adhesion molecule
36 N-cadherin (CDH2) within the malignant bone marrow (BM) microenvironment and associated
37 signalling pathways that may bear promise as therapeutic targets. Additionally, we discuss
38 microenvironment driven treatment resistance and relapse, and elaborate the role of CDH2
39 mediated cancer cell protection from chemotherapy. Finally, we review emerging therapeutic
40 approaches that directly target CDH2 mediated adhesive interactions between the BM cell and
41 leukaemia cells.

42 **Background**

43 Leukaemia accounts for 31% of cancer diagnoses in children up to 14 years of age in the UK,
44 of which 401 children are diagnosed with acute lymphoblastic leukaemia (ALL) and 79 with
45 acute myeloid leukaemia (AML) per year(1), overall, the five-year survival of ALL and AML is
46 over 90% and 67%, respectively (1, 2). N-cadherin (CDH2) is a cell adhesion molecule that
47 mediates adhesive interactions between leukaemia cells and the cells of the BM (3, 4). These
48 interactions facilitate leukaemia cell survival, evasion from apoptosis and cell dormancy
49 ultimately resulting in treatment resistance (3, 4). Indeed, the niche-protected, dormant, non-
50 apoptotic leukaemia cells may re-emerge in relapsed cases to develop resistance to therapy
51 (5, 6).

52 Dysregulation of normal blood homeostasis is the main underlying developmental anomaly
53 that leads to ALL and AML. Leukaemogenesis usually comprises a series of steps with an
54 accumulation of genetic and epigenetic changes, inducing extensive alterations impacting cell
55 growth, metabolism, cell cycle progression, cell death and differentiation, leading to
56 preleukaemic haematopoietic stem cells (HSCs) and subsequently the development of ALL

57 and AML (7-9). Due to the complexity of epigenetic and genetic mutations, it is not fully
58 understood what cascade of events occur to give rise to the leukaemia phenotype and which
59 perturbations are responsible for driving leukaemogenesis. Through advancement of
60 technology, single-cell RNA sequencing has become more accurate, and been applied to
61 examine leukaemia cells at the transcriptional level. A study by Watcham, Kucinski and
62 Gottgens, 2019, presented data that suggest many leukaemia perturbations can gain
63 advantage over wild-type cells, and drive cells into a more active state (10). Indeed, many
64 studies show that *in utero* mutations are becoming more recognised as commonplace in acute
65 leukaemia and could be responsible for fusion genes in paediatric patients with ALL and AML,
66 these mutations are known as an initiating event (11-13). Fusion genes are chromosomal
67 aberrations that have a role in leukemogenesis (14), and can involve genes associated with
68 protein kinase pathways, transcription factor and epigenetic modifications (11).

69 Across mammals the number of HSCs per individual is thought to be conserved, with
70 approximately 300 HSCs at birth, compared to between 11,000 – 22,000 in adults.
71 Development of childhood leukaemia depends on initial somatic mutations in HSCs, and due
72 to the small HSC pool size these mutations are more likely to have a greater impact on the
73 HSC population (15). The Knudson 'two-hit' hypothesis, established in 1971, suggested that
74 dominantly inherited predisposition to cancer begins with a germline mutation, as can be seen
75 with fusion genes, however a second, somatic mutation is needed for tumorigenesis (16).

76 For example, only about 1% of children born with the ETV6-RUNX1 fusion gene develop the
77 second-hit mutation that is needed to transform to ALL, indicating the fusion gene mutation is
78 weakly penetrant (17). Many somatic mutations such as *TP53*, *RUNX1* and *IKZF1* are found
79 at the same sites of germline mutations in children who develop leukaemia (18-20). For
80 example, a germline mutation at *CEBPA* leads to the development of AML with almost
81 complete penetrance, this mutation is known to present favourable outcome (21).

82 Intensification of chemotherapeutic regimens is thought to be one of the main reasons for
83 increased survival in childhood leukaemia; however, such treatment is associated with high

84 morbidity and mortality rates. For example, in AML, high dose cytarabines used in young
85 adults (15 – 24 years old) were reported to have a benefit to outcome, however these results
86 could not be translated to paediatric patients. The COG trial AAML1031 intensified induction
87 chemotherapy with mitoxantrone and cytarabine and found that intensification did not achieve
88 a survival benefit in paediatric patients, since remission rates were comparable to the
89 AAML0531 trail which did not include intensifying induction chemotherapy. Moreover,
90 additional haematological toxicity was found to be associated with treatment intensification,
91 therefore showing an increased toxicity without any proportional benefit in treatment (22).
92 Studies have been conducted worldwide to analyse toxicity of paediatric acute leukaemia
93 treatment (23-25). Table 1 shows comparisons of different paediatric ALL protocols in the UK
94 and two European countries and their associated toxicities. Results show that up to 49% of
95 patients experienced an adverse event due to the chemotherapeutic agents used in their
96 treatment, with toxicity-induced mortality rates up to 3.7%. The children studied by
97 Zawitkowska et al, 2019 were evaluated for the 'grade' of toxicity, it was found that children
98 with grade 3 or higher were found to have a lower overall survival and event-free survival rate
99 compared to children with a lower grade of treatment toxicity (24).

100
101
102
103
104
105
106
107
108
109
110

111

112

Study	Hough et al., 2015	Zawitkowska et al., 2019	Franca et al., 2015
Study Location	UK	Poland	Italy
Study size (Patient numbers)	3126	1872	508
Total Adverse Events (AEs)	1835	3190	311
Patients affected by AEs	1164 (37.2%)	902 (48%)	251 (49.4%)
Most common toxicity	Infection (17.5%)	Infective episodes (32.3%)	Hepatic toxicity (40.2%)
Second most common toxicity	Methotrexate encephalopathy (8%)	Hepatotoxicity (28.2%)	Gastrointestinal toxicity (12.4%)
Third most common toxicity	Septicaemia (5.8%)	Gastrointestinal toxicities (20.4%)	Neurological toxicity (8.7%)
Non-relapse toxic mortality	35 (1.1%)	69 (3.7%)	N/A

113 **Table 1.** Comparison of paediatric ALL protocols from the UK and two European countries,
 114 including study size and number of patients affected by adverse events. The table shows the
 115 top three most common toxicities and alongside mortality rates due to treatment (23-25).

116

117 Chemoprotection induced by the leukaemia microenvironment is important in conferring
 118 treatment protection to cancer cells via mechanisms that include leukaemia cell – BM niche
 119 interactions and malignant dormancy (26, 27). ALL chemotherapies include DNA damaging
 120 and spindle poisons, which target the S and M phase of the cell cycle. These therapies rely
 121 on targeting actively cycling leukaemia cells, and therefore are ineffective against dormant
 122 cells which consequently lead to treatment resistance and relapse (28, 29). To improve
 123 efficacy of treatment and limit treatment failure and relapse, approaches including targeted
 124 therapy, immunotherapy and gene therapy are being explored.

125 Targeted therapy includes risk stratification, an approach where patients are grouped based
 126 on disease risk or therapy response from diagnostic tests. In a clinical trial for paediatric ALL
 127 (JPLSG MLL-10 trial), patients were stratified into 3 risk groups according to their *KMT2A* gene
 128 rearrangement status (*KMT2A-r*), age and presence of central nervous system (CNS)

129 leukaemia (30). High-dose cytarabine was given to KMT2A-r patients with haematopoietic
130 stem cell transplant (HSCT) option being reserved for high-risk patients. Consequently, this
131 removed the requirement for HSCT in patients with KMT2A-r (30). Whilst patient stratification
132 has contributed to the improved survival rates for paediatric ALL, intensifying chemotherapy
133 reaches a plateau where there is no additional benefit to patients but only an increased toxicity
134 exposure. To overcome the limitations of targeted therapy, novel approaches need to be
135 incorporated into the treatment protocol.

136 Immunotherapies have been explored to overcome the challenges presented by conventional
137 targeted therapies. For example, blinatumomab presented promising results in a phase I/II trial
138 with paediatric patients with relapsed/refractory ALL (29). In a phase III trial in paediatric
139 patients with B-ALL at high risk of relapse, blinatumomab was superior to conventional
140 consolidation therapy (31). However, blinatumomab presents unique and significant toxicities
141 of neurological events and cytokine release syndrome (CRS), which includes pyrexia,
142 headache, nausea, fatigue, and hypotension, although these findings were presented from
143 adults with relapsed B-ALL (32). CRS has been seen to be infrequent in low minimal residual
144 disease (MRD) settings and most neurological events could be reversed through interrupting
145 infusions (33), suggesting that blinatumomab could be effective with minimal toxicity in patients
146 with low MRD, although alternatives would be needed in other patients.

147 Gene therapy is another emerging route to overcome the challenges of conventional
148 therapies. T cell therapy involves genetically engineering chimeric antigen receptor (CAR) T
149 cells, coupling an anti-CD19 domain to intracellular T cell signalling domains of the T cell
150 receptor, which redirects cytotoxic T lymphocytes to cells expressing the CD19 antigen, in B
151 cell leukaemia (34). Anti-CD19 CAR T cell therapy, tisagenlecleucel, has been FDA-approved
152 after high remission rates were found in patients with ALL and whilst severe toxicities were
153 observed these effects were reversible (35, 36).

154

155 **The roles of Cadherins in the leukaemia microenvironment**

156 Classical cadherins are a calcium-dependent adhesion molecule family, grouped into Type-I
157 and Type-II subgroups based on the molecular features of their interactions via the cadherin
158 motifs (37). Neural (N)-cadherin (CDH2) and epithelial (E)-cadherin (CDH1) are Type-I
159 cadherins which are characterised by the cell adhesion recognition motif His-Ala-Val (HAV) in
160 their first extracellular domain (38, 39). CDH1 is a tumour suppressor protein which plays an
161 important role in regulating tissue homeostasis by modulating permeability barriers (i.e. tight
162 junctions) between compartments, and the functional state of CDH1 determines metastatic
163 potential (40). Functional activity of CDH1 can be modified in response to environmental
164 factors and CDH1 can be activated by monoclonal antibodies to inhibit metastasis at multiple
165 stages of the metastatic cascade (40). CDH2 is typically known for its role in morphogenetic
166 processes in health such as during the formation of cardiac and neural tissue, and in diseases
167 like solid tumours. Moreover, recent research indicate overexpression of *CDH2* in HSCs show
168 increased HSC attachment to BM endosteal surfaces (5). In disease, loss of *CDH1* and
169 upregulation of *CDH2* in cancer cells, leads to metastatic dissemination and activation of
170 several Epithelial-Mesenchymal Transition (EMT) transcription factors (41). EMT is a cellular
171 morphogenetic transition from a non-motile, epithelial phenotype into a migratory,
172 mesenchymal-like phenotype and is thought to be a driving force in tumourigenesis and
173 metastasis (42-46). CDH2 has been identified as an important molecule of interest in
174 leukaemia. A recent study demonstrated that this adhesion molecule was upregulated in
175 leukaemia cells primed by BM niche cells (4). Furthermore, MILE study and Bloodspot
176 database showed that multiple haematological malignancies exhibited *CDH2* upregulation
177 compared to healthy bone marrows (47, 48).

178 On a related note, osteoblast (OB)-Cadherin (CDH11) a type-II cadherin (35) important in the
179 formation of the neural crest (NC) cells, has been further shown in disease models to cause
180 tumour growth, cell survival and EMT (49-51). It has been further suggested that intracellular
181 downstream signalling of CDH11 is essential for maintenance and survival of premigratory NC

182 cells. in addition, cells require CDH11 for physiological cell-cell, adhesion-related EMT in the
183 preparatory steps prior to migration (52). However, the biological role of CDH11 in leukaemia
184 has not yet been explored.

185 **The role of CDH2 in the bone marrow niches and in chemoprotection**

186 Biological systems are complex where their complexity is characterised by multicellularity,
187 degeneracy and redundancy of the component cell types. The BM is a viscous tissue within
188 the bone comprised of two-well defined niches - endosteal and perivascular, where HSCs are
189 found in close proximity to osteoblasts (OB) and endothelial cells (EC) (53). All blood lineages
190 and immune cells are derived from the common precursor, HSC (54), which retains the ability
191 for both multipotency and self-renewal (55). The two niches are intertwined to create a
192 functional microenvironment, that facilitates cell communication during HSC development
193 consequently helping to maintain the full blood cell forming potential of HSCs (55).

194 It has been well-established that the endosteal niche is filled with mesenchymal stromal cells
195 (MSCs), osteoprogenitor cells, pre-OBs, mature OBs, osteocytes, and osteoclasts (56). OBs
196 play an important role in maintaining a functional microenvironment and are involved in stem
197 cell quiescence and proliferation (57). For example, SDF-1 α in OBs is associated with HSC
198 mobility (58). In disorders that effect HSCs, such as myelodysplastic syndrome, it has been
199 shown that suppressing osteogenic differentiation from MSC leads to their impairment in
200 supporting HSCs (59). Non collagenous bone matrix proteins, such as osteopontin (OPN) and
201 osteocalcin (OC), regulate cell migration and bone mineralisation and are believed to be linked
202 to cell proliferation, osteogenic differentiation and angiogenesis, however these processes are
203 yet to be defined in leukaemia cell biology (60).

204 Due to inaccessibility of reliable animal models, the niche microenvironment of ALL has not
205 been well-established. However, the remodelling of the BM vasculature following AML
206 leukaemogenesis has been studied and it was found that AML cells aid the niche
207 transformation into a preferential leukaemia microenvironment. These changes are

208 anatomically diverse; whilst vasculature in the endosteum was lost through disease
209 progression, central vessels survived with compromised function. This process was thought
210 to be due to the production of pro-inflammatory and anti-angiogenic cytokines from AML cells
211 in the endosteal lining which degrade the surrounding endothelium, as well as stromal
212 osteoblastic cells, together leading to the reduced capacity to support HSCs. Vasculature was
213 maintained in T-ALL murine models suggesting this vascular remodelling is specific to AML
214 (26, 61). The inflammatory cytokine, TNF- α , secreted by AML cells, directly induces E-selectin
215 which plays a role in promoting malignant cell survival, proliferation and chemoresistance (27).
216 AML engraftment also induces exogenous nitric oxide overproduction, which affects HSC
217 motility and increases HSC activation leading to reduction in their repopulating activity (26).
218 Increased vascular leakiness was observed in AML xenografts after induction therapy, leading
219 to poor drug delivery and the formation of areas with low perfusion rates, where leukaemia
220 cells migration resulted in microenvironment-induced treatment resistance (26).

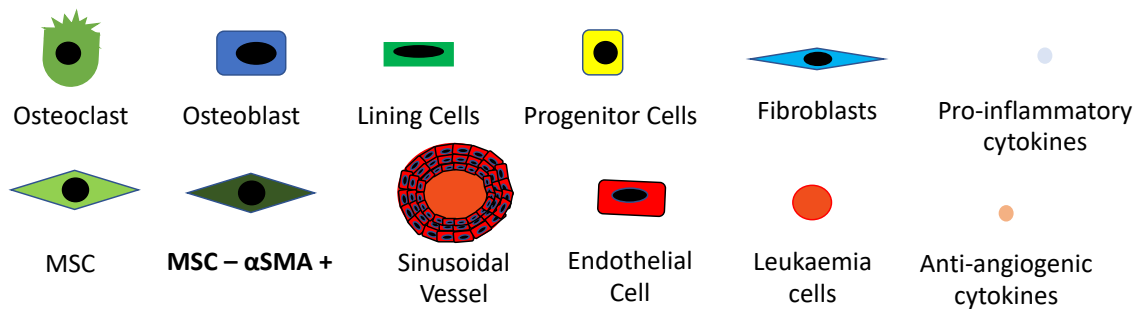
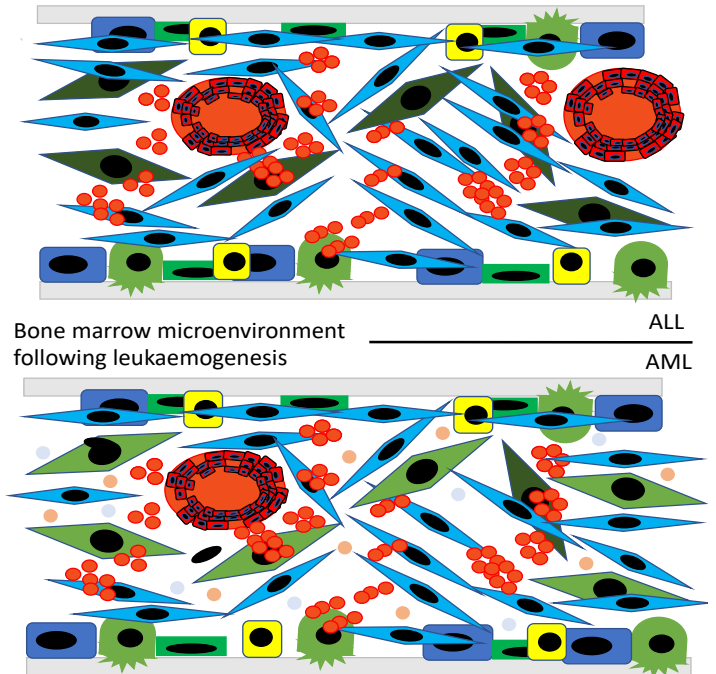
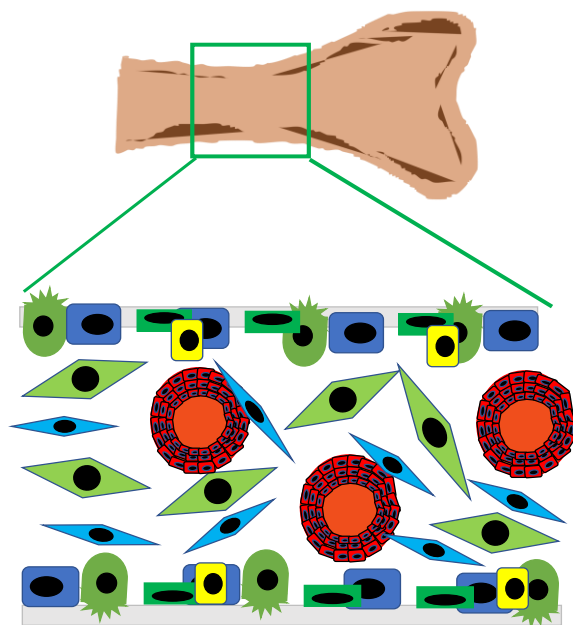
221 Peri-arteriolar stromal cells which are innervated by the sympathetic nervous system and
222 express neural markers NG2 and Nestin (NG2+/Nestin+ MSCs), have previously been found
223 to control HSC quiescence and hematopoiesis (62). The BM is known to be the site of dormant
224 disseminated tumour cells (DTC) and Nobre et al., 2021 found that NG2+/Nestin+ MSC drive
225 DTC dormancy which indicate that the perivascular niche is important for both HSC and DTC
226 dormancy. NG2+/Nestin+ MSCs produce TGF β 2 and BMP7, which signal a quiescent
227 pathway through TGFBRIII and BMPRII, thereby activating SMAD, p38 and p27 pathways
228 leading to dormancy (63). Treatment induced damage of endosteal and perivascular niches
229 have also been reported (64). Further research determined that leukaemia cells have an
230 important function in the development of a new therapy-induced niche formation. Following
231 treatment, secretion of cytokines and growth factors were found to increase in the
232 microenvironment likely due to secretion by the leukaemia cells (64). Indeed the leukaemia
233 niche has been reported to be transient, beginning initially as Nestin+ cells maturing into α -
234 SMA+ cells before terminating with fibre residues (64).

235

236

237

Healthy bone marrow microenvironment



238

239 Figure 1. Schematic of the bone marrow microenvironment in health and following
 240 leukaemogenesis and treatment in AML (top right) and ALL (top left). After leukaemogenesis
 241 and treatment, the microenvironment is remodelled, pro-inflammatory and anti-angiogenic
 242 cytokines are produced resulting in the loss of vasculature in the endosteal and osteoblastic
 243 cells. Of note, several cell types within the human bone marrow express CDH2, including but
 244 not limited to, MSC, fibroblasts, pericytes, osteoblasts and related cell types. Recent research
 245 suggests CDH2 expression to be upregulated in leukaemic bone marrows. Adapted from (4,
 246 26, 27, 53-63).

247

248

249

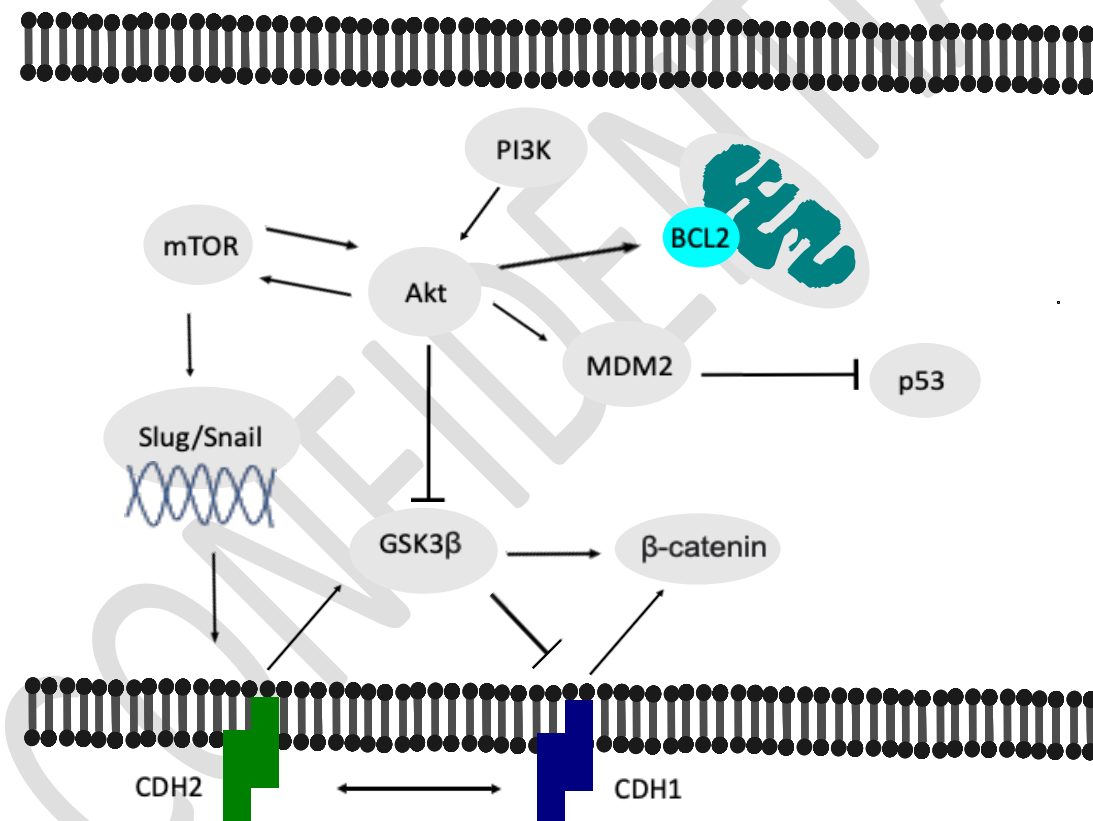
250 In keeping with these studies, recent research shows upregulation of CDH2, a known marker
251 of EMT, in niche primed leukaemia cells. This study demonstrated that knockdown of CDH2
252 in leukaemia cells reduce their proliferation while increasing sensitivity to dexamethasone
253 treatment (3, 4). Under physiological conditions, CDH2 plays a role in osteogenesis in the
254 endosteal niche, specifically in maintaining the precursor osteoblast pool (65). CDH2-
255 mediated interactions with osteoblasts are thought to play a role in supporting HSC function,
256 with HSC – osteoblast cell interactions enabling adhesion of HSCs to cells present in the
257 endosteal niche (5, 66). CDH2 is also expressed by various cell types associated with the
258 HSC niche (Figure 1), including stromal cells in the endosteal niche, and ECs and their
259 associated pericytes in the microvascular of the perivascular niche (5).

260 *CDH2* upregulation has been reported in human leukaemic bone marrows (4). A recent study
261 has shown that *CDH2* upregulation by niche-primed leukaemia is associated with increased
262 cancer proliferation and acquisition of treatment resistance and importantly this interaction is
263 druggable using the CDH2 antagonist ADH1 (3, 4). In adult AML CDH2 supports tumour
264 growth and aids in maintaining self-renewal characteristics of leukaemia stem cells (LSCs), as
265 CDH2⁺ cells have been found to engraft on NOD/SCID mice at a higher proportion than CDH2⁻
266 cells (6). CDH2 is also thought to support microenvironment-induced treatment protection in
267 AML (67). Indeed, adhesion interactions between LSCs and the BM microenvironment
268 activate signalling cascades, which regulate functions including cell survival, evasion of
269 apoptosis and cell dormancy. LSC interactions with the BM microenvironment enable them to
270 evade the cytotoxic effects of chemotherapeutic agents, suggesting there is a reliance on
271 adhesive interactions between AML LSCs and the BM for chemoprotection (5, 68). *CDH2*
272 overexpression in HSCs decreases *in vitro* cell division rate, this is likely due to the
273 sequestration of the CDH2 binding, intracellular β -catenin to the plasma membrane, thus
274 suppressing its activity as a transcription factor in the nucleus (6, 38). In support of this, adult
275 AML BM contains CDH2⁺ LSCs which are found in a quiescent state in G0/G1 cell cycle arrest,
276 which renders them less sensitive to chemotherapy (6, 69). Lastly, in adult AML, CDH2 is also

277 thought to play a role in drug resistance, CDH2⁺ LSCs were found to have a higher IC₅₀ of VP-
 278 16, an anti-leukaemia therapeutic drug, than the
 279 CDH2⁻ population (6, 70).

280 **Pathways associated with CDH2**

281 There are many pathways that are associated with CDH2 in various malignancies. Two
 282 pathways of relevance to this review are the Wnt/ β -catenin pathway and the PI3K/Akt/mTOR
 283 pathway, as detailed in Figure 2. It is of note, that there is limited research into these signalling
 284 pathways in acute leukaemia, clearly indicating an area warranting further exploration.



285

286 Figure 2. A schematic of the pathways and transcription factors associated with CDH2,
 287 including the PI3K/Akt/mTOR pathway and the Wnt/ β -catenin pathway. Arrows represent
 288 activation; bars represent inhibition, double-ended arrows in the pathway indicates
 289 upregulation of a molecule results in downregulation of the other and vice versa. Adapted from
 290 (71-85).

291

292

293 Hyperactivation of the phosphoinositide 3-kinases (PI3K)/Akt/mTOR signalling pathway has
294 been reported in 88% of ALL patients and is associated with poor prognosis and
295 chemotherapeutic resistance (74). The PI3K-Akt-mTOR pathway is important for
296 haematopoietic cells, regulating functions such as HSC proliferation, differentiation and
297 survival, and is furthermore constitutively activated in AML cells (78, 79). The presence of
298 PI3K/Akt/mTOR pathway has been well established in solid tumours and dimerization and
299 phosphorylation of PI3K leads to the downstream activation of Akt. Akt stimulates cell survival
300 by upregulating mouse double minute 2 homolog (*MDM2*), which inhibits *p53*, and upregulates
301 *BCL2*, both leading to inhibition of apoptosis. Akt activation subsequently triggers the
302 phosphorylation of mammalian target of rapamycin (mTOR) (81). mTOR is a conserved
303 serine/threonine kinase that belongs to the PI3K-related kinase family and has also been well-
304 established in solid tumours. It is a constituent of two signalling complexes, mTORC1 involved
305 in mRNA translation and protein synthesis and mTORC2 which controls cell survival and
306 migration (82, 84, 85). There is evidence to link p70S6K to the Akt/mTOR pathway in AML
307 (77). In solid tumours p70S6K activates the transcription factors, Slug and Snail, which
308 downregulates CDH1 and upregulates CDH2, leading to EMT (75).

309 Dysregulation in the Wnt/ β -catenin pathway can lead to initiation and progression of cancer,
310 including haematological malignancies, and β -catenin activation has been found to
311 contribute to ALL and AML drug resistance (71, 72, 83, 86). The inactivation of *GSK3 β* from
312 Akt-dependent phosphorylation prevents β -catenin phosphorylation, leading to the
313 activation of β -catenin independent genes and uncontrolled cell proliferation (71). CDH2
314 regulates Wnt/ β -catenin signalling, a conserved pathway that plays a role in physiological
315 processes, including differentiation, proliferation, and cell fate determination. CDH1 is
316 known to inhibit the activation of the Wnt pathway, and in ALL CDH1 has been shown to be
317 decreased, indicating Wnt pathway activation (76). Additionally in ALL, Akt has been shown
318 to inhibit *GSK3 β* leading to the activation of β -catenin (80).

319

320 **Therapeutic Approaches targeting CDH2**

321 **Exherin (ADH-1)**

322 The CDH2 antagonist ADH-1 is a cyclic pentapeptide which competitively inhibits CDH2
323 (Figure 3), as it contains the cadherin cell adhesion recognition sequence His-Ala-Val (38, 87).

324 The proposed mechanism of action of ADH-1 in cancer is that it results in apoptosis *in vitro*,
325 and causes inhibition of tumour cell migration in addition to altering the tumour vasculature *in*

326 *vivo* (88-90). Pal et al., 2022 found that ADH-1 showed high efficacy *in vitro* and *in vivo* against
327 patient-derived ALL cells, where ADH-1 reduced proliferation of ALL cells *in vitro*, as indicated

328 by a reduced number of blasts in the S phase of the cell cycle (4). This research further
329 assessed ADH-1 activity on CDH2 knockdown ALL cells, where ADH-1 treatment sensitivity

330 was confirmed only in the wildtype ALL cells that did not harbour the CDH2 knockdown,
331 thereby corroborating specificity of ADH-1 against CDH2 and suggesting against the likelihood

332 of off-target effects (4). This study further validated ADH-1 to show efficacy both as a single
333 agent and in combination with dexamethasone, in a patient-derived-xenograft (PDX) mouse

334 model, where addition of ADH-1 to dexamethasone did not result in any additional toxicity (4).
335 Of note, ADH-1 is an FDA approved compound with “orphan drug” status for use in

336 melanomas(91), and ADH-1 treatment in patients with solid tumours was well tolerated
337 resulting only in a few adverse events, most of which were grade 1 or 2, thereby showing a

338 better tolerance than most current treatments (92). These findings indicate ADH-1 to be a
339 potentially promising therapeutic agent that could be repurposed from solid cancers to

340 leukaemia treatment.

341

342

343

344

345

346

347

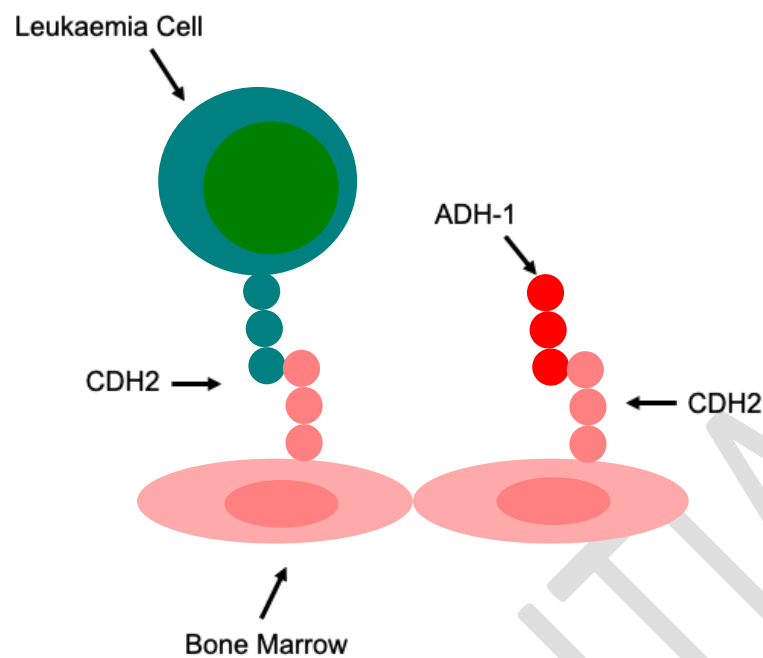
348

349

350

351

352



353 Figure 3. ADH-1 competitively binds to CDH2 on bone marrow cells, preventing leukaemia-
354 niche cell binding of leukaemia cells within the bone marrow microenvironment. Adapted from
355 (38, 87).

356

357 In addition, ADH-1-modified liposomes (A-LP) have been successfully constructed with the

358 aim of enhancing chemotherapy efficacy and preventing metastasis and was tested using a

359 PTX-resistant breast cancer cell line, MCF7 PTX-R, which was established into a tumour

360 model using subcutaneous inoculation into the right flanks of female BALB/c nude mice.

361 Results found that cellular uptake was increased due to the CDH2 expressed after EMT in the

362 MCF7 PTX-R cells (92). Treatment with the A-LP showed cancer cells to have an increased

363 chemo-sensitivity, with EMT to be somewhat suppressed.

364 ADH-1 has been further shown to improve immunotherapy by tumour-infiltrating lymphocyte

365 (TIL)-related treatment. The immune dysfunction mechanism including programmed death

366 ligand-1 (PDL1) and indole amine 2,3-dioxygenase (IDO-1) induces apoptosis, both PD-L1

367 and IDO-1 are increased after EMT and immunosuppression is enforced. Therefore targeting

368 CDH2 improved the efficacy of (TIL)-related treatment by decreasing PD-L1 and IDO-1, and

369 indeed ADH-1 with (TIL)-treatment reduced tumour size and increased survival in the mouse

370 models (93). While ADH-1 has been documented in cancer pre-clinical studies and solid
371 tumour clinical trials, in-depth mechanism of action of this drug remains unexplored. Further
372 research needs to be conducted to develop an in-depth understanding of ADH-1, including
373 scrutiny of any possible mechanisms of resistance that could arise following ADH-1 treatment.
374 In addition, several next generation antagonists, including small molecule inhibitors of CDH2
375 are being developed (38) and their role as potential anti-leukaemia treatment needs to be
376 investigated.

377 **CDH2 small molecule antagonists**

378 Much less is known concerning the biological effects of other types of CDH2 antagonists, as
379 they have not been extensively developed for use as cancer therapeutics (38). A large number
380 of non-peptidyl peptidomimetics of ADH-1 have been recently identified (94, 95), for example,
381 the small molecule LCRF-0006 is an ADH-1 peptidomimetic that inhibits CDH2 function,
382 induces apoptosis in multiple myeloma (MM) and synergises with bortezomib to enhance MM
383 cell death *in vitro* (94).

384 Non-peptide peptidomimetics of the CDH2 Trp-containing amino-terminus have also been
385 discovered and are being developed as cancer therapeutics (38, 96). In particular, the
386 peptidomimetic designated Compound 15, a piperidin-4-amine which acts as a CDH2
387 antagonist, has been shown to induce apoptosis of multiple myeloma, glioblastoma and
388 pancreatic cancer cells, as well as fibroblast and cancer-associated death *in vitro* (38, 97).
389 However, the ability of this small molecule to effect leukaemia blast viability as well as its
390 mechanism of action remains unexplored.

391

392

393

394

395

396 **Targeting other pathways in combination with CDH2**

397 Although targeted therapy underpinning oncogene addiction has shown great promise in
398 cancer treatment, it is associated with emergence of treatment resistant clones. Combinatorial
399 therapies target multiple cancer pathways, and thereby aim to mitigate occurrence of treatment
400 resistance. Furthermore, up to 40% of ALL patients present with CNS involvement, due to the
401 ability of leukaemia cells to penetrate the blood-brain-barrier (BBB) (98). Although it is now
402 well established that achieving CNS clearance in ALL is essential for long term disease cure,
403 CNS-directed therapy is associated with significant toxicity (99). This highlights need for new
404 and improved combinatorial treatments in ALL to prevent treatment resistance and mitigate
405 treatment toxicity. Indeed combination therapies containing dexamethasone, a glucocorticoid
406 routinely used to treat ALL, with venetoclax or ADH-1 have been shown to increase leukaemia-
407 free long-term survival in pre-clinical mouse models and patients (4, 100, 101). Furthermore,
408 ADH-1 and dexamethasone have been found to
409 show high efficacy when tested in combination on patient derived xenograft (PDX) mouse
410 models transplanted with high risk ALL. The ADH1/dexamethasone combination was found to
411 significantly reduce the proportion of leukaemia blasts *in vivo* compared to the
412 dexamethasone-only arm, and moreover addition of ADH-1 to dexamethasone did not result
413 in any additional toxicity (4).

414 Adults with BCR-ABL+ ALL have poor prognosis; therefore, dexamethasone was tested in a
415 triple combination with venetoclax and tyrosine kinase inhibitors (TKI), imatinib or dasatinib.
416 Both combinations were shown to be superior to single agents and double combinations in
417 terms of tumour size and survival, although the combination with dasatinib was shown to be
418 more effective (101). Researching the value of adding ADH-1 to a
419 dexamethasone/venetoclax/TKI is warranted especially in high risk and/or refractory disease
420 to assess if this combination would improve efficacy and minimise emergence of treatment
421 resistant clones (101). Other drugs and pathways where adding ADH-1 as a combinatorial
422 treatment might be valuable is as discussed below.

423 Dysregulation in the PI3K/Akt/mTOR pathway has been well established as a component of
424 AML pathogenesis. Many pharmacological inhibitors within this pathway have been evaluated
425 in preclinical settings, however there is yet to be meaningful clinical effectiveness of inhibition
426 of this pathway for AML. Buparlisib is an oral pan-class I PI3K inhibitor, has completed a Phase
427 I trial of patients with acute leukaemia and at doses of 80mg/day was found to be tolerable
428 with a modest single-agent efficacy. Buparlisib has also been seen to cross the blood-brain-
429 barrier which is of importance in ALL with CNS infiltration (102, 103).

430 Idelalisib is a PI3K- δ inhibitor, more specifically p110 δ a primary PI3K isoform in B cells and
431 has shown activity in lymphoid malignancies and been FDA approved for relapsed CLL,
432 follicular lymphoma and small lymphocytic lymphoma (104). Haematological malignancies
433 such as relapsed CLL, follicular lymphoma and small lymphocytic lymphoma have been
434 observed to depend on pre-B cell receptor signalling, which can also be seen in the majority
435 of TCF3-PBX1 BCP-ALLs. The specificity of idelalisib to p110 δ , results in a low toxicity profile,
436 making it a promising therapeutic for TCF3-PBX1 BCP-ALL patients (105, 106). Interestingly,
437 significant CDH2 upregulation in TCF3-PBX1 leukaemic BM combined with high ADH1
438 efficacy seen in TCF3-HLF PDX samples would suggest that combining a CDH2 antagonist
439 with idelalisib might be potentially beneficial.

440 mTOR inhibitors have shown promise in preclinical models of ALL through direct inhibition of
441 tumour cell growth and reversal of glucocorticoid resistance and have demonstrated *in vitro*
442 synergy with dexamethasone (107). Everolimus, presents these preclinical characteristics as
443 a single agent, making it a good candidate for combination treatment. Moreover, there is a
444 phase II study of Everolimus in combination with vincristine, prednisone, pegaspargase and
445 doxorubicin in relapsed ALL (108). Everolimus was also tested in chronic myeloid leukaemia
446 patients and found that in combination with imatinib, treatment was effective in both sensitive
447 and resistant cases (109).

448 BEZ-235 is a dual pan-class I PI3K and mTOR inhibitor that has been tested in adult patients
449 with relapsed/refractory acute leukaemia. Clinical development of BEZ-235 has been

450 terminated due to suboptimal pharmacokinetic properties. Although this study found that
 451 efficacy observed in ALL patients warrant further clinical exploration into dual PI3K/mTOR
 452 inhibitors, in particular patients with Ph + BCP-ALL or T-ALL may benefit from these
 453 treatments (110). Given the link between CDH2, mTOR and EMT all of which play an important
 454 role in cancer biology (the role of EMT in non-epithelial cancers such as leukaemia is an
 455 emerging concept (111)), including niche-driven leukaemia cell behaviour, combining a CDH2
 456 antagonist with mTOR inhibitors may have a potential therapeutic benefit.

457 Despite the role Wnt plays within acute leukaemias and its connection with CDH2, there has
 458 not been any clinical or preclinical testing with Wnt inhibitors, suggesting a potential area of
 459 further research, some pre-clinical antagonists are highlighted in table 2 (112-116). Table 2
 460 highlights inhibitors that have been tested against other cell lines and malignancies.

Name	Target	Clinical Trial Progression?
ADH-1	CDH2	Phase II (solid tumours)
AZD2014	mTORC1 and mTORC2	Phase II (Solid tumours)
AZD8055	mTORC1 and mTORC2	Phase I (Halted Development)
BEZ-235	PI3K and mTOR	Phase I (ALL) Phase II (Solid tumours)
Buparlisib	PI3K	Phase I
Capmatinib	Wnt	Pre-clinical
Compound 15	CDH2	Pre-clinical (solid and haematological cancers)
Dasatinib	Tyrosine kinase	FDA approved for paediatric chronic myeloid leukemia
Dexamethasone	Glucocorticoid Receptors	FDA approved
Everolimus	mTOR	FDA approved (Solid tumours)
Idelalisib	PI3K- δ	FDA approved for chronic lymphocytic leukemia
Imatinib	Tyrosine kinase	FDA approved for chronic myeloid leukemia
Isoquercitrin	Wnt	Pre-clinical
IWP-4	Wnt	Pre-clinical
XAV-939	Wnt	Pre-clinical
Venetoclax (ABT-199)	BCL-2	FDA approved for AML

461 Table 2. A list of the therapeutics, their targets, and their progressions through clinical trials

462 **Conclusion**

463 In conclusion, CDH2 is an important molecule in both the healthy and malignant BM
 464 microenvironment, supporting both non-malignant haematopoietic cells and leukaemia cells.
 465 CDH2 supports tumour growth and promotes microenvironment-mediated treatment

466 protection, decrease cell division rate, and potentially plays a role in cancer dormancy. ADH-
467 1, a first generation CDH2 inhibitor used in solid tumour clinical trials, demonstrated a well-
468 tolerated toxicity profile and therefore may be an ideal candidate for combinatorial treatment
469 in acute leukaemia. It is important to note that only CDH2 antagonists target the extracellular
470 domain of cell surface receptors making them a unique class of therapeutic drugs.
471 Furthermore, targeting other pathways that are associated with CDH2 may overcome
472 environment-mediated drug resistance and may help reduce the rate of relapse in paediatric
473 acute leukaemia. Next generation CDH2 antagonists such as small molecule inhibitors with
474 improved potency and formulation are emerging as a unique class of anti-cancer therapeutics.
475 These are potentially capable of targeting microenvironment mediated malignant dormancy
476 and treatment resistance in leukaemia and following in-depth preclinical and clinical validation
477 may provide improved and low toxicity treatment options in paediatric leukaemia.

478 **Competing Interests**

479 OWB holds shares in Zonula Incorporated. The company is developing N-cadherin
480 antagonists (such as compound 15) for the treatment of fibroblast-associated diseases. DP
481 and her team which includes JP and SH, are collaborating in studies investigating the ability
482 of compound 15 to act as a therapeutic for the treatment of ALL.

483
484
485
486
487
488
489
490
491
492
493
494
495
496
497
498
499
500
501

502 **References:**

503

504 1. England PH. Children, teenagers and young adults UK cancer statistics report 2021.
505 In: England PH, editor. Internet2021.506 2. Bartram J, Veys P, Vora A. Improvements in outcome of childhood acute
507 lymphoblastic leukaemia (ALL) in the UK - a success story of modern medicine through
508 successive UKALL trials and international collaboration. *Br J Haematol.* 2020;191(4):562-7.509 3. Borbaran-Bravo N, Arreba-Tutusaus P, Ritter MU, Nasri M, Klimiankou M, Skokowa
510 J. Regenerative medicine meets translational oncology: Modeling leukemic bone marrow
511 niche. *Cell Rep Med.* 2022;3(8):100724.512 4. Pal D, Blair HJ, Parker J, Hockney S, Beckett M, Singh M, et al. hiPSC-derived bone
513 marrow milieu identifies a clinically actionable driver of niche-mediated treatment resistance
514 in leukemia. *Cell Reports Medicine.* 2022;3(3):100717.515 5. Mroziak KM, Blaschuk OW, Cheong CM, Zannettino ACW, Vandyke K. N-cadherin in
516 cancer metastasis, its emerging role in haematological malignancies and potential as a
517 therapeutic target in cancer. *BMC cancer.* 2018;18(1):1-16.518 6. Zhi L, Gao Y, Yu C, Zhang Y, Zhang B, Yang J, et al. N-Cadherin aided in
519 maintaining the characteristics of leukemic stem cells. *The Anatomical Record.*
520 2016;299(7):990-8.521 7. Yamashita M, Dellorusso PV, Olson OC, Passegué E. Dysregulated haematopoietic
522 stem cell behaviour in myeloid leukaemogenesis. *Nature Reviews Cancer.* 2020;20(7):365-
523 82.524 8. Belver L, Ferrando A. The genetics and mechanisms of T cell acute lymphoblastic
525 leukaemia. *Nat Rev Cancer.* 2016;16(8):494-507.526 9. Iacobucci I, Mullighan CG. Genetic Basis of Acute Lymphoblastic Leukemia. *J Clin*
527 *Oncol.* 2017;35(9):975-83.528 10. Watcham S, Kucinski I, Gottgens B. New insights into hematopoietic differentiation
529 landscapes from single-cell RNA sequencing. *Blood, The Journal of the American Society of*
530 *Hematology.* 2019;133(13):1415-26.531 11. Chen X, Wang F, Zhang Y, Ma X, Cao P, Yuan L, et al. Fusion gene map of acute
532 leukemia revealed by transcriptome sequencing of a consecutive cohort of 1000 cases in a
533 single center. *Blood Cancer Journal.* 2021;11(6):1-10.534 12. Shin S-Y, Lee H, Lee S-T, Choi JR, Jung CW, Koo HH, et al. Recurrent somatic
535 mutations and low germline predisposition mutations in Korean ALL patients. *Scientific*
536 *Reports.* 2021;11(1):8893.537 13. Winer P, Muskens IS, Walsh KM, Vora A, Moorman AV, Wiemels JL, et al. Germline
538 variants in predisposition genes in children with Down syndrome and acute lymphoblastic
539 leukemia. *Blood Advances.* 2020;4(4):672-5.540 14. Wang Y, Wu N, Liu D, Jin Y. Recurrent fusion genes in leukemia: an attractive target
541 for diagnosis and treatment. *Current Genomics.* 2017;18(5):378-84.542 15. Rozhok AI, Salstrom JL, DeGregori J. Stochastic modeling reveals an evolutionary
543 mechanism underlying elevated rates of childhood leukemia. *Proceedings of the National*
544 *Academy of Sciences.* 2016;113(4):1050-5.545 16. Knudson Jr AG. Mutation and cancer: statistical study of retinoblastoma.
546 *Proceedings of the National Academy of Sciences.* 1971;68(4):820-3.547 17. Böiers C, Richardson SE, Laycock E, Zriwil A, Turati VA, Brown J, et al. A human
548 IPS model implicates embryonic B-myeloid fate restriction as developmental susceptibility to
549 B acute lymphoblastic leukemia-associated ETV6-RUNX1. *Developmental cell.*
550 2018;44(3):362-77. e7.551 18. Demir S, Boldrin E, Sun Q, Hampp S, Tausch E, Eckert C, et al. Therapeutic
552 targeting of mutant p53 in pediatric acute lymphoblastic leukemia. *Haematologica.*
553 2020;105(1):170.554 19. Maciel ALT, da Conceição Barbosa T, Blunck CB, Wolch K, Machado AdAL, da
555 Costa ES, et al. IKZF1 deletions associate with CRLF2 overexpression leading to a poor

- 556 prognosis in B-cell precursor acute lymphoblastic leukaemia. *Translational Oncology*.
557 2022;15(1):101291.
- 558 20. Malla TM, Shah ZA, Bhat AH, Malik MA, Baba RA, Rasool R, et al. Fishing for
559 ETV6/RUNX1 fusion and MLL gene rearrangements and their additional abnormalities in
560 childhood acute lymphoblastic leukemia patients of Kashmir. *Gene*. 2023;856:147128.
- 561 21. Liao Xy, Fang Jp, Zhou Dh, Qiu Ky. CEBPA are independent good prognostic factors
562 in pediatric acute myeloid leukemia. *Hematological Oncology*. 2022;40(2):258-68.
- 563 22. Elgarten CW, Wood AC, Li Y, Alonzo TA, Brodersen LE, Gerbing RB, et al.
564 Outcomes of intensification of induction chemotherapy for children with high-risk acute
565 myeloid leukemia: A report from the Children's Oncology Group. *Pediatr Blood Cancer*.
566 2021;68(12):e29281.
- 567 23. Hough R, Rowntree C, Goulden N, Mitchell C, Moorman A, Wade R, et al. Efficacy
568 and toxicity of a paediatric protocol in teenagers and young adults with Philadelphia
569 chromosome negative acute lymphoblastic leukaemia: results from UKALL 2003. *British*
570 *Journal of Haematology*. 2016;172(3):439-51.
- 571 24. Zawitkowska J, Lejman M, Zaucha-Prażmo A, Drabko K, Płonowski M, Balsa J, et al.
572 Grade 3 and 4 toxicity profiles during therapy of childhood acute lymphoblastic leukemia. *in*
573 *vivo*. 2019;33(4):1333-9.
- 574 25. Franca R, Rebora P, Bertorello N, Fagioli F, Conter V, Biondi A, et al.
575 Pharmacogenetics and induction/consolidation therapy toxicities in acute lymphoblastic
576 leukemia patients treated with AIEOP-BFM ALL 2000 protocol. *The Pharmacogenomics*
577 *Journal*. 2017;17(1):4-10.
- 578 26. Passaro D, Di Tullio A, Abarategi A, Rouault-Pierre K, Foster K, Ariza-McNaughton
579 L, et al. Increased vascular permeability in the bone marrow microenvironment contributes to
580 disease progression and drug response in acute myeloid leukemia. *Cancer cell*.
581 2017;32(3):324-41. e6.
- 582 27. Barbier V, Erhani J, Fiveash C, Davies JM, Tay J, Tallack MR, et al. Endothelial E-
583 selectin inhibition improves acute myeloid leukaemia therapy by disrupting vascular niche-
584 mediated chemoresistance. *Nature communications*. 2020;11(1):2042.
- 585 28. Ebinger S, Özdemir EZ, Ziegenhain C, Tiedt S, Alves CC, Grunert M, et al.
586 Characterization of rare, dormant, and therapy-resistant cells in acute lymphoblastic
587 leukemia. *Cancer cell*. 2016;30(6):849-62.
- 588 29. Pal D, Heidenreich O, Vormoor J. Dormancy stems the tide of chemotherapy. *Cancer*
589 *cell*. 2016;30(6):825-6.
- 590 30. Tomizawa D, Miyamura T, Imamura T, Watanabe T, Moriya Saito A, Ogawa A, et al.
591 A risk-stratified therapy for infants with acute lymphoblastic leukemia: a report from the
592 JPLSG MLL-10 trial. *Blood*. 2020;136(16):1813-23.
- 593 31. von Stackelberg A, Locatelli F, Zugmaier G, Handgretinger R, Trippett TM, Rizzari C,
594 et al. Phase I/Phase II Study of Blinatumomab in Pediatric Patients With
595 Relapsed/Refractory Acute Lymphoblastic Leukemia. *J Clin Oncol*. 2016;34(36):4381-9.
- 596 32. Topp MS, Gökbuget N, Stein AS, Zugmaier G, O'Brien S, Bargou RC, et al. Safety
597 and activity of blinatumomab for adult patients with relapsed or refractory B-precursor acute
598 lymphoblastic leukaemia: a multicentre, single-arm, phase 2 study. *Lancet Oncol*.
599 2015;16(1):57-66.
- 600 33. Gökbuget N, Dombret H, Bonifacio M, Reichle A, Graux C, Faul C, et al.
601 Blinatumomab for minimal residual disease in adults with B-cell precursor acute
602 lymphoblastic leukemia. *Blood*. 2018;131(14):1522-31.
- 603 34. Kalos M, Levine BL, Porter DL, Katz S, Grupp SA, Bagg A, et al. T cells with chimeric
604 antigen receptors have potent antitumor effects and can establish memory in patients with
605 advanced leukemia. *Science translational medicine*. 2011;3(95):95ra73-95ra73.
- 606 35. Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H, et al.
607 Tisagenlecleucel in Children and Young Adults with B-Cell Lymphoblastic Leukemia. *N Engl*
608 *J Med*. 2018;378(5):439-48.

- 609 36. Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, et al. Chimeric
610 antigen receptor T cells for sustained remissions in leukemia. *New England Journal of*
611 *Medicine*. 2014;371(16):1507-17.
- 612 37. Nollet F, Kools P, Van Roy F. Phylogenetic analysis of the cadherin superfamily
613 allows identification of six major subfamilies besides several solitary members. *Journal of*
614 *molecular biology*. 2000;299(3):551-72.
- 615 38. Blaschuk OW. Potential Therapeutic Applications of N-Cadherin Antagonists and
616 Agonists. *Frontiers in Cell and Developmental Biology*. 2022;10:866200.
- 617 39. Kashef J, Köhler A, Kuriyama S, Alfandari D, Mayor R, Wedlich D. Cadherin-11
618 regulates protrusive activity in *Xenopus* cranial neural crest cells upstream of Trio and the
619 small GTPases. *Genes & Development*. 2009;23(12):1393-8.
- 620 40. Na T-Y, Schecterson L, Mendonsa AM, Gumbiner BM. The functional activity of E-
621 cadherin controls tumor cell metastasis at multiple steps. *Proceedings of the National*
622 *Academy of Sciences*. 2020;117(11):5931-7.
- 623 41. Onder TT, Gupta PB, Mani SA, Yang J, Lander ES, Weinberg RA. Loss of E-
624 cadherin promotes metastasis via multiple downstream transcriptional pathways. *Cancer*
625 *research*. 2008;68(10):3645-54.
- 626 42. Christofori G. Changing neighbours, changing behaviour: cell adhesion molecule-
627 mediated signalling during tumour progression. *The EMBO journal*. 2003;22(10):2318-23.
- 628 43. Gupta GP, Massagué J. Cancer metastasis: building a framework. *Cell*.
629 2006;127(4):679-95.
- 630 44. Thiery JP, Sleeman JP. Complex networks orchestrate epithelial–mesenchymal
631 transitions. *Nature reviews Molecular cell biology*. 2006;7(2):131-42.
- 632 45. Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in
633 development and disease. *cell*. 2009;139(5):871-90.
- 634 46. Craene BD, Berx G. Regulatory networks defining EMT during cancer initiation and
635 progression. *Nature Reviews Cancer*. 2013;13(2):97-110.
- 636 47. Bagger FO, Kinalis S, Rapin N. BloodSpot: a database of healthy and malignant
637 haematopoiesis updated with purified and single cell mRNA sequencing profiles. *Nucleic*
638 *acids research*. 2019;47(D1):D881-D5.
- 639 48. Haferlach T, Kohlmann A, Wieczorek L, Basso G, Kronnie GT, Béné M-C, et al.
640 Clinical utility of microarray-based gene expression profiling in the diagnosis and
641 subclassification of leukemia: report from the International Microarray Innovations in
642 Leukemia Study Group. *Journal of clinical oncology*. 2010;28(15):2529-37.
- 643 49. Piao J, You K, Guo Y, Zhang Y, Li Z, Geng L. Substrate stiffness affects epithelial-
644 mesenchymal transition of cervical cancer cells through miR-106b and its target protein
645 DAB2. *International Journal of Oncology*. 2017;50(6):2033-42.
- 646 50. Row S, Liu Y, Alimperti S, Agarwal SK, Andreadis ST. Cadherin-11 is a novel
647 regulator of extracellular matrix synthesis and tissue mechanics. *Journal of cell science*.
648 2016;129(15):2950-61.
- 649 51. Yoshioka R, Kita Y, Nagahira A, Manno A, Makita N, Tomita U, et al. Quantitative
650 analysis of cadherin-11 and β -catenin signalling during proliferation of rheumatoid arthritis-
651 derived synovial fibroblast cells. *Journal of Pharmacy and Pharmacology*. 2015;67(8):1075-
652 82.
- 653 52. Manohar S, Camacho-Magallanes A, Echeverria Jr C, Rogers CD. Cadherin-11 is
654 required for neural crest specification and survival. *Frontiers in Physiology*. 2020;11:563372.
- 655 53. Bello AB, Park H, Lee S-H. Current approaches in biomaterial-based hematopoietic
656 stem cell niches. *Acta biomaterialia*. 2018;72:1-15.
- 657 54. Jiang N, Chen M, Yang G, Xiang L, He L, Hei TK, et al. Hematopoietic stem cells in
658 neural-crest derived bone marrow. *Scientific reports*. 2016;6(1):1-11.
- 659 55. Zhang P, Zhang C, Li J, Han J, Liu X, Yang H. The physical microenvironment of
660 hematopoietic stem cells and its emerging roles in engineering applications. *Stem Cell*
661 *Research & Therapy*. 2019;10(1):1-13.
- 662 56. Le PM, Andreeff M, Battula VL. Osteogenic niche in the regulation of normal
663 hematopoiesis and leukemogenesis. *Haematologica*. 2018;103(12):1945.

- 664 57. Kajiume T, Kawahara Y, Yuge L, Kobayashi M. Osteoblastic adherence regulates
665 hematopoietic stem cell self-renewal and differentiation: A conceptual in vitro and in vivo
666 study. *Stem Cell Investigation*. 2021;8:21.
- 667 58. Mangialardi G, Cordaro A, Madeddu P. The bone marrow pericyte: an orchestrator of
668 vascular niche. *Regenerative Medicine*. 2016;11(8):883-95.
- 669 59. Hayashi Y, Kawabata KC, Tanaka Y, Uehara Y, Mabuchi Y, Murakami K, et al. MDS
670 cells impair osteolineage differentiation of MSCs via extracellular vesicles to suppress
671 normal hematopoiesis. *Cell Reports*. 2022;39(6):110805.
- 672 60. Carvalho MS, Poundarik AA, Cabral J, da Silva CL, Vashishth D. Biomimetic
673 matrices for rapidly forming mineralized bone tissue based on stem cell-mediated
674 osteogenesis. *Scientific reports*. 2018;8(1):1-16.
- 675 61. Duarte D, Hawkins ED, Akinduro O, Ang H, De Filippo K, Kong IY, et al. Inhibition of
676 endosteal vascular niche remodeling rescues hematopoietic stem cell loss in AML. *Cell stem
677 cell*. 2018;22(1):64-77. e6.
- 678 62. Kunisaki Y, Bruns I, Scheiermann C, Ahmed J, Pinho S, Zhang D, et al. Arteriolar
679 niches maintain haematopoietic stem cell quiescence. *Nature*. 2013;502(7473):637-43.
- 680 63. Nobre AR, Risson E, Singh DK, Di Martino JS, Cheung JF, Wang J, et al. Bone
681 marrow NG2+/Nestin+ mesenchymal stem cells drive DTC dormancy via TGF- β 2. *Nature
682 cancer*. 2021;2(3):327-39.
- 683 64. De Rooij JD, Zwaan CM, van den Heuvel-Eibrink M. Pediatric AML: from biology to
684 clinical management. *Journal of clinical medicine*. 2015;4(1):127-49.
- 685 65. Alimperti S, Andreadis ST. CDH2 and CDH11 act as regulators of stem cell fate
686 decisions. *Stem cell research*. 2015;14(3):270-82.
- 687 66. Zhao M, Tao F, Venkatraman A, Li Z, Smith SE, Unruh J, et al. N-cadherin-
688 expressing bone and marrow stromal progenitor cells maintain reserve hematopoietic stem
689 cells. *Cell reports*. 2019;26(3):652-69. e6.
- 690 67. Tabe Y, Konopleva M. Role of microenvironment in resistance to therapy in AML.
691 *Current hematologic malignancy reports*. 2015;10(2):96-103.
- 692 68. Barwe SP, Quagliano A, Gopalakrishnapillai A. Eviction from the sanctuary:
693 development of targeted therapy against cell adhesion molecules in acute lymphoblastic
694 leukemia. *Seminars in oncology*. 2017;44(2):101-12.
- 695 69. Arai F, Hosokawa K, Toyama H, Matsumoto Y, Suda T. Role of N-cadherin in the
696 regulation of hematopoietic stem cells in the bone marrow niche. *Annals of the New York
697 Academy of Sciences*. 2012;1266(1):72-7.
- 698 70. Zhi L, Wang M, Rao Q, Yu F, Mi Y, Wang J. Enrichment of N-Cadherin and Tie2-
699 bearing CD34+/CD38-/CD123+ leukemic stem cells by chemotherapy-resistance. *Cancer
700 letters*. 2010;296(1):65-73.
- 701 71. Chiarini F, Paganelli F, Martelli AM, Evangelisti C. The role played by Wnt/ β -catenin
702 signaling pathway in acute lymphoblastic leukemia. *International journal of molecular
703 sciences*. 2020;21(3):1098.
- 704 72. Dandekar S, Romanos-Sirakis E, Pais F, Bhatla T, Jones C, Bourgeois W, et al. Wnt
705 inhibition leads to improved chemosensitivity in paediatric acute lymphoblastic leukaemia.
706 *British journal of haematology*. 2014;167(1):87-99.
- 707 73. Gang EJ, Hsieh YT, Pham J, Zhao Y, Nguyen C, Huantes S, et al. Small-molecule
708 inhibition of CBP/catenin interactions eliminates drug-resistant clones in acute lymphoblastic
709 leukemia. *Oncogene*. 2014;33(17):2169-78.
- 710 74. Grüninger PK, Uhl F, Herzog H, Gentile G, Andrade-Martinez M, Schmidt T, et al.
711 Functional characterization of the PI3K/AKT/MTOR signaling pathway for targeted therapy in
712 B-precursor acute lymphoblastic leukemia. *Cancer Gene Therapy*. 2022;29(11):1751-60.
- 713 75. Liang F, Ren C, Wang J, Wang S, Yang L, Han X, et al. The crosstalk between
714 STAT3 and p53/RAS signaling controls cancer cell metastasis and cisplatin resistance via
715 the Slug/MAPK/PI3K/AKT-mediated regulation of EMT and autophagy. *Oncogenesis*.
716 2019;8(10):59.

- 717 76. Ma Y, Li Y, Huang M, Meng Y. Triptolide inhibits T-cell acute lymphoblastic
718 leukaemia by affecting aberrant epigenetic events in the Wnt signalling pathway. *Journal of*
719 *Chemotherapy*. 2022;1-10.
- 720 77. Murray HC, Miller K, Brzozowski JS, Kahl RG, Smith ND, Humphrey SJ, et al.
721 Synergistic targeting of DNA-PK and KIT signaling pathways in KIT mutant acute myeloid
722 leukemia. *Molecular & Cellular Proteomics*. 2023;22(3):100503.
- 723 78. Nepstad I, Hatfield KJ, Grønningsæter IS, Aasebø E, Hernandez-Valladares M,
724 Hagen KM, et al. Effects of insulin and pathway inhibitors on the PI3K-Akt-mTOR
725 phosphorylation profile in acute myeloid leukemia cells. *Signal transduction and targeted*
726 *therapy*. 2019;4(1):20.
- 727 79. Nepstad I, Hatfield KJ, Tvedt THA, Reikvam H, Bruserud Ø. Clonal heterogeneity
728 reflected by PI3K-AKT-mTOR signaling in human acute myeloid leukemia cells and its
729 association with adverse prognosis. *Cancers*. 2018;10(9):332.
- 730 80. Perry JM, Tao F, Roy A, Lin T, He XC, Chen S, et al. Overcoming Wnt-β-catenin
731 dependent anticancer therapy resistance in leukaemia stem cells. *Nature Cell Biology*.
732 2020;22(6):689-700.
- 733 81. Pothongsrisit S, Pongrakhananon V. Targeting the PI3K/AKT/mTOR signaling
734 pathway in lung cancer: an update regarding potential drugs and natural products.
735 *Molecules*. 2021;26(13):4100.
- 736 82. Rabanal-Ruiz Y, Otten EG, Korolchuk VI. mTORC1 as the main gateway to
737 autophagy. *Essays in biochemistry*. 2017;61(6):565-84.
- 738 83. Wang Y, Krivtsov AV, Sinha AU, North TE, Goessling W, Feng Z, et al. The Wnt/β-
739 catenin pathway is required for the development of leukemia stem cells in AML. *Science*.
740 2010;327(5973):1650-3.
- 741 84. Zhang Y, Manning BD. mTORC1 signaling activates NRF1 to increase cellular
742 proteasome levels. *Cell cycle*. 2015;14(13):2011-7.
- 743 85. Zou Z, Chen J, Yang J, Bai X. Targeted inhibition of rictor/mTORC2 in cancer
744 treatment: a new era after rapamycin. *Current cancer drug targets*. 2016;16(4):288-304.
- 745 86. Gang EJ, Hsieh Y-T, Pham J, Zhao Y, Nguyen C, Huantes S, et al. Small-molecule
746 inhibition of CBP/catenin interactions eliminates drug-resistant clones in acute lymphoblastic
747 leukemia. *Oncogene*. 2014;33(17):2169-78.
- 748 87. Yarom N, Stewart D, Malik R, Wells J, Avruch L, J Jonker D. Phase I clinical trial of
749 Exherin (ADH-1) in patients with advanced solid tumors. *Current clinical pharmacology*.
750 2013;8(1):81-8.
- 751 88. Shintani Y, Fukumoto Y, Chaika N, Grandgenett PM, Hollingsworth MA, Wheelock
752 MJ, et al. ADH-1 suppresses N-cadherin-dependent pancreatic cancer progression.
753 *International journal of cancer*. 2008;122(1):71-7.
- 754 89. Li H, Price DK, Figg WD. ADH1, an N-cadherin inhibitor, evaluated in preclinical
755 models of angiogenesis and androgen-independent prostate cancer. *Anti-cancer drugs*.
756 2007;18(5):563-8.
- 757 90. Lammens T, Swerts K, Derycke L, De Craemer A, De Brouwer S, De Preter K, et al.
758 N-cadherin in neuroblastoma disease: expression and clinical significance. *PLoS One*.
759 2012;7(2):e31206.
- 760 91. Perotti A, Sessa C, Mancuso A, Noberasco C, Cresta S, Locatelli A, et al. Clinical
761 and pharmacological phase I evaluation of Exherin™(ADH-1), a selective anti-N-cadherin
762 peptide in patients with N-cadherin-expressing solid tumours. *Annals of oncology*.
763 2009;20(4):741-5.
- 764 92. Guo Z, Li W, Yuan Y, Zheng K, Tang Y, Ma K, et al. Improvement of
765 chemosensitivity and inhibition of migration via targeting tumor epithelial-to-mesenchymal
766 transition cells by ADH-1-modified liposomes. *Drug delivery*. 2018;25(1):112-21.
- 767 93. Sun Y, Jing J, Xu H, Xu L, Hu H, Tang C, et al. N-cadherin inhibitor creates a
768 microenvironment that protect TILs from immune checkpoints and Treg cells. *Journal for*
769 *immunotherapy of cancer*. 2021;9(3):e002138.

- 770 94. Mrozik KM, Cheong CM, Hewett DR, Noll JE, Opperman KS, Adwal A, et al. LCRF-
771 0006, a small molecule mimetic of the N-cadherin antagonist peptide ADH-1, synergistically
772 increases multiple myeloma response to bortezomib. *FASEB BioAdvances*. 2020;2(6):339.
773 95. GOUR BJ, BLASCHUK OW, ALI A, NI F, CHEN Z, inventors Peptidomimetic
774 modulators of cell adhesion. United States 2000.
- 775 96. Vaisburg A, Blaschuk OW, inventors Modulators of cell adhesion, methods and
776 compositions therefor. United States 2018.
- 777 97. Smits IP, Blaschuk OW, Willerth SM. Novel N-cadherin antagonist causes
778 glioblastoma cell death in a 3D bioprinted co-culture model. *Biochemical and Biophysical
779 Research Communications*. 2020;529(2):162-8.
- 780 98. Mitchell CD, Richards SM, Kinsey SE, Lillieyman J, Vora A, Eden TO, et al. Benefit of
781 dexamethasone compared with prednisolone for childhood acute lymphoblastic leukaemia:
782 results of the UK Medical Research Council ALL97 randomized trial. *British journal of
783 haematology*. 2005;129(6):734-45.
- 784 99. Halsey C, Escherich G. A "Goldilocks" approach to CNS leukemia is needed. *Blood*.
785 2021;138(4):288-9.
- 786 100. Peirs S, Matthijssens F, Goossens S, Van de Walle I, Ruggero K, De Bock CE, et al.
787 ABT-199 mediated inhibition of BCL-2 as a novel therapeutic strategy in T-cell acute
788 lymphoblastic leukemia. *Blood, The Journal of the American Society of Hematology*.
789 2014;124(25):3738-47.
- 790 101. Scherr M, Kirchhoff H, Battmer K, Wohlan K, Lee C-W, Ricke-Hoch M, et al.
791 Optimized induction of mitochondrial apoptosis for chemotherapy-free treatment of BCR-
792 ABL+ acute lymphoblastic leukemia. *Leukemia*. 2019;33(6):1313-23.
- 793 102. Ragon BK, Kantarjian H, Jabbour E, Ravandi F, Cortes J, Borthakur G, et al.
794 Buparlisib, a PI3K inhibitor, demonstrates acceptable tolerability and preliminary activity in a
795 phase I trial of patients with advanced leukemias. *American journal of hematology*.
796 2017;92(1):7-11.
- 797 103. de Gooijer MC, Zhang P, Buil LC, Çitirikkaya CH, Thota N, Beijnen JH, et al.
798 Buparlisib is a brain penetrable pan-PI3K inhibitor. *Scientific reports*. 2018;8(1):10784.
- 799 104. Brown JR, Byrd JC, Coutre SE, Benson DM, Flinn IW, Wagner-Johnston ND, et al.
800 Idelalisib, an inhibitor of phosphatidylinositol 3-kinase p110 δ , for relapsed/refractory chronic
801 lymphocytic leukemia. *Blood, The Journal of the American Society of Hematology*.
802 2014;123(22):3390-7.
- 803 105. Miller BW, Przepiorka D, de Claro RA, Lee K, Nie L, Simpson N, et al. FDA Approval:
804 Idelalisib Monotherapy for the Treatment of Patients with Follicular Lymphoma and Small
805 Lymphocytic Lymphoma FDA Approval of Idelalisib for Non-Hodgkin Lymphoma and SLL.
806 *Clinical cancer research*. 2015;21(7):1525-9.
- 807 106. Eldfors S, Kuusanmäki H, Kontro M, Majumder M, Parsons A, Edgren H, et al.
808 Idelalisib sensitivity and mechanisms of disease progression in relapsed TCF3-PBX1 acute
809 lymphoblastic leukemia. *Leukemia*. 2017;31(1):51-7.
- 810 107. Silic-Benussi M, Sharova E, Ciccarese F, Cavallari I, Raimondi V, Urso L, et al.
811 mTOR inhibition downregulates glucose-6-phosphate dehydrogenase and induces ROS-
812 dependent death in T-cell acute lymphoblastic leukemia cells. *Redox Biology*.
813 2022;51:102268.
- 814 108. Place AE, Pikman Y, Stevenson KE, Harris MH, Pauly M, Sulis ML, et al. Phase I trial
815 of the mTOR inhibitor everolimus in combination with multi-agent chemotherapy in relapsed
816 childhood acute lymphoblastic leukemia. *Pediatric Blood & Cancer*. 2018;65(7):e27062.
- 817 109. Alves R, Gonçalves AC, Jorge J, Alves J, Alves da Silva A, Freitas-Tavares P, et al.
818 Everolimus in combination with Imatinib overcomes resistance in Chronic myeloid
819 leukaemia. *Medical Oncology*. 2019;36:1-10.
- 820 110. Lang F, Wunderle L, Badura S, Schleyer E, Brüggemann M, Serve H, et al. A phase I
821 study of a dual PI3-kinase/mTOR inhibitor BEZ235 in adult patients with relapsed or
822 refractory acute leukemia. *BMC Pharmacology and Toxicology*. 2020;21(1):1-14.

- 823 111. Chen X, Wang F, Zhang Y, Wang M, Tian W, Teng W, et al. Retrospective analysis
824 of 36 fusion genes in 2479 Chinese patients of de novo acute lymphoblastic leukemia.
825 Leukemia Research. 2018;72:99-104.
- 826 112. Amado NG, Predes D, Fonseca BF, Cerqueira DM, Reis AH, Dudenhoeffer AC, et al.
827 Isoquercitrin suppresses colon cancer cell growth in vitro by targeting the Wnt/ β -catenin
828 signaling pathway. Journal of Biological Chemistry. 2014;289(51):35456-67.
- 829 113. Chen B, Dodge ME, Tang W, Lu J, Ma Z, Fan C-W, et al. Small molecule-mediated
830 disruption of Wnt-dependent signaling in tissue regeneration and cancer. Nature chemical
831 biology. 2009;5(2):100-7.
- 832 114. Liu N, Shi S, Deng M, Tang L, Zhang G, Liu N, et al. High levels of β -catenin
833 signaling reduce osteogenic differentiation of stem cells in inflammatory microenvironments
834 through inhibition of the noncanonical Wnt pathway. Journal of Bone and Mineral Research.
835 2011;26(9):2082-95.
- 836 115. Pan F, Shen F, Yang L, Zhang L, Guo W, Tian J. Inhibitory effects of XAV939 on the
837 proliferation of small-cell lung cancer H446 cells and Wnt/ β -catenin signaling pathway in
838 vitro. Oncology letters. 2018;16(2):1953-8.
- 839 116. Sohn S-H, Kim B, Sul HJ, Kim YJ, Kim HS, Kim H, et al. INC280 inhibits Wnt/ β -
840 catenin and EMT signaling pathways and its induce apoptosis in diffuse gastric cancer
841 positive for c-MET amplification. BMC research notes. 2019;12(1):1-7.
842